

ed by injecting a mouse with the human enzyme.

Our identification of a monoclonal antibody that discriminates between MAO-A and -B is consistent with recent evidence of structural differences in the active site subunits of the two species (8, 9, 20). Our results also agree with McCauley and Racker's (11) finding that bovine liver MAO-B has at least one antigenic site not found on bovine liver MAO-A. Conversely, Powell and Craig (12) have prepared antisera that react preferentially with human MAO-A. Our results do not support the conclusion of Dennick and Mayer (13) or Russell *et al.* (21) that human liver MAO-A and -B are immunologically indistinguishable. It should be noted, however, that these investigators raised antisera against MAO purified from liver mitochondria, which contain both MAO-A and -B. It is possible that their purified MAO contained a mixture of inactive MAO-A and active MAO-B (22), and therefore elicited antisera containing a mixture of antibodies to MAO-A and antibodies to MAO-B. Such antisera could inhibit both MAO-A and -B even if no single antibody recognized both molecules. In view of these uncertainties, additional monoclonal antibodies should be evaluated for their ability to discriminate MAO-A and -B.

Correlations between the level of MAO-B activity in human platelets and a variety of psychiatric and neurological disorders have been reported by numerous investigators (23), but there has been no method for determining whether the low MAO-B activities observed in some individuals are due to low amounts of normal enzyme or normal amounts of altered enzyme. Preliminary experiments show that crude extracts of human liver and platelets inhibit the binding of [<sup>3</sup>H]pargyline-labeled MAO-B to MAO-1C2 and suggest that this competition can form the basis for a highly sensitive quantitative assay for MAO-B protein in these extracts.

The ready availability and high specificity of antibody MAO-1C2 should prove valuable in a wide range of studies of human MAO-B, including measurement of MAO protein in platelets from patients with neurological and psychiatric disorders and preparation of human MAO-B suitable for structural studies.

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## References and Notes

- C. Schnaitman, V. G. Erwin, J. W. Greenawalt, *J. Cell Biol.* **32**, 719 (1967); J. W. Greenawalt and C. Schnaitman, *ibid.* **46**, 173 (1970).
- T. N. Tozer and N. H. Neff, *J. Pharmacol. Exp. Ther.* **153**, 177 (1966); S. Spector, R. Gordon, A. Sjoerdsma, S. Udenfriend, *Mol. Pharmacol.* **3**, 549 (1967).
- C. J. Fowler, B. A. Callingham, T. J. Mantle, K. F. Tipton, *Biochem. Pharmacol.* **27**, 97 (1978); C. H. Donnelly and D. I. Murphy, *ibid.* **26**, 853 (1977); C. J. Fowler, L. Orelund, J. Marcusson, B. Winblad, *Arch. Pharmacol.* **311**, 263 (1980); R. McCauley, *Arch. Biochem. Biophys.* **189**, 8 (1978); R. Lewinson, V. Glover, M. Sandler, *Biochem. Pharmacol.* **29**, 777 (1980).
- J. P. Johnston, *Biochem. Pharmacol.* **17**, 1285 (1968); N. H. Neff and H. Y. T. Yang, *Life Sci.* **14**, 2061 (1974).
- J. Knoll and K. Magyar, *Adv. Biochem. Psychopharmacol.* **5**, 393 (1972).
- H. L. White and A. T. Glassman, *J. Neurochem.* **29**, 987 (1977).
- L. Hellerman and V. G. Erwin, *J. Biol. Chem.* **243**, 5234 (1968); T. Singer, in *Monoamine Oxidase: Structure, Function and Altered Functions*, T. P. Singer, R. W. VonKorff, D. L. Murphy, Eds. (Academic Press, New York, 1979), pp. 7-24.
- B. A. Callingham and D. Parkinson, in *Monoamine Oxidase: Structure, Function and Altered Functions*, T. P. Singer, R. W. VonKorff, D. L. Murphy, Eds. (Academic Press, New York, 1979), pp. 81-86.
- G. K. Brown, J. F. Powell, I. W. Craig, *Biochem. Pharmacol.* **29**, 2595 (1980).
- R. M. Cawthon, J. E. Pinter, F. P. Haseltine, X. O. Breakefield, *J. Neurochem.* **37**, 363 (1981).
- R. McCauley and E. Racker, *Mol. Cell. Biochem.* **1**, 73 (1973).
- J. F. Powell and I. W. Craig, *Biochem. Soc. Trans.* **5**, 180 (1977).
- R. G. Dennick and R. J. Mayer, *Biochem. J.* **161**, 167 (1977).
- G. Köhler and C. Milstein, *Nature (London)* **256**, 495 (1975); *Eur. J. Immunol.* **6**, 511 (1976).
- Curr. Top. Microbiol. Immunol.* (1978), vol. 81; R. H. Kennett, T. J. McKearn, K. B. Bechtol, Eds., *Monoclonal Antibodies* (Plenum, New York, 1980).
- R. H. Kennett, K. A. Denis, A. S. Tung, N. R. Klinman, *Curr. Top. Microbiol. Immunol.* **81**, 77 (1978).
- C. W. Abell, R. R. Fritz, J. Landowski, J. D. Bessman, S. Castellani, J. A. Boeringa, R. M. Rose, in *Proteins of the Nervous System: Structure and Function*, B. Haber, J. R. Perez-Polo, J. D. Coulter, Eds. (Liss, New York, in press).
- In small-scale experiments, chromatofocused, [<sup>3</sup>H]pargyline-labeled human platelet MAO has been purified to homogeneity by high-performance liquid chromatography. The purified material migrates as a single band in SDS-polyacrylamide gels and comigrates with the pargyline-labeled subunit of MAO (arrow in Fig. 1A).
- R. M. Denney, N. T. Patel, R. R. Fritz, C. W. Abell, unpublished manuscript.
- R. M. Cawthon and X. O. Breakefield, *Nature (London)* **281**, 692 (1979).
- S. M. Russell, J. Davey, R. J. Mayer, *Biochem. J.* **181**, 15 (1979).
- Dennick and Mayer (13) and Russell *et al.* (21) reported that the enzyme they used for immunization lacked significant activity with serotonin and therefore contained little enzymatically active MAO-A. However, R. McCauley [*Arch. Biochem. Biophys.* **189**, 8 (1978)] noted that, although purified MAO from pig liver had little activity with serotonin, it appeared to contain inactive MAO-A, which could well be antigenic.
- M. Sandler, M. A. Reveley, V. Glover, *J. Clin. Pathol.* **34**, 292 (1981).
- R. J. Wurtman and J. Axelrod, *Biochem. Pharmacol.* **12**, 1439 (1963).
- G. Fairbanks, T. L. Steck, O. F. H. Wallach, *Biochemistry* **10**, 2606 (1971).
- C. R. Merrill, D. Goldman, S. A. Sedman, M. H. Ebert, *Science* **211**, 1437 (1981).
- M. M. Bradford, *Anal. Biochem.* **72**, 248 (1976).
- We thank J. Knoll for providing deprenyl, J. M. Smith for supplying human liver tissue, V. Kloeris, A. Waguespack, S. Stafford, W.-S. Lin, and D. Eberle for excellent technical assistance, and C. B. Denney for helpful comments on the manuscript. Supported by National Institute of Mental Health grant NIMH-34757 and by the Hogg Foundation, the Cullen Foundation, and the Multidisciplinary Research Program on Schizophrenia, University of Texas Medical Branch, Galveston. Correspondence should be addressed to R.M.D.

25 November 1981

## Brain Target Sites for 1,25-Dihydroxyvitamin D<sub>3</sub>

**Abstract.** Autoradiographic studies with <sup>3</sup>H-labeled 1,25-dihydroxyvitamin D<sub>3</sub> [1,25(OH)<sub>2</sub>D<sub>3</sub>] demonstrate, in certain neurons of rat forebrain, hindbrain, and spinal cord, a nuclear retention and concentration of radioactivity, which can be prevented by treatment with 1,25(OH)<sub>2</sub>D<sub>3</sub>, but not with 25-hydroxyvitamin D<sub>3</sub>. These results indicate the presence of brain receptors in addition to pituitary receptors for 1,25(OH)<sub>2</sub>D<sub>3</sub> and suggest a central modulation of calcium homeostasis and other central effects for this hormone. The existence of a brain-pituitary axis for certain 1,25(OH)<sub>2</sub>D<sub>3</sub>-mediated endocrine-autonomic effects is postulated.

Calcium homeostasis was generally not considered to be centrally regulated until 1979, when evidence for a direct central regulation was provided by our autoradiographic studies. After radioactively labeled 1,25-dihydroxyvitamin D<sub>3</sub> [1,25(OH)<sub>2</sub>D<sub>3</sub>] was injected in rats fed a vitamin D-deficient diet, radioactivity was found to be concentrated in the nuclei of certain cells in the pituitary pars distalis and infundibular process (1). Target cells in the anterior pituitary were identified, through combined autoradiography and immunohistochemistry, as thyrotrophs (2). Subsequently, biochemical studies demonstrated binding proteins for 1,25(OH)<sub>2</sub>D<sub>3</sub> in a pituitary tumor cell line and in homogenates of whole pituitaries, without consideration

of the specific pituitary lobes or cell types (3). The biochemical studies did not provide evidence of receptors for 1,25(OH)<sub>2</sub>D<sub>3</sub> in rat brain (3, 4); in contrast, our autoradiographic experiments revealed neuronal target sites for this steroid hormone in the brain and spinal cord (5), as well as 25 new target cell types in peripheral organs (6).

Nine 21-day-old male Holtzman Sprague-Dawley rats were fed a vitamin D-deficient diet (7) for 5 to 6 weeks and kept at a photoperiodic cycle of 12 hours of light and 12 hours of darkness. All of the animals were given intravenous injections (0.19 µg per 100 g of body weight) of 1,25(OH)<sub>2</sub>[26,27-<sup>3</sup>H]D<sub>3</sub> (specific activity 160 Ci/mmol), dissolved in 75 percent ethanol-isotonic saline. Three

of these animals were injected intravenously with 10  $\mu\text{g}$  of unlabeled  $1,25(\text{OH})_2\text{D}_3$  and one was injected with 10  $\mu\text{g}$  of 25-hydroxyvitamin  $\text{D}_3$  (25-OH- $\text{D}_3$ ) 30 minutes before injection of the

labeled compound. Two hours after injection of the labeled compound, the animals were killed. Brain, cervical spinal cord, pituitary, and intestine were dissected. Samples were mounted and

frozen onto tissue holders, sectioned in a cryostat at 4  $\mu\text{m}$ , and thaw-mounted on slides coated with photographic emulsion (Kodak NTB3) (8). The autoradiograms were exposed for 75 to 150 days, then photographically processed and stained with methyl green pyronine.

After the injection of  $^3\text{H}$ -labeled  $1,25(\text{OH})_2\text{D}_3$ , nuclear concentration and retention of radioactivity is seen in autoradiograms not only in the absorptive epithelial cells of the intestine (1) and in certain cells of the pituitary (2), but under the same conditions, in the brain and spinal cord (Fig. 1, a to f). Injection of  $1,25(\text{OH})_2\text{D}_3$ , but not of 25-OH- $\text{D}_3$ , before the injection of  $^3\text{H}$ -labeled  $1,25(\text{OH})_2\text{D}_3$  abolished or diminished nuclear labeling. In the brain, the labeled cells were probably neurons, as judged by their size and location. In the fore-brain, labeled cells were found in the nucleus centralis of the amygdala and in the rostralateral portion of the bed nucleus of the stria terminalis. Nuclear labeling also occurred in neurons of the rostral thalamus, including the nucleus periventricularis, nucleus parataenialis, and nucleus rhomboides. Further nuclear labeling was found in the medulla oblongata in cells in the area postrema and its immediate vicinity in the nucleus tractus solitarii, and in scattered cells in the caudal spinal trigeminal nucleus. The labeled cells of the latter were continuous with labeled cells in the cervical spinal cord lamina II.

Among the labeled neurons, the strongest nuclear concentration of radioactivity was observed in the bed nucleus of the stria terminalis and the nucleus centralis of the amygdala. Neurons in the dorsal thalamus are weakly labeled in comparison with the other regions in the brain and the duodenum (9). The observation that the nuclear labeling in most brain regions is equal to or stronger than that in the duodenum suggests—in conjunction with the results of the competition studies—that the nuclear radioactivity represents  $1,25(\text{OH})_2\text{D}_3$ .

The anatomical distribution of the target neurons in the brain indicates the existence of specific neuronal circuits for  $1,25(\text{OH})_2\text{D}_3$ , similar to those proposed for estradiol (10, 11). The region involved includes a portion of the stria terminalis and a sensory relay system in the spinal cord dorsal horn, the spinal trigeminal nucleus, and the area postrema–nucleus tractus solitarii. The stria terminalis is a neural pathway that interlinks brain regions related to olfaction and reproduction, including certain amygdaloid and preoptic-hypothalamic cell groups (11). Target neurons for

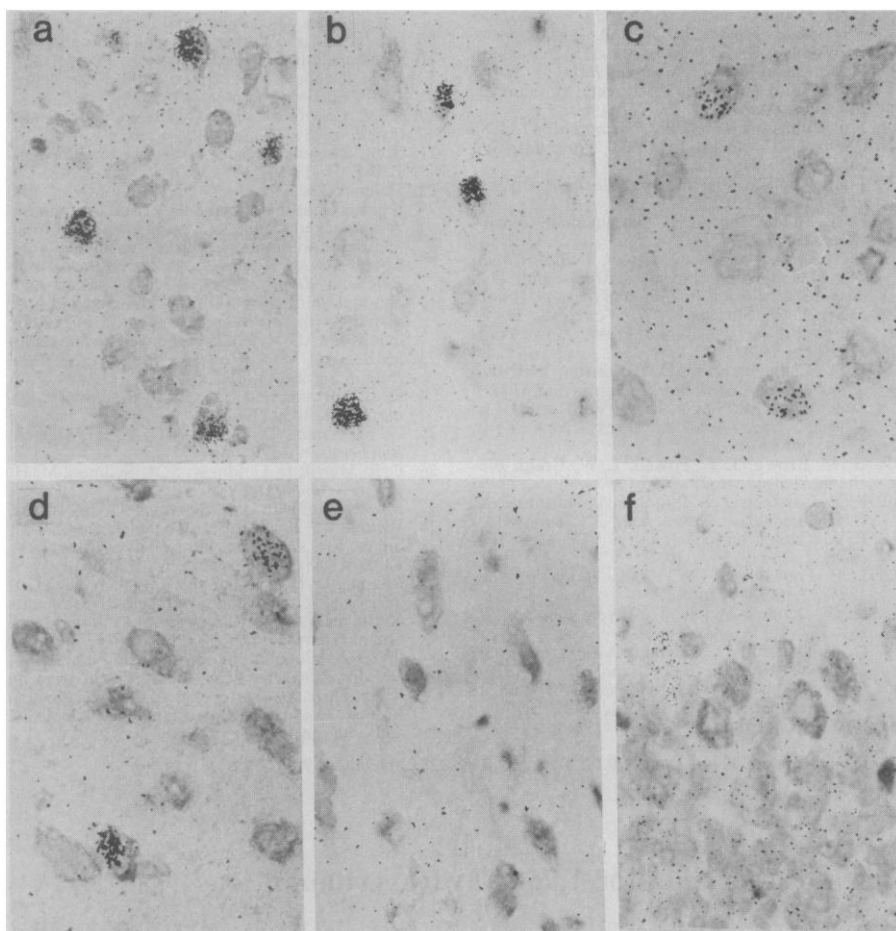
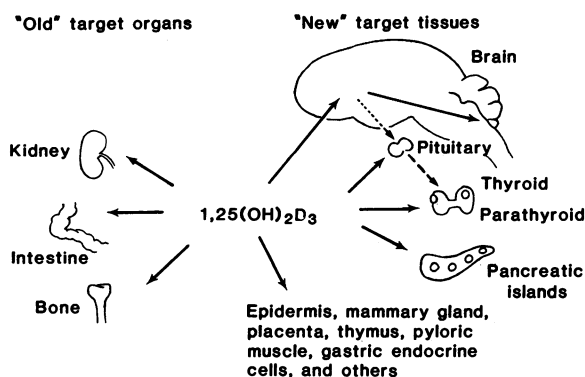


Fig. 1. Autoradiograms of rat brain showing nuclear concentration of radioactivity 2 hours after injection of  $^3\text{H}$ -labeled  $1,25(\text{OH})_2\text{D}_3$  in neurons of the (a) nucleus centralis of the amygdala, (b) nucleus interstitialis of the stria terminalis, and (c) the outer zone of the nucleus spinalis caudalis of the trigeminus. In competition studies designed to chemically characterize the nuclear radioactivity, unlabeled substances were injected before treatment with labeled  $1,25(\text{OH})_2\text{D}_3$ . (d) 25-OH- $\text{D}_3$  (nucleus centralis amygdalae) did not prevent nuclear uptake; (e)  $1,25(\text{OH})_2\text{D}_3$  (nucleus interstitialis striae terminalis) prevented nuclear uptake of radioactivity. (f) Neurons of the cerebellar cortex do not show nuclear labeling in the same animals that show labeling in (a to d). Scale bar, 4  $\mu\text{m}$ ; stain, methyl green pyronine. Magnification (a, b, e, and f),  $\times 560$ ; (c and d)  $\times 880$ . Exposure time (a to c, e, and f), 150 days; (d), 75 days.

Fig. 2. Target cells for  $1,25(\text{OH})_2\text{D}_3$  (solid lines) have been identified by autoradiography, not only in the three classical target organs (left), but in many new target tissues (6) (right). Evidence for a vitamin  $\text{D}$ -related pituitary-thyroid functional connection (dashed line) has been obtained (12), and target cells for  $1,25(\text{OH})_2\text{D}_3$  are found in both neurons and pituitary cells. Therefore, a related brain-pituitary link is postulated (dotted line), and the existence of a neuroendocrine brain-pituitary-intestinal axis, which is involved in the central regulation of calcium homeostasis, is proposed. Like other steroid hormones,  $1,25(\text{OH})_2\text{D}_3$  may act on many tissues, some of which do not seem to be directly related to calcium metabolism, but involve other functions as well.



1,25(OH)<sub>2</sub>D<sub>3</sub> in the central amygdaloid nucleus, in the bed nucleus of the stria terminalis, and in the anterior thalamus are located within projection fields of the stria terminalis, of which they seem to form a subsystem. An analogy to relationships between other steroid hormones and brain-endocrines is suggested (Fig. 2), and 1,25(OH)<sub>2</sub>D<sub>3</sub>-related peptidergic messengers and links to the portal system in the median eminence and to pituitary secretion can be expected. The finding of 1,25(OH)<sub>2</sub>D<sub>3</sub> in the brain, the previous demonstration of 1,25(OH)<sub>2</sub>D<sub>3</sub> in the pituitary (1, 2), and the effects of this hormone on thyroid-stimulating hormone secretion (12) suggest central modulation of calcium homeostasis involving a brain-pituitary-thyroid axis (Fig. 2). The possibility of a brain-intestine link in endocrine regulation must also be considered, since 1,25(OH)<sub>2</sub>D<sub>3</sub> apparently acts on structures in the amygdala, which, when lesioned or electrically stimulated, produce changes in gastric motility and secretion (13).

Vitamin D-inducible calcium-binding protein has been found not only in many peripheral target tissues for 1,25(OH)<sub>2</sub>D<sub>3</sub> but also in cell bodies, dendrites, and axons of Purkinje cells of chicken brain (14). We did not observe a concentration of radioactivity in these cells in our present studies (Fig. 1f). Although a correspondence between 1,25(OH)<sub>2</sub>D<sub>3</sub> target cells and cells containing vitamin D-dependent calcium-binding protein seems to exist—at least in some tissues—such a correspondence needs to be established through the use of our autoradiography-immunohistochemistry technique (15). This technique permits the simultaneous visualization of radioactivity and antibodies in the same preparation. Clinical reports indicate the importance of vitamin D action for normal brain function. In humans, retardation of mental growth has been observed in both vitamin D hypervitaminosis and hypovitaminosis (16). The frequency of epileptic seizures was reduced after vitamin D treatment, and this effect was unrelated to changes in serum calcium and magnesium (17). In hypoparathyroidism and pseudohypoparathyroidism, conditions in which 1,25(OH)<sub>2</sub>D<sub>3</sub> serum levels are low, seizures are a frequent symptom (18). Facilitated by the present anatomical data, the nature of the effects can now be studied and it can be clarified to which degree general or local actions of the hormone are involved.

1,25(OH)<sub>2</sub>D<sub>3</sub> may act to maintain normal brain functions by providing normal serum calcium and phosphate levels. In addition, it may selectively affect specif-

ic neuronal populations with changes in metabolism, neuritic growth, alterations of sensitivity thresholds, and changes in production and secretion of certain aminergic-peptidergic messengers, as has been demonstrated for other steroid hormones.

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#### References and Notes

1. W. E. Stumpf, M. Sar, F. A. Reid, Y. Tanaka, H. F. DeLuca, *Science* **206**, 1188 (1979).
2. M. Sar, W. E. Stumpf, H. F. DeLuca, *Cell Tissue Res.* **209**, 161 (1980).
3. J. W. Pike, L. L. Gooze, M. R. Haussler, *Life Sci.* **26**, 407 (1980); H. A. Gelbard, P. H. Stern, D. C. U'Prichard, *Science* **209**, 1247 (1980).
4. K. Colston, M. Hirst, D. Feldman, *Endocrinology* **107**, 1916 (1980).
5. W. E. Stumpf, M. Sar, S. A. Clark, E. Lieth, H. F. DeLuca, *Neuroendocrinol. Lett.* **2**, 297 (1980).
6. W. E. Stumpf, M. Sar, H. F. DeLuca, in *Hormonal Control of Calcium Metabolism*, D. V. Cohn, R. V. Talmage, J. L. Matthews, Jr., Eds. (Excerpta Medica, Amsterdam, 1981), p. 222; R. Narbaitz, W. E. Stumpf, M. Sar, *J. Histochem. Cytochem.* **29**, 91 (1981); S. A. Clark, W. E. Stumpf, M. Sar, H. F. DeLuca, Y. Tanaka, *Cell Tissue Res.* **209**, 515 (1980).
7. T. Suda, H. F. DeLuca, Y. Tanaka, *J. Nutr.* **100**, 1049 (1970).
8. W. E. Stumpf and L. J. Roth, *J. Histochem. Cytochem.* **14**, 274 (1966); W. E. Stumpf and M. Sar, *Methods Enzymol.* **36**, 135 (1975).
9. Quantitative assessment of the number of molecules in target cells was done by the method described by W. E. Stumpf, M. Sar, T. J. Zuber, E. Soini, and P. Tuohimaa [*J. Histochem. Cytochem.* **29** (1A), 201 (1981)]. Under the present experimental conditions, the numbers of molecules concentrated per nucleus in select target cells with high nuclear uptake are 15,000 to 23,000 in nucleus interstitialis striae terminalis and nucleus centralis amygdalae; 600 to 1200 in dorsal thalamus; 3600 to 8500 in area postrema and vicinity; 6800 to 7200 in spinal cord lamina II; and 2000 to 3500 in epithelial crypt cells of duodenum. These data are preliminary and need to be verified statistically with larger numbers of animals. Competition experiments with 25-OH-D<sub>3</sub> show results similar to those with <sup>3</sup>H-labeled 1,25(OH)<sub>2</sub>D<sub>3</sub> alone. In competition experiments with 1,25(OH)<sub>2</sub>D<sub>3</sub>, no nuclear concentration of radioactivity is obtained.
10. W. E. Stumpf, *Science* **162**, 1001 (1968); *Am. J. Anat.* **129**, 207 (1970); — and M. Sar, in *Anatomical Neuroendocrinology*, W. E. Stumpf and L. D. Grant, Eds. (Karger, Basel, 1975), p. 254. First evidence for brain target sites for estrogen, androgen, and progesterone was also provided by autoradiography, contrasting with negative biochemical results; reviewed by W. E. Stumpf and M. Sar, in *Steroid Hormone Regulation of the Brain*, K. Fuxe, J.-A. Gustafsson, L. Wetterberg, Eds. (Pergamon, New York, 1981), pp. 41–50.
11. L. Heimer, in *Anatomical Neuroendocrinology*, W. E. Stumpf and L. D. Grant, Eds. (Karger, Basel, 1975), p. 30; W. E. Stumpf, in *ibid.*, p. 2.
12. Treatment of vitamin D-deficient rats with 1,25(OH)<sub>2</sub>D<sub>3</sub> results in elevation of blood levels of thyroid-stimulating hormone [M. Sar, W. L. Miller, W. E. Stumpf, *Physiologist* **24**, 70 (1981)].
13. H. Kokegami, A. Kimoto, C. Kido, *Folia Psychiatr. Neurol. Jpn* **7**, 86 (1953); W. R. Hess, *Das Zwischenhirn: Syndrome, Lokalisationen, Funktionen* (Schwabe, Basel, 1954); R. N. Sen and B. K. Anand, *Indian J. Med. Res.* **45**, 515 (1957); P. G. Henke, *J. Comp. Physiol. Psychol.* **94**, 313 (1980).
14. A. N. Taylor, *J. Nutr.* **107**, 480 (1977); S. S. Jande, S. Tolani, D. E. M. Lawson, *Histochemistry* **71**, 99 (1981). The identification of "calcium-binding protein" in tissues appears complicated by possible artifacts related to technique and the existence of certain tissue-specific calcium-binding proteins, which may require the use of tissue-specific antibodies instead of antibodies against intestinal epithelium. As demonstrated earlier, for instance, with goblet cells, and cell types of kidney, pancreatic islets and pituitary, our autoradiographic techniques have proven to be sensitive and to provide reliable results as a guide for other histochemical and biochemical studies.
15. M. Sar and W. E. Stumpf, *J. Histochem. Cytochem.* **29** (1A), 161 (1981).
16. M. S. Seelig, *Ann. N.Y. Acad. Sci.* **147**, 539 (1969).
17. D. Christiansen, P. Rodbro, O. Sjö, *Br. Med. J.* **2**, 258 (1974).
18. D. H. Shelling and D. Remsen, *Bull. Johns Hopkins Hosp.* **57**, 158 (1935).
19. Supported by PHS grants NS09912 and AM14881.

23 July 1981; revised 4 January 1982

## Intraspecific Vertical Stratification as a Mate-Finding Mechanism in Tropical Cockroaches

**Abstract.** *Cockroaches in a tropical forest stratify vertically both inter- and intraspecifically along micrometeorological gradients. At night, low wind speeds and unstable atmospheric conditions result in efficient vertical mixing of the air near the ground. Convective ascent of warm air imparts directionality to the pheromone-dispersion process. The occurrence of males at greater heights than pheromone-emitting conspecific females appears to be a mate-finding strategy.*

For airborne chemicals to be effective signals in communication, their release and reception must correlate with favorable micrometeorological conditions. To maximize their efficiency in finding pheromone-emitting females, males should occupy ranges above, overlapping, or below the females' vertical ranges, depending upon the prevailing atmospheric conditions. Since meteorological patterns exhibit circadian cycles, they com-

plement the orientation behavior to specific times during the day or night. If, for example, buoyant (1) atmospheric conditions prevail at night, males should occupy perches at greater heights than females. Conversely, under stable (1) atmospheric conditions with temperature inversion, males should overlap with the vertical distribution of the females or range below them.

Dispersion models (2) describe plumes