REFERENCES AND NOTES

- 1. I. Farnan and J. F. Stebbins, J. Am. Chem. Soc. 112, 32 (1990); J. F. Stebbins, I. Farnan, X. Xue, Chem. Geol., in press.
- 2 P. Richet, Geochim. Cosmochim. Acta 48, 471 (1984).
- 3. B. O. Mysen, J. Geophys. Res. 95, 15733 (1990); F. A. Scifert, B. O. Mysen, D. Virgo, Geochim. Cosmochim. Acta 45, 1879 (1981).
- G. A. Waychunas, G. E. Brown, Jr., C. W. Ponader, W. E. Jackson, *Nature* **332**, 251 (1988); W. E. Jackson, thesis, Stanford University (1991). 4.
- 5. M. E. Brandriss and J. F. Stebbins, Geochim. Cosmochim. Acta 52, 2659 (1988). J. F. Stebbins, Nature 351, 638 (1991).
- _____, J. B. Murdoch, A. Pines, I. S. E. Car-michael, *ibid.* **314**, 250 (1985); J. F. Stebbins, E. Schneider, J. B. Murdoch, A. Pines, I. S. E. Car-michael, *Rev. Sci. Instrum.* 57, 39 (1986); S.-B. Liu, A. Pines, M. Brandriss, J. F. Stebbins, *Phys. Chem.* Minerals 15, 155 (1987); S.-B. Liu, J. F. Stebbins, E. Schneider, A. Pines, Geochim. Cosmochim. Acta 52, 527 (1988); J. F. Stebbins, I. Farnan, P. Fiske, Eos 72, 572 (1991).
- 8. J. F. Stebbins, J. Non-Cryst. Solids 106, 359 (1988).
- S. Shimokawa et al., Chem. Lett. 1990, 617 (1990). F. Taulelle, J. P. Coutures, D. Massiot, J. P. Rifflet, Bull. Magn. Reson. 11, 318 (1989); D. Massiot, F. Taulelle, J. P. Coutures, Collog. Phys. 51, C5-425
- (1990).B. Coté, D. Massiot, F. Taulelle, J. P. Coutures, abstract, 4th Silicate Melt Workshop, Le Hohwald, France, 1991; B. T. Poe et al., Eos 72, 572 (1991).
- The nominal compositions of samples studied, be-12. fore high-T NMR runs, are listed in Table 1. Errors for major elements are about ±1% relative, for Gd₂O₃ about 20% relative. All but one contained a small amount of a paramagnetic impurity (Gd_2O_3) added to decrease ²⁹Si relaxation times. Samples were synthesized from reagent-grade alkali carbon-ates, $Al(OH)_3$, Gd_2O_3 , and SiO_2 . Si at natural isotopic abundance was used. Each sample listed in Table 1 was synthesized separately. Hyphenated sample numbers (for example, NDS-1) in text and figures indicate different portions of the same batch. Glasses both before and after high-*T* experiments were shown by ²⁹Si NMR and optical microscopy to be crystal-free. Only sample L40A5 could not readi-ly be quenched to a glass. Minor changes in compo-sition during high-T NMR experiments (primarily due to a small amount of contamination from the BN sample capsules) were noted by small differences in MAS peak positions between starting glasses and those quenched from high-*T* NMR runs (0.3 to 0.4 ppm more shielded), by semiquantitative ¹¹B MAS spectra (showing up to about 1% B_2O_3), and by small changes in high-*T* peak positions during relatively long runs (0.2 to 0.4 ppm). None of these similarity offset conclusions but for consistence. significantly affect conclusions, but for consistency some data collected during relatively slow initial heating of the liquids have been omitted from the figures. Alkali loss during NMR experiments is insignificant for the large samples (0.3 to 0.7 g) and the short run durations used here.
- 13. NMR spectra were recorded with a Varian VXR 400S spectrometer. High-T data were collected with a home-built probe [J. F. Stebbins, Chem. Rev. 91, 1353 (1991)]. The sample and radio-frequency (ff) coil were resistively heated in a small, water-cooled furnace with vertical Mo wire windings, blanketed by N_2 -3% H_2 gas. Temperatures were controlled by a thermocouple in the furnace windings that we calibrated before the NMR runs by placing a second thermocouple in a sample container in the rf coil and are accurate to about 5°C. The only possible instrumental source of variation in NMR resonant frequency with T is perturbation of the magnetic field at the sample caused by the current flow in the heater. This effect is minimized by careful, noninductive winding. We routinely mea sured and corrected residual field shifts of 1 to 3 ppm by collecting data with heater current flowing in opposite directions, and these shifts introduce uncertainties of about 0.2 ppm in measured chemi-cal shifts. Spectra were recorded at 79.4 MHz for ²⁹Si, 104.2 MHz for ²⁷Al, and 105.8 MHz for

²³Na. For ²⁹Si high-T spectra, data from 1000 to 2000 pulses, requiring about 5 min to collect, were averaged. For ²⁷Al and ²³Na, 100 to 400 pulses were used, requiring less than 1 min. Signal to noise ratios for ²³Na, ²⁷Al, and ²⁹Si were typically at least ratios for ^{-1}Na , ^{-1}Al , and ^{-1}Sl were typically at least 100:1, 50:1, and 10:1, respectively. For these nuclei, samples of 1 M NaCl, 1 M Al(NO₃)₃, and pure Si(CH₃)₄ were run at ambient T as frequency references. For ^{27}Al , 90° pulse times were measured at high T, and comparison with results for crystalline Al2O3 showed that liquid spectra collected above about 1200°C were fully averaged over all transitions. Under these conditions, peaks for ²³Na and ²⁷Al were accurately fitted by single Lorentzians, and dispersion versus absorption (DISPA) plots were circular. Both indicate that exchange dynamics were rapid enough to eliminate dynamic quadrupolar line shifts [L. G. Werbelow, J. Chem. Phys. 70, 5381 (1979); ______ and A. G. Marshall [J. Magn. Reson. 43, 443 (1981)]. MAS data were collected with a commercial high-speed probe. Spinning rates of about 6 kHz for ²⁹Si (Ål₂O₃ rotors) and up to 12 kHz for ²⁷Al (ZrO₂ rotors) were used in a probe from Doty Scientific Acquisition delays were chosen to eliminate any effects of differential relaxation rates on peak shapes. We subtracted a significant ²⁷Al background from the probe by collecting data with an empty rotor under conditions identical to those for glass samples. For 29 Si and the central (1/2 to -1/2) transition for 27 Al, MAS peak centroids were determined that included the first set of spinning side bands. For 27 Al, the centroid of the 1/2 to 3/2 transition spinning sideband manifold was estimated from side-band peak centers and used in conjunction with the main peak to estimate isotropic chemical shifts in the glasses [H. J. Jacobsen, J. Skibsted, H. Bildsoe, N. C. Nielsen, J. Magn. Reson. 85, 173 (1989); E. Lippmaa, A Samosen, M. Mägi, J. Am. Chem. Soc. 108, 1730 (1986)]. The relatively low intensity of the satellite side bands introduced an uncertainty of about 3 ppm in this estimation. We compared data for Gd-doped and undoped samples of the same lithium aluminosilicate composition (L18A5BD and L18A5BU). ²⁹Si and ²⁷Al NMR line shapes in both liquid and glass samples were the same, and peak positions differed by

- less than 0.5 ppm. For review, see G. Engelhardt and D. Michel,
- 14. High-Resolution Solid-State NMR of Silicates and

Zeolites (Wiley, New York, 1987).

- Stoichiometry requires about one nonbridging O per tetrahedral cation in most of the compositions 15. studied. This does not allow for time-averaged local symmetry to be increased to cubic (reducing the electric field gradient to 0) simply by local motion of the cations around their mean positions. Y. Bottinga and D. F. Weill, Am. J. Sci. 272, 438
- 16. (1972)
- D. B. Dingwell and S. L. Webb, *Phys. Chem. Minerals* 16, 508 (1989). 17.
- P. F. McMillan, B. T. Poe, B. Coté, D. Massiot, J. P. Coutures, *Eos* 72, 572 (1991).
 S. Hafner and N. H. Nachtrieb, *J. Chem. Phys.* 40, 2891 (1964); Y. Nakamura, Y. Kitazawa, M. Shimoji, O. M. Kitazawa, M. Shimoji, M. Kitazawa, M. Shimoji, O. M. Kitazawa 19
- S. Shimokawa, J. Phys. Chem. 87, 5117 (1983) R. M. Hazen, in Microscopic to Macroscopic, S
- ś. w. 20 Kieffer and A. Navrotsky, Eds. (Mineralogical Society of America, Washington, DC, 1985), pp. 317-346.
- This assumption might not be the case if there was extensive clustering of Al polyhedra. There is not clear evidence that this occurs in these compositions. In tectosilicate glass compositions, NMR studies have suggested that Al and Si distributions are random [see (14)].
- S. H. Risbud, R. J. Kirkpatrick, A. G. Taglialavore,
 B. Montez, J. Am. Ceram. Soc. 70, C10 (1987); R.
 K. Sato, P. F. McMillan, P. Dennison, R. Dupree, 22 Phys. Chem. Glasses 32, 149 (1991).
- R. K. Sato, P. F. McKillan, P. Dennison, R. Dupree, J. Phys. Chem. 95, 4484 (1991).
 X. Xue, J. F. Stebbins, M. Kanzaki, P. F. McMillan, B. Poe, Am. Mineral. 76, 8 (1991).
 J. B. Murdoch, J. F. Stebbins, I. S. E. Carmichael, Net Compared and Compared an 23. 24
- ibid. 70, 332 (1985).
- For a Si site with four O neighbors, the number of 26. O atoms bonded to other network-forming cations (Si^{IV}, Al^{IV}, and so forth) is indicated by n in Q^n . B. O. Myscn, *Structure and Properties of Silicate Melts*
- (Elsevier, Amsterdam, 1988)
- Supported under National Science Foundation 28. grants EAR 85-53024 and EAR 89-05188. We thank B. Coté, J. P. Coutures, D. Massiot, P. McMillan, and F. Taulelle for useful discussions; P. Fiske for help with sample preparation; and two anonymous reviewers for helpful comments.

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Experimental Phylogenetics: Generation of a Known Phylogeny

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Although methods of phylogenetic estimation are used routinely in comparative biology, direct tests of these methods are hampered by the lack of known phylogenies. Here a system based on serial propagation of bacteriophage T7 in the presence of a mutagen was used to create the first completely known phylogeny. Restriction-site maps of the terminal lineages were used to infer the evolutionary history of the experimental lines for comparison to the known history and actual ancestors. The five methods used to reconstruct branching pattern all predicted the correct topology but varied in their predictions of branch lengths; one method also predicts ancestral restriction maps and was found to be greater than 98 percent accurate.

HE DEVELOPMENT OVER THE PAST four decades of explicit methods for phylogenetic inference (1) has permitted biologists to reconstruct the broad outlines of evolutionary history and to interpret comparative biological studies within an evolutionary framework (2). However, evolutionary history usually cannot be observed directly, at least over the course of relevant magnitudes of change, so that assessment of phylogenetic methods has relied on numerical simulations. Although simulations have provided considerable insight into the effectiveness of various

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phylogenetic algorithms, they are limited by an incomplete knowledge of biology: all models incorporate untested assumptions about evolutionary processes.

Direct tests of organismal phylogenetic histories are limited to a few studies of strains of laboratory animals (3) and plant cultivars (4). Even these cases have shortcomings: the organisms underwent little genetic differentiation, phylogenies were produced over the course of decades or centuries, and the histories are in-



Fig. 1. True phylogeny for the experimental lineages of bacteriophage T7. The ancestors at each node are labeled with letters A through F and W (the latter represents wild-type T7). The numbers represent the number of restriction-site differences scored between the phages at each node of the phylogeny.

completely known. In contrast, viruses can be manipulated in the laboratory through thousands of generations per year, and mutation rates of viruses can easily be elevated through the use of mutagens, so that experimental studies of phylogenies with viruses should be feasible (5, 6). We report the creation of a known phylogeny of lineages derived from bacteriophage T7 and provide fine-scale restriction maps of the entire genomes of the experimental lineages, including the ancestors. Our purpose was to test the effectiveness of methods for inferring phylogeny and ancestral genetic character states by comparing the inferred evolutionary history against a known phylogeny and the true ancestors.

We chose to construct a symmetric phylogeny with equal distances among nodes (Fig. 1) (7). This topology (tree shape) has proven especially amenable to accurate reconstruction in theoretical studies (8-10) and may thus be regarded as a "null" model against which other topologies may be compared. We created a phylogeny of nine taxa (eight ingroup lineages and one outgroup lineage to root the tree) by serially propagating T7 phage in the presence of a mutagen and dividing the lineages at predetermined intervals (7); a clonal stock of wild-type T7 phage was the common ancestor of all lineages. There are 135,135 possible



Fig. 2. (a) DNA from each terminal lineage cleaved with the restriction enzyme Xmn I, separated by electrophoresis on 0.8% wedge-shaped agarose gels and stained with ethidium bromide. Lanes 1, 7, and 13 contain size standards (sizes shown in kilobase pairs); lane 2 contains wild-type T7 DNA; lanes 3 through 6 contain DNA from lineages R, Q, P, and O, respectively; lanes 8 through 12 contain DNA from lineages R, Q, P, and O, respectively; lanes 8 through 12 contain DNA from lineages N, M, L, K, and J, respectively. (b) Strategy for mapping restriction site variation. Viral DNA was cleaved with the restriction enzyme Stu I (which has a single recognition site in T7 that remained invariant in all lineages) and then partially digested with the restriction enzyme to be mapped. After electrophoresis on 0.8% wedge-shaped agarose gels, Southern blots were successively hybridized with four oligonucleotides that matched sequences located at each end of T7 DNA and on either side of the Stu I fragment (A through C and E). To improve resolution of the map for several restriction sites, we cleaved with the restriction enzyme Apa LI (which also has an invariant recognition site in all lineages), partially digested with the enzyme to be mapped, and hybridized the Southern blot with an oligonucleotide located at position D. (c) An example of an autoradiogram produced as described in (b), probed with oligonucleotide B. Lane 1 contains size standards (indicated in kilobase pairs); lanes 2 through 4 contain DNA from lineages O, P, and Q partially cleaved with Sau 3A1 (an isoschizomer of Mbo I.

bifurcating trees for this many taxa, so the likelihood of inferring the correct phylogeny by chance alone is minimal. We compared the actual phylogeny (Fig. 1) to estimated phylogenies from five reconstruction methods; estimates were based on restriction-site maps produced for 34 restriction endonucleases in all terminal lineages (Figs. 2 and 3). To avoid bias, the actual phylogeny was unknown to the person mapping the restriction sites. We also produced restriction maps for the ancestral phage at each of the nodes of the true phylogeny (Fig. 3). Three aspects of the inferred phylogeny were compared to the actual phylogeny: branching topology, branch lengths, and ancestral states. The five methods of phylogenetic inference evaluated were parsimony (12), the Fitch-Margoliash method (13), the Cavalli-Sforza method (14), neighbor-joining (15), and the unweighted pair-group method of arithmetic averages (UPGMA) (16).

All methods predicted the correct branching order of the known phylogeny, but no method predicted the actual branch lengths for every branch (17). To compare the five methods for their ability to predict branch lengths, the correlation between observed and predicted branch lengths was calculated for each method. These five correlations were significantly heterogeneous, with parsimony yielding the highest value and UPGMA yielding the lowest value (18). The UPGMA method is known to be sensitive to unequal rates of change (1), and the number of changes per branch was quite variable in the true phylogeny [although the number of changes per ingroup branch is not significantly heterogeneous from the expectation under a Poisson distribution; test from (19)].

The experimental system also enabled us to determine ancestral states directly. Of the methods tested, only parsimony makes predictions about ancestral character states (parsimony may be used to optimize states onto phylogenies inferred by other methods, but, for these data, all methods estimated the same branching pattern). In comparing inferred ancestral states to the actual ancestral states, three outcomes are possible: the ancestral states may be (i) correctly inferred, (ii) incorrectly inferred, or (iii) ambiguous (when more than one character optimization is possible). For 202 variable sites assayed in each of seven ancestors, parsimony correctly inferred 1369 states (97.3%), incorrectly inferred 18 states (1.3%), and was ambiguous about 20 states (1.4%). Seven states (of four sites) could not be observed in some ancestors because they fell under deletion mutations (Fig. 3). If the 91 wild-type states that were invariant in all lineages are included, the inferred restriction maps are an average of 98.6% identical to the actual maps (with either delayed or accelerated

	-	
BglII* 0.4+⊕	MboI 13.0	Scal 26.3+
SpeI 0.8++	XbaI 13.0+	MboI 26.4+
MDOI 0.8++++++	XmnI 13.2 + + + + - + + + + - + + + +	MooI 27.0++++++-+-
MOOI 1.0 +	Mini 13.3+	Hpai $27.1 + + + + + + - + - + - + - $
	HCII- 13.6	
	FIDI 13.8	MOL 21.1+
Built 1.3+++	Asso 14.2	RGRL 27/24 + + + + + + - + + + + + + + + + + + +
Del: 1.5-2.7+++	NdeT 14306 + + + + + + + + - +	Bankitt 28.2
Del: 1.7-2.6 ++++++	Bcll* 14.4++	MboI 28.2++
Del: 2.0-2.9++	ClaI 14.7+	ECONI 28749 + + + + + + + + + + + + + + + + +
BglII* 1.5 -+DD+D	BanHI* 15.3+++++-⊕-	HpaI 29.1
MboI 1.6D+-DD	MboI 15.4+	AseI 29.2 ++++++
SpeI 1.9 DDDDD++D-DDD+D+D	Scal 15515 + + + + - + + + + + + + + + + + + + +	BclI* 29.4+++
ClaI* 2.7DD-+D	χ_{mnI} 16008 + + + + - + + + - + + Θ + + + +	NsiI 29.5 - +
Mbol 3.0	Mbol 16.1 +	HpaI 29587 $+++++++-+++$
		MOOL 29.6+
Britt 3.3	Salt 16.8+++	Moot 29/17 + + - + - + - + + + - + + + + + + + +
PstT 3.7+	MooT 17.2+	For PT 29.8 - + + + - +
MooI 3.9+	Scel 17279 + + + + + + + + + + + + + + + + +	
MboI 4.0+	AseI 17.4+	PvuI* 30.0+
MboI 4.3 +	MboI 17.5 +	BstBI 30.0 +
ClaI 4.6+	KpnI 17.6+	HindIII 30.1 + + +
MboI 5.0++	BclI* 17.6+	BstBI 30.3 +++++++-+-
BamHI* 5.2++	ClaI 17.7++	HpaI 30.8+
ECONI 5.5 +++	MboI 17.8+	EcoRV 31.2+
Ase I 5.5 - + + +	EcoRI 17.9 + - +	ECORI 31.2+
		$x_{mn1} = 31202 + + + + + + + + + + + + + + + + + + $
Empty 59	Exert 18.4 $++++++++-++-++-++-+++-+++++++++++++$	MOI 31.4+++++++-+-
MooI 6.0 + + + + + + +	AseI 18.6 + + +	AseT 31.7+
BclI* 6.2 ++++++++	BstBI 19213 ++++++++++++++++++++++++++++++++++++	BallI* 32.0 +
NsiI 6.2+	XmnI 19.3 +++	ClaI* 32.1+
MooI 6.3++	SacI 19.6 +	DraI 32.2+++
BglII* 6.4+	MooI 19.7 + + - +	MooI 32.2 + + +
SacI 6.7+	MooI 19.9 - + + + +	Xm1 32.2 - + + +
MDOI 6.9 + + + + + + +	BSTELL 20065 +++++++++++++++++++++++++++++++++++	MooI 32.4 ++++++++
	Mool 20.0	SCal 32.6 ++++
		MODI 32.8+
Sept 7.2 - + + +	Salt 21.2+	Mot 33.0
SSDI 7.3 ++++++	MooI 21.6++	XmnI 33.0+
HpaI 7752 + + + + + + + + - +	MooI 21.7+	MooI 33.4
BstNI 8188 ++++++++++++++	MccI 21.9+	EcoRV 33.5+
DraI 8.6+	McoI 22.1 + + - + + ⊞-E-	MooI 33.5++++++-+-
Sall 8.8+	HindIII 22.5++++++-+-	XmnI 33.8 -+
PwiII 8.9+	NsiI 22.6+	NsiI 33.9+
	Mooi 22.7+	Scal $34099 + + + - + + + + + + + + + + + + + + +$
		CIAL* 34.9 -+
XmnT 0569 + + + + + + + + - +	MonT 22.9	YmpT 351 +
SsoI 10.2	NheI 2296 - + + + + - + - + + + + +	Mot 35.4++
SacI 10.4+	MooI 23.4++++++	MooI 35684 + + + + - + + - + + + + + + + + + + +
EcoRI 10.5+	MooI 23.6 - + +	BstEII 35.8 ++++++
McoI 11.2 ++++-+-+++++++	RpnI 23740 + + + + + - + + + + + + + + + + +	Xml 36134 +++++++++++++++++++++++++++++++++++
PviII 11481 +++++++++++++++++++++++++++++++++	HpaI 24.1 +	MboI 36.7 -+
HpaI 11804 $+++-++++++++++++$	EcoRV 24.5+	ClaI 37078 +++++++++++++++++++++++++++++++++++
ECORI 11.8 +++++++++	MDDI 24.6 +	ECORI 37.2++
SSD1 12.1+	NS11 29.8 +	Na11 37.2+
	MonT 25.2	MODI 37.6+
Front 12.4	AseT 25.3+	MonT 38.2
EcoRI 12.5 -+	MooI 25.6++	MboI 38.4+
BclI* 12.6+++	Sall 25.8+-	MboI 38.5 - + + + +
MboI 12.8++	MboI 25.8+	ClaI 38.9 +
EcoRI 12.9++++++-+-	MboI 26.1+	NsiI 39133 -+++++++++++++++
		14-T 20.3 .

Enzyma/site JKLMNOPORABCDEFW

e/site JKLMNOPOPARCDEE

e/site JKLMNOPORABCDEFW

Fig. 3. Variable restriction enzyme cleavage sites and deletions of the terminal lineages (J through R) and ancestral nodes (A through F) compared to the wild-type T7 genome (W). Wild-type sites for these enzymes that are not listed are found among all the lineages. Sites with positions shown without decimals are wild-type sites that have been lost in some of the lineages; their positions were determined from the complete sequence of T7 DNA (11). The remaining site locations were mapped as shown in Fig. 2; these positions are indicated in kilobases. Sites marked with an asterisk are also Mbo I sites; they are listed only once unless multiple changes produced differences between Mbo I and the larger recognition sequence in some of the taxa (for example, the Mbo I and Bam HI sites at position 5.2). Sites in ancestors that were inferred incorrectly by parsimony analysis of the terminal lineages are circled; those for which inference was ambiguous are surrounded by squares. A letter "D" indicates that the site occurs in a region that has been deleted in the respective lineage; these sites were coded as unknown in the analyses.

character optimization used) (1). Three of the incorrectly inferred states involved sites that were found only in the ancestral lineages, hence would not have been detected as variable characters if ancestors were unavailable (as is typically the case in phylogenetic studies).

The results of this study directly support the legitimacy of methods for phylogenetic estimation, not only with regard to reconstructing branching relationships, but also branch lengths and ancestral genotypes. Perhaps more importantly, they point the direction to a field of research in which methods of reconstruction can be tested against various known phylogenies of real organisms differing in topological and other evolutionary

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characteristics in the same fashion that tests have been conducted with simulated, theoretical phylogenies. Experimental phylogenetics is not a substitute for numerical studies, nor is it likely that laboratory phylogenies will ever display the full complexity of phylogenies produced over long-term evolution, but such studies will fill an important void in the science of phylogenetic reconstruction.

REFERENCES AND NOTES

- 1. D. L. Swofford and G. J. Olsen, in Molecular Systematics, D. M. Hillis and C. Moritz, Eds. (Sinauer, Sunderland, MA, 1990), pp. 411–501. N. Eldredge and J. Cracraft, *Phylogenetic Patterns and*
- the Evolutionary Process (Columbia Univ. Press, New York, 1980); J. Felsenstein, Am. Nat. 125, 1 (1985); D. R. Brooks and D. A. McLennan, Phylogeny, Ecology, and Behavior (Univ. of Chicago

Press, Chicago, 1991); P. H. Harvey and M. D. Pagel, The Comparative Method in Evolutionary Biol-

- ogy (Oxford Univ. Press, Oxford, 1991). 3. W. M. Fitch and W. R. Atchley, *Science* **228**, 1169 (1985); in Molecules and Morphology in Evolution: Conflict or Compromise?, C. Patterson, Ed. (Cam-bridge Univ. Press, Cambridge, 1987), pp. 203– 216; W. R. Atchley and W. M. Fitch, Science 254, 54 (1991)
- B. R. Baum, in Cladistics: Perspectives on the Reconstruction of Evolutionary History, T. Duncan and T. F. Stuessy, Eds. (Columbia Univ. Press, New York, 1984), pp. 192–220.
 F. W. Studier, in Genes, Cells, and Behavior: A View Columbia Univ. Press, New York, 1984).
- of Biology Fifty Years Later, N. H. Horowitz and E. Hutchings, Jr., Eds. (Freeman, San Francisco, 1980), pp. 72–78.
 M. E. White, J. J. Bull, I. J. Molineux, D. M. Hillis,
- in Proceedings of the Fourth International Congress of Systematic and Evolutionary Biology, E. Dudley, Ed. (Dioscorides Press, Portland, OR, 1991), pp. 935-**943**
- 7. T7 phage was grown in 1-ml cultures of Escherichia coli strain W3110 in the presence of the mutagen N-methyl-N'-nitro-N'-nitrosoguanidine (20 µg/ ml). After lysis, a new culture was infected with 10 µl of the lysate. After every five serial lysates, phage were plated on agar and a single plaque was randomly selected for further propagation. The distance between each node of the ingroup phylogeny (lineages J through P and their ancestors) was equal to 40 serial lysates; each lysate is the consequence of two to three bursts (generations) of phage. The outgroup lineage (R) was derived from wild-type T7 by a similar protocol (6). To guard against contamination among lineages, we assayed DNA from each isolated plaque with restriction enzymes whose restriction patterns evolve rapidly in this system (either Mbo I or a combination of Hae II, Hpa I, Kpn I, and Spe I) and looked for massive convergence of the restriction patterns. Two cases of contamination were detected by this procedure; after verifying the contamination with additional restriction digests, we regrew the lineages from the last contamination-free ancestor. J. Felsenstein, Syst. Zool. 27, 401 (1978). J. P. Huelsenbeck, *ibid.* 40, 257 (1991).

- J. A. Lake, Mol. Biol. Evol. 4, 167 (1987).
 J. J. Dunn and F. W. Studier, J. Mol. Biol. 166, 477 (1983).
- D. L. Swofford, Phylogenetic Analysis Using Parsimo-ny (Illinois Natural History Survey, Champaign, 1990)
- W. M. Fitch and E. Margoliash, Science 155, 279 13. (1967).
- 14 L. L. Cavalli-Sforza and A. W. F. Edwards, Am. J. Hum. Genet. 19, 233 (1967). 15. N. Saitou and M. Nei, Mol. Biol. Evol. 4, 406
- (1987). 16. R. R. Sokal and C. D. Michener, Univ. Kansas Sci.
- Bull. 38, 1409 (1958).
- For the parsimony reconstruction, bootstrap confi-dence intervals were 100% for all nodes except A 17. (92%; refer to Fig. 1 for this node), based on 1000 replications; the consistency index for informative characters was 0.750.
- Correlations between the actual and predicted 18. branch lengths were 0.91 for parsimony (average of accelerated and delayed character transformation), 0.89 for the Fitch-Margoliash method, 0.88 for the Cavalli-Sforza method, 0.88 for the neighbor-joining method, and 0.82 for the UPGMA method. As the estimates of branch lengths by these methods may violate the assumptions underlying parametric statistics, we tested the possibility of statistically significant differences among the five correlations in the following fashion. For each branch i, each of the five estimates of that branch was assigned at random and without replacement to one of the five recon-struction methods; the resulting matrix was thus a shuffling of the original matrix, with the constraint that an estimate of branch i was always assigned as an estimate of branch i, even though the method to which it was assigned varied. Correlations between actual and estimated branch lengths were computed for each of the methods in this randomized matrix, and these data were recorded; the process was repeated 10,000 times. The heterogeneity of corre-

lations from the actual estimates was found to be significantly greater than that of the randomized data (P = 0.017), with the correlation for UPGMA being significantly less than the randomized value minimum (P < 0.011). The heterogeneity of correlations among the set of four methods with UP-GMA removed is no longer significantly large, but the correlation for parsimony is larger than the maximum of randomized values at P = 0.05. By these criteria, UPGMA appears to be significantly worse than the other methods, and there is some evidence that parsimony is superior.

- G. W. Snedecor and W. G. Cochran, Statistical Methods (Iowa State Univ. Press, Ames, ed. 7, 1980).
- 20. Supported by the NSF (to D.M.H.) and the NIH (to I.J.M.). We thank F. W. Studier, I. Tessman, and T. Kunkel for advice on mutagenesis in phage.

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Chemical Signals from Host Plant and Sexual Behavior in a Moth

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In the phytophagous corn earworm, *Helicoverpa* (*Heliothis*) zea, females delay their reproductive behaviors until they find a suitable host on which to deposit their eggs. Perception of volatile chemical signals from corn silk triggers the production of sex pheromone followed by its release, which leads to mating. Several natural corn silk volatiles, including the plant hormone ethylene, induced pheromone production in *H. zea* females. Because *H. zea* larvae feed on the fruiting parts of a wide variety of hosts, ethylene, which is associated with fruit ripening, could act as a common cue.

EMALES OF MOST SPECIES OF MOTHS use sex pheromones to attract their mates. Production of the pheromone in a number of moth species is controlled by a peptide hormone, the pheromone biosynthesis-activating neuropeptide (PBAN) (1). Release of PBAN in laboratory-reared females of the corn earworm, Helicoverpa (Heliothis) zea, is regulated primarily by photoperiod (2). However, in nature, photoperiodic control appears to be superceded by signals from the host plant (2). For instance, female progeny of H. zea collected from a corn field did not produce pheromone during scotophase until they encountered corn, one of the hosts for this insect (3). The presence of silk from an ear of corn was sufficient to elicit pheromone production followed by "calling" behavior or pheromone release (3). Moreover, physical contact between the females and corn silk was not required, which indicated the involvement of a volatile factor (or factors) produced by the host plant.

Several years ago, trans-2-hexenal, a volatile component of oak leaves, was implicated in the induction of sex pheromone production in the polyphemus moth, *Antheraea polyphemus* (4, 5). However, this phenomenon could not be confirmed in a subsequent study (6). Hence the signal (or signals) and the mechanism involved in the process have remained a matter of conjecture. We show the efficacy of several chemical constituents of corn silk volatiles, including the plant hormone ethylene, in inducing pheromone production in *H. zea* females. Ethylene, widely used by plants in fruit ripening (7), may act as a common cue because *H. zea* larvae feed on the fruiting parts of a wide variety of hosts. Thus, the corn earworm female can recognize these plant chemicals and exploit this ability to coordinate its reproductive behavior with the availability of food for the offspring.

Larvae of H. zea were collected from corn fields (8) and reared through one generation on an artificial diet. The resulting females (F1) were used in all the experiments reported here. Pheromone titers [expressed as the quantity of its major component, (Z)-11hexadecenal] were determined by capillary gas chromatography (9). The wild F_1 females not exposed to corn produce only trace amounts of the pheromone (<3 ng per female). However, in the presence of the plant host (corn silk), a 20- to 30-fold increase in pheromone production occurred (Table 1). Removal of corn silk before the onset of scotophase resulted in a much lower production of pheromone, showing that the presence of plant volatiles during scotophase is required for maximal induction. Pheromone production was also induced, although to varying degrees, when moths were provided with volatiles collected from corn silk, ether-soluble and water-soluble fractions of corn silk extracts, or an intact tomato fruit (also a host for H. zea) (10). These results show that wild females stand in contrast to moths reared in the laboratory **Table 1.** Effect of host plant on sex pheromone production by wild F_1 females of *H. zea.* Individual 2-day-old virgin females were placed in cylindrical plastic containers (10 cm tall, 5 cm in diameter, and 150 ml volume) with snap-on lids. Where indicated, corn silk (1 g) was placed under a Whatman filter paper at the bottom of the container for 18 hours (that is, until pheromone extractions were carried out) in corn silk–1. In 2, corn silk was removed before the onset of scotophase or 5 hours before extraction time. All means were significantly different at $\alpha = 0.05$ (Dunn's test: distribution free multiple comparisons based on Kruskal-Wallis rank sums).

Treatment	n	Z-11-hexadecenal (ng/female ± SEM)
Control	10	2.1 ± 0.3
Corn silk–1	10	54.1 ± 6.4
Corn silk–2	5	11.1 ± 3.2

on artificial diet for many generations, which do not require host plant for the production of pheromone (2).

Because most species belonging to the Heliothis-Helicoverpa group feed on fruiting parts of various host plants and because the gaseous plant hormone ethylene is commonly produced by flowering plants, we tested the possibility that ethylene may act as a cue for pheromone production. First, using gas chromatography, we checked whether ethylene was a constituent of the volatiles produced by corn silk. On a fresh weight basis, a gram of corn silk produced 2.07 ± 0.47 (n = 6) and 2.53 \pm 0.67 (*n* = 6) nl of ethylene per hour over a 24-hour period in two separate tests. We then determined the effect of exogenous ethylene on pheromone production in H. zea females. Ethylene at various concentrations was introduced into containers holding individual H. zea females in their second photophase. Pheromone was extracted during the following scotophase



Fig. 1. Pheromone production in *H. zea* wild F_1 females as a function of ethylene concentration (mean \pm SEM, n = 7).

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