and summer volume transports from figure 2 of

- 8. L. V. Worthington, On the North Atlantic Circulation (Johns Hopkins Press, Baltimore, 1976), p. 34.
- 9. D. H. Roemmich, "The application of inverse methods to problems in ocean circulation, thesis. Massachusetts Institute of Technology Woods Hole Oceanographic Institution Joint Program (1979).
 D. Halpern, J. R. Holbrook, R. M. Reynolds,
- D. Halpern, J. R. Holbrook, R. M. Reynolds, "A compilation of wind, current, and temper-ature measurements: near Spanish Sahara dur-ing March and April 1974" (Reference M75-103, Department of Oceanography, University of Washington, Seattle, 1975); R. D. Pillsbury, J. S. Bottero, R. E. Still, E. Mittelstaedt, "A com-pilation of observations from moored current meters: wind, currents, and temperature off northwest Africa along 21^{e40} N during JOINT-1, February-April 1974" (Reference 74-20, School of Oceanography, Oregon State University, Corvallis, 1974).
 L. Fu and C. Wunsch, POLYMODE News (Woods Hole Oceanographic Institution, unpub-lished document) 60, 1 (1979).
- 11.

- 12. L. M. Fomin, The Dynamic Method in Oceanography (Elsevier, New York, 1964). 13. W. D. Sellers, *Physical Climatology* (Univ. of
- Chicago Press, Chicago, 1965). A. F. Bunker, Mon. Weather Rev. 104, 1122 14. A.
- (1976)H. U. Sverdrup, Hand. Phys. 48, 608 (1957)
- Support for this research was provided by a summer fellowship from C. S. Payson and by grants OCE 77-19403 and OCE 77-22887 from the Office for the International Decade of Ocean Exploration of the National Science Founda tion. R. deSzoeke suggested searching for a sec tion where depth-averaged potential temper-ature has small variation. We thank K. Bryan, A. Gill, P. Rhines, and H. Stommel for stimulating discussions and R. Montgomery, P. Niller, B. Warren, and V. Worthington for helpful criti-cism. A. Bunker's help in locating values of wind stress and of energy exchange between for this study were invaluable. Woods Hole Oceanographic Institution contribution 4433 and POLYMODE contribution 124.

17 September 1979; revised 27 November 1979

Free-Radical Oxidants in Natural Waters

Abstract. Photooxidation of cumene (isopropylbenzene) and pyridine in dilute solution in natural waters gives products characteristic of reactions with alkylperoxy (RO_{2}) and hydroxyl (HO) radicals. On the basis of the rates of formation of the products, the average concentrations of RO_2 and HO are estimated to be about 10^{-9} and 10^{-17} mole per liter, respectively. The concentration of RO_2 is large enough that, for some classes of reactive chemicals, oxidation can be an important process in natural waters.

An amazing variety of synthetic organic chemicals are found in the waterways of the United States, many in significant concentrations (1). Their fates are largely unknown, although dilution, biodegradation, volatilization, hydrolysis, and photooxidation processes have all been implicated in their loss (2). Several investigators have reported evidence of free-radical oxidations of chemicals exposed to sunlight (or light of wavelength above 300 nm) in natural waters but not in pure water (2). The implication is that when the natural organics present in many water bodies undergo photolysis, they generate free radicals such as alkylperoxy (RO_2) or hydroxyl (HO) and other oxidants such as singlet oxygen $({}^{1}O_{2})$ (3) and that these oxidants contribute to the transformation of some synthetic chemicals in water. We have investigated the importance of free radicals as oxidants for organic chemicals in a variety of natural waters, using cumene (isopropylbenzene) and pyridine (C_5H_5N) to probe quantitively for RO_2 . and HO.

In our laboratory experiments only one kind of radical was generated in pure water; we used azo-bis(2-carbomethoxypropane) (MAB) as a thermal source of RO_2 · (4) and $10^{-3}M$ H₂O₂ in water, photolyzed with > 290-nm light, as a source of HO \cdot (4a). All the significant oxidation products from each kind of experiment were measured and identified.

Table 1 summarizes the experimental results and shows that cumene (CuH) at

Table 1. Distribution of oxidation products from CuH oxidized in pure water at 50°C (concentrations are in $M \times 10^{-6}$).

Radical (source)	[CuH] ₀	[Products]*			
		Side chain [†] (S)	Ring‡ (R)	S/R	
RO ₂ · (MAB)§	303	9.27 ± 0.5	< 0.02	> 500	
RO ₂ (MAB)	245	7.65 ± 0.4	< 0.01	> 800	
RO ₂ (MAB)¶	23	3.44 ± 0.2	< 0.01	> 300	
$HO (H_2O_2)#$	~ 220	2.85 ± 0.2	6.68 ± 0.4	0.43	

*Values averaged from three or more analyses; we analyzed the products by using an hplc $C_{18} \mu$ -Bondapak column (4 mm by 30 cm) and gradient elution of an acetonitrile-water mixture from 10 to 100 percent in acetonitrile in 30 minutes with 2 ml min⁻¹ throughput. An ultraviolet detector set at 254 nm was used to measure products by comparison with peak areas for known mixtures. \uparrow Sum of CuOH, CuOOH, and $C_6H_5COCH_3$. \ddagger Sum of 2- and 4-hydroxyisopropylbenzenes; identity of these products was confirmed by gas chromatography-mass spectroscopy. $\{MAB\}_0 = 1.45 \times 10^{-4}M; 24$ -hour experiment. $\|[MAB]_0 = 1.03 \times 10^{-4}M; 72$ hour experiment. $\#[H_2O_2]_0 = 1.0 \times 10^{-3}M$; experiments were carried out at 30°C with a xenon light source (6).

0036-8075/80/0222-0886\$00.50/0 Copyright © 1980 AAAS

 $10^{-4}M$ in pure water oxidizes readily in the presence of RO_2 and O_2 to give only products from side-chain oxidation: cumyl hydroperoxide (CuOOH), cumyl alcohol (CuOH), and acetophenone ($C_6H_5COCH_3$). However, HO· oxidizes CuH in water to form both side-chain and ring oxidation products, including 2and 4-isopropylphenols in a 30:70 ratio (5). The following reactions summarize the radical oxidation of CuH:

$$\begin{split} & \text{RO}_2 \cdot + \text{CuH} \rightarrow \text{RO}_2\text{H} + \text{Cu} \cdot \\ & \text{Cu} \cdot + \text{O}_2 \rightarrow \text{CuO}_2 \cdot \\ & \text{CuO}_2 \cdot + \text{CuH} \rightarrow \text{CuO}_2\text{H} + \text{Cu} \cdot \\ & \text{CuO}_2 \cdot + \text{RO}_2 \cdot \rightarrow \text{CuO} \cdot + \text{RO} \cdot + \text{O}_2 \\ & \text{CuO} \cdot \rightarrow \text{C}_6\text{H}_5\text{COCH}_3 + \text{CH}_3 \cdot \\ & \text{CH}_3 \cdot + \text{O}_2 + \text{CuO}_2 \cdot \rightarrow \text{CH}_2\text{O} + \\ & \text{CuOH} + \text{O}_2 \text{ (two steps)} \\ & \text{HO} \cdot + \text{CuH} \rightarrow \text{H}_2\text{O} + \text{Cu} \cdot \\ & \text{O} \cdot + \text{O}_2 + \text{CuH} \rightarrow \text{HOC}_6\text{H}_4\text{-}i\text{-}\text{C}_3\text{H}_7 \\ & + \text{HO}_2 \cdot \text{ (two steps)} \end{split}$$

where HOC_6H_4 -*i*- C_3H_7 is hydroxyisopropylbenzene.

H

These experiments provide the basis for interpreting results from experiments with CuH dissolved in natural waters and exposed to light. Photolyses of CuH dissolved in filter-sterilized natural waters and then exposed to either sunlight or to a xenon lamp (6) gave mixtures of the same side-chain and ring oxidation products found in radical-initiated reactions (Table 1). Table 2 summarizes these results. In all cases except for pure water, the proportions of side-chain and ring oxidation products were such that participation by both RO2 · and HO · is required to account for the results.

The ratio of products from side-chain and ring oxidation (S/R) evidently does not vary widely either in the same water source sampled at different times or in difference sources; the Aucilla River, a highly eutrophic river in northern Florida, showed somewhat greater activity than Coyote Creek or Boronda Lake and also exhibits much higher absorptivity in the solar region. The different relative proportions of oxidation products of RO_2 or HO_2 found may be due to different secondary reactions as well as different spectral, photochemical, and chemical properties of the waters (7).

We measured the products and the S/Rratio from CuH at several times in one experiment with photolyzing Aucilla River water containing $1 \times 10^{-4}M$ CuH. No significant change in S/R was found over a 144-hour period (measured at 24, 72, and 144 hours); this result indicates that phenols did not arise from the photolysis of initially formed CuOOH.

SCIENCE, VOL. 207, 22 FEBRUARY 1980

We also used C_5H_5N as a probe for natural radical oxidants when we discovered that RO₂ reacts slowly with C₅H₅N in water to give only pyridine N-oxide (C₅H₅NO), whereas HO· (from H_2O_2) gives hydroxypyridines and more polar products but no N-oxide:

 $C_5H_5N + RO_2 \cdot \rightarrow C_5H_5NO + RO \cdot$ $C_5H_5N + HO \rightarrow 2$ -, 3-, and

 $4-HOC_5H_4N$ + unidentified products

We also found that, although C_5H_5N does not photolyze in pure water at wavelengths above 300 nm, it undergoes significant loss when it photolyzes in natural waters, both in the absence and in the presence of oxygen to give very polar (hplc) unidentified products, possibly by way of the Dewar-C₅H₅N form (8). In aerated, photolyzed natural waters under conditions similar to those described in Table 2, both C₅H₅NO and hydroxypyridines were formed in small amounts together with the larger amounts of polar products.

Kinetic analysis of the experiments with CuH and C₅H₅N is straightforward. The rates of formation of oxidation products of HO· or RO_2 · with CuH or C_5H_5N follow from the assumption that each radical is in a steady-state concentration and that each radical produces a unique set of oxidation products. Thus, the CuH oxidation follows the relations

$$dS/dt + dR/dt = - (dCuH/dt) =$$

$$(k_{p}[RO_{2}\cdot] + k_{A}[HO\cdot])[CuH] \qquad (1)$$

$$k_{T} = k_{p}[RO_{2}\cdot] + k_{A}[HO\cdot] \qquad (2)$$

where $k_{\rm p}$ and $k_{\rm A}$ are the specific rate constants for H atom transfer or addition by RO_2 · and HO·, respectively (9). Integration of Eq. 1 between time limits of t = 0 and t = t gives expressions for the averaged concentrations of RO_2 and HO in natural waters exposed to sunlight or simulated sunlight (10):

$$[\text{RO}_{2} \cdot]_{\text{av}} = [S]_{t} k_{\text{T}} / [\text{CuH}]_{0} k_{\text{P}} (1 - e) (3)$$

$$[\text{HO} \cdot]_{\text{av}} = [R]_{t} k_{\text{T}} / [\text{CuH}]_{0} k_{\text{A}} (1 - e) (4)$$

A similar relation holds for the formation of C_5H_5NO in two natural waters by reaction of RO_2 with C_5H_5N . Hydroxypyridines could not be measured reliably; therefore, C₅H₅N was not used to estimate the concentration of HO·:

$$[\mathbf{RO}_2 \cdot] = [\mathbf{C}_5 \mathbf{H}_5 \mathbf{NO}]_t k^{1}_{\mathrm{T}} / [\mathbf{C}_5 \mathbf{H}_5 \mathbf{N}]_0 \times k_{\mathrm{ox}} (1 - e)$$
(5)

where, in this case, $k_{\rm T}^1$ is the rate constant for the loss of C₅H₅N by all processes known or otherwise. Table 3 summarizes the concentrations of RO_2 and HO· in natural waters as calculated from Table 2. Distribution of oxidation products from CuH photolyzed in natural waters (concentrations are in $M \times 10^{-6}$). Water samples were filter-sterilized through 0.22-µm Micropore filters and photolyzed for 5 days with a xenon source (6) (light > 290 nm).

		[Products]*		
Water source	[CuH] ₀	Side chain† (S)	Ring‡ (R)	S/R
Boronda Lake§	246	5.8	5.8	1.0
Coyote Creek	208	9.1	4.2	2.2
-	74	1.97	0.61	3.2
Aucilla River¶	274	13	6.9	1.9
Pure water#	229	0.09	0.5	0.18

*See asterisked footnote in Table 1. \dagger Sum of CuOH, CuOOH, and C₆H₃COCH₃. \ddagger Sum of 2-and 4-hydroxyisopropylbenzenes. \$Eutrophic lake, Palo Alto, California. \parallel Eutrophic rivereek, San Jose, California. \P Eutrophic river, northern Florida. #Purified with the Milli-O water puri-fication unit (Millipore Corporation).

these experiments. The fair agreement between values of $[RO_2 \cdot]$ found when CuH was used as a probe and those found when C₅H₅N was used gives some confidence that the true value is larger or smaller by less than a factor of 5 than the estimated value.

Because many natural eutrophic water bodies contain particulates which will scatter light and dissolved organics which will absorb light, the estimated $[RO_2 \cdot]_{av}$ and $[HO \cdot]_{av}$ values reported here for filtered water, photolyzed in thin layers, are upper bounds on the concentrations expected to be present in eutrophic water bodies more than 1 or 2 m in depth (10). Half-lives estimated from these concentrations will be minimum values.

From the $[RO_2 \cdot]$ measured in this way, coupled with the values of k_p for a range of organic structures (11), we can estimate minimum half-lives for these struc-

Table 3. Averaged steady-state concentrations of radicals in photolyzed natural waters. Concentrations are based on experiments with a xenon lamp adjusted to closely resemble the sunlight in brightness and spectral distribution (6); values of $[RO_2]$ and [HO] are applicable only to near-surface photolysis conditions in natural waters (see text).

$[RO_2 \cdot] [HO \times 10^9 \times 10^1$ Aucilla River CuH 2.8 1.8	Average radical concen- tration (M)*		
Aucilla River CuH 2.8 1.8]		
Boronda Lake CuH 9.5 0.15	i		
Boronda Lake C ₅ H ₅ N 0.45			
Coyote Creek CuH 9.1 1.6			
Coyote Creek C_5H_5N 5.0			

^{*}Calculated from Eqs. 3, 4, or 5; $k_{\rm T}^1 = 1.7 \times 10^{-7}$ sec⁻¹; $k_{\rm p} = 10M^{-1}$ sec⁻¹; and $k_{\rm ox} = 0.67M^{-1}$ sec⁻¹.

tures in reactions with RO_2 · in aquatic systems. In general, only the most reactive chemicals will be transformed by RO_2 , with half-lives of a few days or less; these include phenols, aromatic amines, hydroxylamines, and hydroquinones.

Despite the very high reactivity of HO, its very low concentrations found here make it unlikely that oxidation by HO· will be an important pathway in aquatic systems for most organic compounds; the half-life for oxidation by HO. of even the most reactive structure will be at least 80 days (12).

> **Theodore Mill*** DALE G. HENDRY

HAROLD RICHARDSON

Physical Organic Chemistry Department, Physical Sciences Division, SRI International, Menlo Park, California 94025

References and Notes

- 1. L. S. Shelton and R. A. Hites, *Environ. Sci. Technol.* 12, 1188 (1978) and references cited therein
- 2. R. Haque and V. Freed, Eds. Environmental Dynamics of Pesticides (Plenum, New York, 1975); J. H. Smith, W. R. Mabey, N. Bohonos, 1973; J. H. Smith, W. K. Madey, N. Bononos, B. R. Holt, S. S. Lee, T. W. Chou, D. C. Bom-berger, T. Mill, "Environmental pathways of se-lected chemicals in freshwater systems, parts I and II" (EPA Reports 600/7-77-113 and 600/ 7-78-074, Environmental Protection Agency, Washington, D.C., October 1977 and May 1970) 1978)

- 1978).
 R. G. Zepp, N. L. Wolfe, G. L. Baughman, R. C. Hollis, Nature (London) 267, 42 (1978).
 A. F. Bickel and W. A. Waters, Rec. Trav. Chim. Pays-Bas 69, 312 (1950).
 L. M. Dorfman and G. E. Adams, "Reactivity of the hydroxyl radical in aqueous solution" (Publication NSRDS-NBS 46, Government Printing Office, Washington, D.C., 1973).
 T. Mill, D. G. Hendry, H. Richardson, in Aquatic Parallel and Biological
- ic Pollutants—Transformations and Biological Effects, O. Hutzinger, L. H. Van Lelyveld, B. C. J. Zoeteman, Eds. (Pergamon, Oxford, 1978),
- 6. A 6000-W xenon lamp controlled for brightness and spectral distribution to closely resemble sunlight was used in most photolysis experiments; the unit is manufactured by the Atlas Electric Devices Company, Chicago, under the name Weather-Ometer
- Organic material in Aucilla River water scavenges RO₂ efficiently competition with added competition with added
- *p*-isopropylphenol. 8. K. E. Wilzbach and D. J. Rausch, J. Am. Chem. Soc. 92, 2178 (1970).
- Soc. 92, 21/8 (19/0).
 9. The rate constant used for H atom transfer from CuH by RO₂ is 10⁻¹ sec⁻¹, larger by about a factor of 10 than reported in (10). The larger value is based on the results of computer modeling of CuH with the CL WH is Charles. CuH oxidation (T. Mill, D. G. Hendry, H. Richardson, in preparation). The rate constant used for the addition of HO· to CuH was $3 \times 10^{9} M^{-1}$ a value close to that for toluene (4a).
- The ultraviolet spectrum of Coyote Creek water shows that it will absorb 99.9 percent of the available light at 350 nm at depths of more than
- D. G. Hendry, T. Mill, L. Piszkiewicz, J. A. Howard, H. K. Eigenmann, J. Phys. Chem. Ref. Data 3, 937 (1974).
- 12. Rate constants for the oxidation of most organic structures fall in the range of 1×10^9 to $1 \times$ $10^{10}M^{-1}$ se (4a)
- 13. We thank D. Thomas for gas chromatographicmass spectrometric identification of isopro-pylphenols and N. L. Wolfe, J. Leckie, D. G. Crosby, and J. R. Plimmer for many helpful suggestions. This study was supported by the Na-tional Science Foundation (Applied Science and Research Applications) under grant ENV 76-11153.

* Reprint requests should be sent to T.M.

26 March 1979; revised 18 June 1979

22 FEBRUARY 1980