

structures, including their stereochemistry. It is of interest that the N^6 position of adenine is in the major groove of the DNA double helix, whereas the N^2 position of guanine is in the minor groove. The significance of attack by BPDE at these two different sites, with respect to stereochemical aspects, interference with base pairing, recognition by DNA repair enzymes, and the carcinogenic process, remains to be determined.

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References and Notes

1. H. V. Gelboin and P. T'so, Eds., *Polycyclic Aromatic Hydrocarbons and Cancer* (Academic Press, New York, 1978), pp. 1-36.
2. A. M. Jeffrey, I. B. Weinstein, K. W. Jennette, K. Grzeskowiak, K. Nakanishi, R. G. Harvey, H. Autrup, C. Harris, *Nature (London)* **269**, 348 (1977).
3. K. Shinohara and P. Cerutti, *Proc. Natl. Acad. Sci. U.S.A.* **74**, 979 (1977); W. M. Baird and L. Diamond, *Biochem. Biophys. Res. Commun.* **77**, 162 (1977).
4. V. Ivanovic, N. E. Geacintov, H. Yamasaki, I. B. Weinstein, *Biochemistry* **17**, 1597 (1978).
5. M. Koreeda, P. P. Moore, P. G. Wislocki, W. Levin, A. H. Conney, H. Yagi, D. M. Jerina, *Science* **199**, 778 (1978).
6. K. Nakanishi, H. Kasai, H. Cho, R. G. Harvey, A. M. Jeffrey, K. W. Jennette, I. B. Weinstein, *J. Am. Chem. Soc.* **99**, 258 (1977).
7. P. A. Cerutti, F. Sessions, P. V. Hariham, A. Lusky, *Cancer Res.* **38**, 2118 (1978).
8. M. R. Osborne, R. G. Harvey, P. Brookes, *Chem.-Biol. Interact.* **20**, 23 (1978).
9. K. M. Straub, T. Meehan, A. L. Burlingame, M. Calvin, *Proc. Natl. Acad. Sci. U.S.A.* **74**, 5285 (1977).
10. K. W. Jennette, A. M. Jeffrey, S. H. Blobstein, F. A. Beland, R. G. Harvey, I. B. Weinstein, *Biochemistry* **16**, 932 (1977).
11. T. Kakefuda and H. Yamamoto, *Proc. Natl. Acad. Sci. U.S.A.* **75**, 415 (1978).
12. H. Kasai, K. Nakanishi, S. Traiman, *J. Chem. Soc. D* (1978), p. 798.
13. A. M. Jeffrey, K. W. Jennette, S. H. Blobstein, I. B. Weinstein, F. A. Beland, R. G. Harvey, I. Miura, K. Nakanishi, *J. Am. Chem. Soc.* **98**, 5714 (1976).
14. The high-resolution spectrum was measured on a JEOL-01SG-2 instrument at 8 kV, 4.4 A; the probe temperature was 250°C. Further losses of 43 (CH_3CO) and 60 (CH_3COOH) mass units occurred from the m/e 444 fragment to give ions at 401 ($\text{C}_{24}\text{H}_{18}\text{NO}_5$, 50 percent) and 384 ($\text{C}_{24}\text{H}_{18}\text{NO}_4$, 100 percent). Consistent with the proposed structure were the ions corresponding to losses from the molecular ion of (i) CH_3COOH to give fragment 761 ($\text{C}_{40}\text{H}_{35}\text{N}_5\text{O}_{11}$, 20 percent), followed by losses of CH_3COO to 702 (20 percent) and CH_3COOH to 642 ($\text{C}_{38}\text{H}_{28}\text{N}_5\text{O}_7$, 85 percent); (ii) CH_3COO and the benzo(a)pyrene moiety to give an ion at m/e 334 ($\text{C}_{14}\text{N}_{16}\text{N}_5\text{O}_5$, 10 percent); and (iii) the ribose moiety and CH_3COO to give an ion at m/e 503 ($\text{C}_{29}\text{H}_{21}\text{N}_5\text{O}_4$, 10 percent), followed by a loss of CH_3CO to 460 ($\text{C}_{27}\text{H}_{18}\text{N}_5\text{O}_3$, 5 percent).
15. M. Park, H. Isemura, H. Yuki, K. Takiura, *Yakugaku Zasshi* **95**, 68 (1975); J. Zemlincka and A. Holy, *Collect. Czech. Chem. Commun.* **32**, 3159 (1967).
16. T. Prusik, N. E. Geacintov, C. Tobiasz, V. Ivanovic, I. B. Weinstein, *Photochem. Photobiol.* **29**, 223 (1979).
17. P. Pulkrabek, S. Leffler, D. Grunberger, I. B. Weinstein, *Biochemistry* **18**, 5128 (1979).
18. We thank Dr. P. Pulkrabek and Dr. D. Grunberger for discussions and for samples of BDPE modified plasmid DNA, and the NMR Center at the University of Pittsburgh for the 250-MHz NMR spectra. Supported by NIH grants CA-21111, CA-13696, and CA-11572 and ACS grant BC-132.

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Generation of Unidirectionally Propagated Action Potentials in a Peripheral Nerve by Brief Stimuli

Abstract. Single, unidirectionally propagated action potentials can be elicited in peripheral nerves by electrical stimuli of short duration. Propagation in one direction is blocked anodically by means of a quasi-trapezoidal stimulus wave form and a modified tripolar electrode configuration. Propagation in the other direction proceeds unhindered. This technique may be applicable to collision blocking of motor nerves for neural prostheses.

Electrical stimulation of a peripheral nerve ordinarily elicits two action potentials, which propagate in opposite directions from the stimulus site. We report a technique in which a single unidirectionally propagated action potential is generated by brief current pulses. This mode of stimulation could be used to effect motor-nerve blockage by introducing antidromic impulses in the peripheral nerve. The block would arise from the head-on collision (and subsequent mutual annihilation) of the natural efferent impulses and the artificially generated antidromic impulses. Directionally controlled stimu-

lation is essential in producing antidromic impulses in order that the muscle receive no artificially generated neural activity.

We demonstrated the technique successfully in 15 cats by using the preparation shown in Fig. 1A. Regulated current pulses were delivered to the sciatic nerve through an asymmetrical tripolar cuff electrode (Fig. 1B). Of the current returning to the cathode, 10 to 30 percent originated from the proximal anode; the remainder flowed from the closer distal anode. Orthodromic propagation was blocked by the potential gradient arising

between the cathode and distal anode, which interfered with the flow of excitatory action currents. Spurious orthodromic excitation that occurred distal to the block as a result of stimulus current spread was suppressed by the tripolar electrode configuration, which tended to contain current flow within the insulator. The stimulus wave form consisted of a square leading edge and a 350- μsec plateau phase followed by an exponential falling phase (1). Electromyogram (EMG) tracings were obtained by recording distally (R1) from the medial gastrocnemius with intramuscular electrodes of fine stainless steel wire. Compound sciatic neurogram tracings were obtained by recording proximally (R2) with 45- μm straight wires inserted into the nerve trunk with hypodermic needles (29 gauge).

Figure 2A shows a typical sequence of responses recorded at R1 and R2 with increasing stimulus amplitude. A maximal EMG response was elicited at an amplitude of 0.5 mA; as the amplitude was increased to 6 mA, the response disappeared. Simultaneously, the neurographic response grew to a maximum as smaller fibers were recruited. This demonstrates the feasibility of preventing orthodromic propagation from the stimulus site to the muscle. The persistence of the antidromic response is implied by the maximal sciatic discharge at R2. However, an additional experiment was required to demonstrate conclusively that the particular axons innervating the medial gastrocnemius were among those discharging antidromically. This was accomplished by establishing that, after the blocking stimulus, these axons were refractory on the proximal side of the blocking cuff.

Stimulating hook electrodes (S2) were placed proximal to the cuff and their stimulus amplitude was adjusted to produce a maximal EMG response with a 100- μsec pulse. Neurogram recording sensitivity was adjusted such that the compound action potential (recorded at R2) elicited by a maximal test stimulus at S1 to the medial gastrocnemius branch was clearly visible (trace a in Fig. 2B). Stimuli were then delivered at S2 immediately after the blocking pulse. The blocking pulse itself elicited a strong neural discharge at R2 (traces b to h). Stimulation at S2 failed to elicit any further neural activity when delayed from the onset of the blocking pulse by 1.75 msec or less (traces b and c). This indicates that all axons normally excited by stimulation at S2, including those innervating the medial gastrocnemius, were refractory in this time period, hav-

ing already propagated an antidromic impulse in response to the blocking stimulus. As the delay was increased to 2.9 msec (traces c to g), the neural response gradually returned to normal and there

was a simultaneous recovery of the maximal EMG response.

This stimulation technique may be applicable to development of a clinically useful motor-nerve block for spasticity

control. For example, a short-term collision block may be very effective in controlling urinary sphincter spasticity (which interferes with efforts to urinate) after certain injuries to the spinal cord (2). Stimulus repetition rates adequate to block natural motor activity have been demonstrated, and successful biphasic coupling of the stimulus has been achieved (3). Although safe levels of electrical charge injection have not been determined for peripheral nerves, the stimulus strengths that we used appear to meet realistic electrode surface-area requirements (4).

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References and Notes

1. These parameters represent the shortest stimulus pulse adequate to sustain an orthodromic block as determined by a study (C. van den Honert and J. T. Mortimer, in preparation) of responses to single pulses. Parameters investigated include pulse width (0 to 8 msec), linear (0 to 10 msec) and exponential (0- to 10-msec time constant) falling edges, fraction of current diverted to the proximal anode (0 to 100 percent), and electrode spacing. Of the current flowing through the proximal anode, the proportion necessary to suppress spurious excitation was never greater than 30 percent, and no adjustment during the course of an experiment was required.
2. D. B. Halverstadt and W. F. Leadbetter, *Br. J. Urol.* **40**, 175 (1968).
3. According to the analysis of A. Iggo [*J. Physiol. (London)* **142**, 110 (1958)], a stimulus rate of $V/2d$ Hz is required to establish a complete block, where V represents nerve conduction velocity and d the distance between the stimulus site and the cell body. (This is subject to the constraint that the interval between efferent impulses is never less than the interstimulus interval.) In practice, a lower frequency would suffice if (i) the refractory period is included in the analysis; (ii) a partial block is adequate (some efferent impulses may escape the block); or (iii) the source of efferent impulses is inhibited by the invasion of antidromic impulses. In particular, the integration of synaptic inputs to the motor neuron is reset by antidromic impulses. A stimulation rate slightly greater than the natural frequency of discharge would thus serve to prevent that integration process from ever achieving threshold. Normal motor-nerve discharge typically has a frequency between 5 and 25 Hz, and rarely exceeds 40 Hz [R. W. Clark, E. S. Luschei, D. S. Hoffman, *Exp. Neurol.* **61**, 31 (1978); B. Derfler and L. J. Goldberg, *ibid.*, p. 592; A. W. Monster and H. Chan, *J. Neurophysiol.* **40**, 1432 (1977); M. Kato and J. Tanji, *Brain Res.* **40**, 345 (1972); J. Tanji and M. Kato, *Exp. Neurol.* **40**, 771 (1973)]. A separate study (C. van den Honert and J. T. Mortimer, in preparation) has indicated that orthodromic blocking is well sustained at stimulation frequencies up to 50 Hz with the stimulus described here and up to 30 Hz with a balanced biphasic stimulus wave form of similar charge. The study uses this technique to give a preliminary demonstration of blockage of muscular contraction. The principal complication envisioned in clinical implementation of collision blocking is excitation of afferent fibers within the nerve and consequent reflex effects. These effects will require individual assessment for each potential application. Techniques for selective excitation of small fibers [N. Accornero, B. Giorgio, G. L. Lenzi, M. J. Manfredi, *J. Physiol. (London)* **273**, 539 (1977); W. Burke and G. L. Ginsborg, *ibid.* **132**, 586 (1956)] may be adaptable to antidromic pulse generation, but the long pulse widths used in the studies cited are not amenable to high repetition rates.
4. C. van den Honert, thesis, Case Western Reserve University (1979).

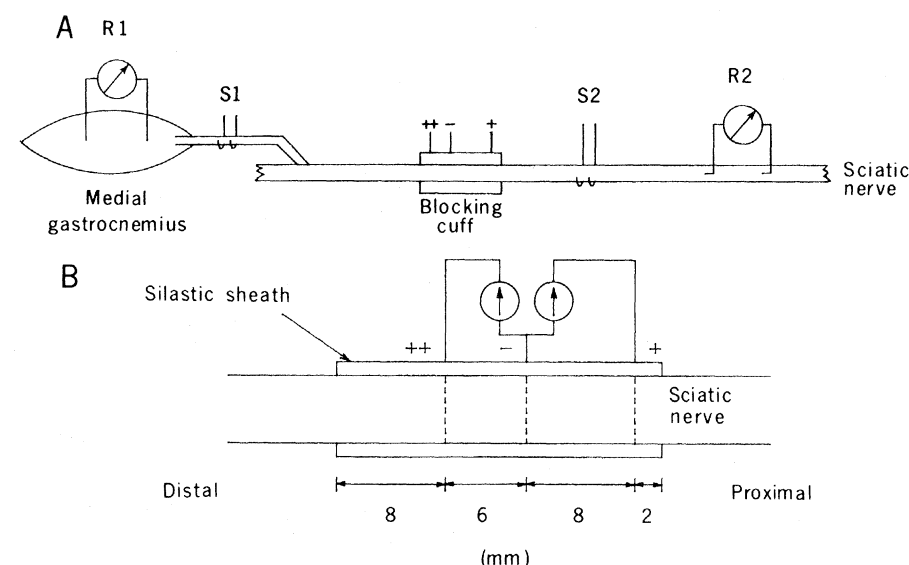


Fig. 1. (A) Experimental preparation: R1 and R2 represent EMG and neurogram recording sites, respectively; S1 and S2 represent silver hook electrodes. (B) Detail of insulated tripolar blocking electrode shown as blocking cuff in (A). Dashed lines represent rings within a silicone-rubber sheath (Silastic). Current is divided unevenly between the anodes.

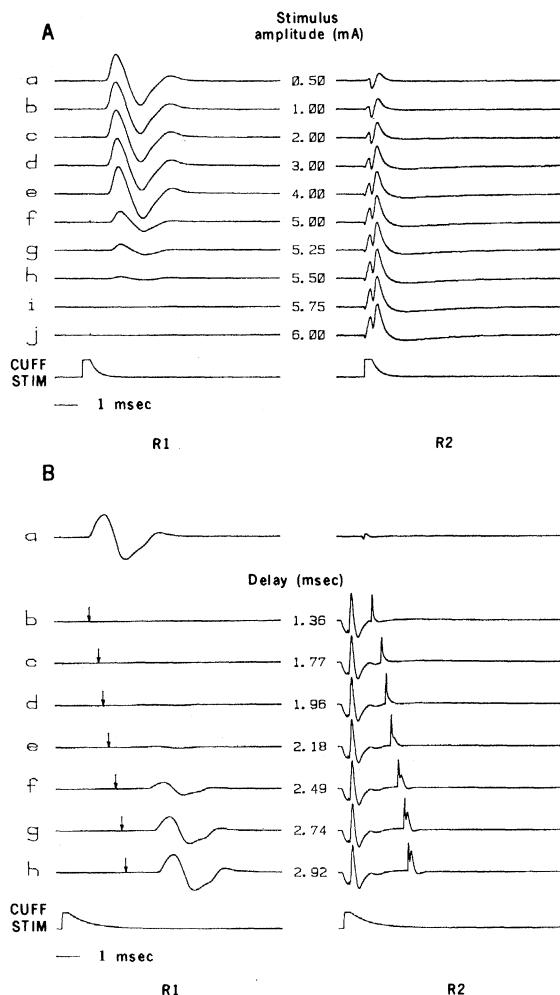


Fig. 2. (A) Electromyograms (R1) and sciatic neurograms (R2) of responses to blocking stimuli of increasing amplitude (350- μ sec plateau, 350- μ sec falling-phase time constant). (B) Confirmation of antidromically propagated excitation in response to the blocking stimulus. Trace a: test stimulus S1 elicits maximal EMG at R1 and compound action potential at R2 (axons to medial gastrocnemius only). Traces b to h: 100- μ sec stimulus pulse at S2 delayed from the leading edge of blocking stimulus (2.8 mA, 1-msec falling-phase time constant). (Arrows indicate points of S2 stimulation on EMG traces.) Blocking stimulus elicits a maximal neural discharge at R2 (early deflection) but no EMG response. With a short delay (traces b and c), stimulus at S2 produces only a stimulus artifact at R2 (delayed deflection). With increased delay (traces d to h), neural response to stimulation at S2 develops as the delayed stimulus occurs outside the refractory period, and maximal EMG response develops.

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