Cyclolobus is confined to the upper Chhidru. All the Salt Range representatives of Cyclolobus for which accurate occurrence data are available came from 10 to 23 m below the top of the Chhidru (5, p. 73). There is no conclusive ammonoid evidence for the precise age of the Kalabagh.

Four additional ammonoids have been reported from the Zewan Formation in the vicinity of Srinagar. "Xenaspis cf. carbonaria Waagen" was recorded by Diener (17) as being represented by three specimens (India Geological Survey, Nos. 11117 to 11119) from the "Spur 2 miles N. of Barus," a well-known locality (12) approximately 11 km south-southeast of Guryul Ravine. The parent horizon is "Unit 3¹/₂" of Middlemiss and probably corresponds to the basal few meters of the Triassic black shales at Guryul Ravine rather than the underlying Zewan Formation. Our recent study of these three specimens reveals that they are at least specifically distinct from the widespread Late Permian Xenodiscus carbonarius, and are best regarded as indeterminate Triassic ophiceratins, possibly Glyptophiceras. An associated bivalve [plate 6, figure 1, in (17)] is Claraia stachei (Bittner). The fourth ammonoid reported from the Zewan Formation (India Geological Survey, No. 11120) was collected near Pahlgam (Pailgam) approximately 95 km east of Srinagar. Stratigraphic occurrence of the specimen is uncertain (12) and Diener's assignment (17) to the Permian genus Popanoceras is certainly incorrect. Our examination leads us to regard the specimen as generically indeterminate, but Triassic in age. Unquestionably Permian ammonoids, including Xenodiscus and Stacheoceras, have been collected recently from Pahlgam and adjacent areas, but the details of occurrence are not yet available.

The discovery of Cyclolobus walkeri near the top of the Zewan Formation of Kashmir strengthens the correlation of the parent beds with the upper Kuling Shale of the central Himalayas, the upper Chhidru Formation of the Salt Range, and the Ankitohazo beds of Madagascar. All occurrences are interpreted as representing the Chhidruan Stage of the middle Dzhulfian Series (18). It follows that if no major hiatus occurs in the upper Zewan, the latest Dzhulfian Changhsingian Stage is represented in the top few meters of the Zewan Formation at Guryul Ravine. Detailed collecting and comparison of the fauna from this interval with that from the type Changhsingian of South China and the correlative Paratirolites beds of Armenia and possibly Madagascar are proceeding (19).

W. M. FURNISH

BRIAN F. GLENISTER

Department of Geology, University of Iowa, Iowa City 52242

KEIJI NAKAZAWA

Department of Geology and

Mineralogy, Kyoto University, Sakyo Ward, Kyoto, Japan

HARI MOHAN KAPOOR Geological Survey of India,

3, Gokhale Marg, Lucknow, India

References and Notes

- 1. J. Phillips, in The Penny Cyclopaedia of the Society for the Diffusion of Useful Knowledge. G. Long, Ed. (Charles Knight, London, 1840), vol. 17, pp. 153–154.
- N. D. Newell, Geol. Soc. Amer. Spec. Pap 89 (1967), p. 63.
- For abstracts of review papers presented at the International Permian-Triassic Conference, the International Permian-Triassic Conference, 23-26 August 1971, Calgary, Alberta, Canada, see: Bull. Can. Petrol. Geol. 19, 313 (1971).
 V. E. Ruzhencev and T. G. Sarycheva, Eds., Akad. Nauk SSSR Trudy Paleontol. Inst.

(1965), vol. 108; D. L. Stepanov et al., Geol. Surv. Iran Rep. 12 (1969); B Kummel and C. Teichert, Bull. Can. Petrol. Geol. 19, 336 (1971); H. Taraz, Amer. Ass. Petrol. Geol.

- (1971); H. Taraz, Amer. Ass. Petrol. Geol. Bull. 55, 1280 (1971).
 5. B. Kummel and C. Teichert, Eds., Univ. Kans. Dep. Geol. Spec. Publ. 4 (1970).
 6. K. Nakazawa et al., Mem. Fac. Sci. Kyoto Univ. Ser. Geol. Mineral. 37, 163 (1970).
 7. W. C. Sweet, Paleontol. Contrib. Univ. Kans. No. 49 (1970).
 8. C. Teichert, B. Kummel, H. M. Kapoor, Science 167, 174 (1970).
 9. K.K. Chao Scientia Sin 14, 1813 (1965).

- 9. K.-K. Chao, Scientia Sin. 14, 1813 (1965). 10. C. Teichert and B. Kummel, Bull. Can. Petrol.
- Geol., in press. 11. H. H. Hayden, Rec. Geol. Surv. India 36, 23
- H. H. Hayden, *Rec. Geol. Surv. India* 36, 23 (1907).
 C. S. Middlemiss, *ibid.* 37, 286 (1909).
 , *ibid.* 40, 206 (1910).
 W. M. Furnish and B. F. Glenister, in Univ. Kans. Dep. Geol. Spec. Publ. 4 (1970), p. 153.
 J. B. Waterhouse, Lethata 5, 251 (1972).
 R. E. Grant, J. Paleontol. 42, 1 (1968).
 G. R. E. Grant, G. Corr, Lethata Belastrat.

- K. E. Orani, J. Falconiol. 42, 1 (1966).
 C. Diener, Mem. Geol. Surv. India Palaeontol. Indica (1915), vol. 5, No. 2.
- 18. W. M. Furnish, Bull. Can. Petrol. Geol., in press
- press. 19. K. Nakazawa et al., in preparation. 20. Financial support for W.M.F. and B.F.G. was provided by NSF grant GB-5530 and the Graduate College, University of Iowa. Field studies by K.N. and H.M.K. were sponsored by the Japanese Ministry of Education and the Geological Survey of India. M. V. A. Sector Collegical Survey of India. M. V. A. Sastry, Geological Survey of India. M. V. A. Sastry, Geological Survey of India (Calcutta), provided access to comparative collections. This report is published with the permission of the director general, Geological Survey of India.
- 16 January 1973

1*α*-Hydroxy Derivative of Vitamin D₃: A Highly Potent Analog of 1α ,25-Dihydroxyvitamin D₃

Abstract. The 1α -hydroxy derivative of vitamin D_3 has been chemically synthesized and tested for its biological activity. This analog has comparable biological activity on a weight basis to 1,25-dihydroxyvitamin D_3 in the stimulation of intestinal calcium transport and bone calcium mobilization in normal and anephric rats. Because the 1α -hydroxy derivative is synthesized from cholesterol, it is easier and less expensive to prepare than 1α ,25-dihydroxy derivative, making it attractive as a drug in the treatment of renal osteodystrophy and hypoparathvroidism.

For vitamin D to carry out its biological functions in stimulating intestinal calcium transport and mobilizing bone calcium, it must be hydroxylated on C-25 in the liver (1) and subsequently on C-1 by the kidney (2). The resulting metabolite, 1α ,25-dihydroxyvitamin D_3 $[1\alpha, 25-(OH)_2D_3]$ (Fig. 1) was isolated in pure form, identified (3), and recently synthesized (4). This metabolite is the most potent form of vitamin D known (5, 6), and it is the only known

Table 1. Intestinal calcium transport and bone calcium mobilization response of rats to 1_{α} -OH-D₃. Results are means of six rats; S.E., standard error.

1_{α} -OH-D ₃ (pmole)	⁴⁵ Ca serosal/ ⁴⁵ Ca mucosal (mean ± S.E.)	Serum Ca (mg/ml) (mean ± S.E.)
	Normal	
None*	1.5 ± 0.2	4.3 ± 0.1
6.2	1.8 ± 0.2	4.8 ± 0.1
62.5	2.8 ± 0.3	5.8 ± 0.1
625	2.9 ± 0.2	6.7 ± 0.1
1250	2.9 ± 0.2	6.9 ± 0.1
	Anephric	
None*	1.5 ± 0.2	4.1 ± 0.1
312	2.3 ± 0.2	5.2 ± 0.1

* The control dose consisted of 50 μ l of 95 percent ethanol alone.

vitamin D metabolite active in anephric rats (7).

During the course of the $1\alpha, 25$ - $(OH)_2D_3$ synthesis we explored the various reactions utilizing a less expensive starting material, namely cholesterol instead of homocholenic acid. As a result, 1α -hydroxyvitamin D₃ (1α -OH-D₃) (Fig. 1) was prepared.

The possibility that such an analog of 1α , 25-(OH)₂D₃ might be biologically active was of obvious interest and consequently was examined. The analog proved to be approximately equal to 1α ,25-(OH)₂D₃ in inducing intestinal calcium transport and bone calcium mobilization in normal and anephric rats.

The synthesis of 1α -OH-D₃ was accomplished by the methods described for 1α ,25-(OH)₂D₃, except that cholesterol was used as starting material (4). The resulting product showed the expected ultraviolet absorption spectrum (λ_{max} 265 nm, λ_{min} 228 nm) for the 5,6-cis triene system and mass spectrum $[m/e 400 (M^+), 287, 152, and 134].$

Furthermore, the compound chromatographed as one peak on gas-liquid chromatography, demonstrating its purity (it is interesting that, although this analog has a 5,6-cis triene system, it does not cyclize in the gas chromatograph to the pyro and isopyro forms as do other vitamin D metabolites, probably because of the close proximity of 1α -OH to the triene system).

Weanling male rats (Holtzman) were fed for 2 weeks a vitamin D-deficient diet containing adequate calcium and phosphorus (8) and then fed a low calcium (0.02 percent), vitamin Ddeficient diet for another week (9). Groups of rats, either bilaterally nephrectomized or normal, received the appropriate dose via the jugular vein in 0.05 ml of 95 percent ethanol (Fig. 2 and Table 1). Intestinal calcium transport and bone mobilization assays were performed as described (9).

The results shown in Fig. 2 demonstrate that 1α -OH-D₃ is capable of stimulating intestinal calcium transport and bone calcium mobilization. This analog elicits an intestinal calcium transport response as early as 3 hours after its administration and shows a maximum response at 6 hours. 1,25-(OH)₂D₃ also gives a maximum response 6 hours after intravenous administration (6). Although 1α -OH-D₃ does not show as large a transport response as 1,25-(OH), D₃ at 6 hours, it sustains the response for at least 24 13 APRIL 1973



Fig. 1. Structures of 1α , 25-(OH)₂D₃ (1), and 1α -OH-D₃ (2).

hours, while the response to 1,25- $(OH)_2D_3$ decreases after 6 hours (6). The 1α -OH-D₃ is capable of mobilizing calcium from bone as early as 3 hours after its administration and shows a maximum response at 14 hours.

Table 1 shows that as little as 62.5 pmole of 1α -OH-D₃ induces intestinal calcium transport and bone mineral mobilization at 14 hours after administration. A similar response to 62.5 pmole of 1,25-(OH)₂D₃ has been demonstrated (10). Of great importance is the fact that 1α -OH-D₃ stimulates both intestinal calcium transport and bone mineral mobilization in anephric rats (Table 1). However, as the dose of 1α -OH-D₃ is increased from 625 pmole to 1250 pmole, there is no significant increase in the intestinal calcium transport response.

Because the 1α -OH-D₃ is less expensive and less difficult to prepare chemically as compared to $1\alpha 25$ -(OH)₂D₃, but has similar activity in both anephric and intact vitamin Ddeficient rats, it is potentially attractive for the treatment of renal osteodystrophy and hypoparathyroidism. It has been suggested that the 5,6-trans form of



Fig. 2. Intestinal calcium transport) and bone calcium mobilization (----) response of vitamin D-deficient rats on a low calcium diet to 625 pmole of 1a-OH-D₃. The vertical bars represent the standard error of the means; six animals were used for each measurement.

vitamin D_3 may be a suitable substitute for $1,25-(OH)_2D_3$ because it can induce both intestinal calcium transport and bone calcium mobilization in anephric rats (9). Although this analog is easy and inexpensive to make it is about 50 to 100 times less active than either the $1,25-(OH)_2D_3$ or 1α -OH-D₃.

It is not possible to predict at this time whether 1α -OH-D₃ is hydroxylated on C-25 in the liver before it is active. It is interesting that the time it takes to produce a maximum response in the intestine parallels that for $1\alpha, 25$ - $(OH)_2D_3$, which would argue against further hydroxylation. On the other hand the 1α -OH-D₃ does not attain the high intestinal calcium transport response seen for 1α ,25-(OH)₂D₃ but can sustain its response for longer periods, which could be consistent with the idea that the 1α -OH-D₃ is further hydroxylated presumably on C-25. Additional investigation will be necessary to answer these questions, but the possible utility of this compound in medicine seems clear.

M. F. HOLICK, E. J. SEMMLER H. K. Schnoes, H. F. DELUCA Department of Biochemistry, College of Agricultural and Life Sciences, University of Wisconsin, Madison 53706

References and Notes

- 1. J. W. Blunt, H. F. DeLuca, H. K. Schnoes, Biochemistry 7, 3317 (1968); M. Horsting and Biochemistry 1, 3317 (1968); M. Horsting and
 H. F. DeLuca, Biochem. Biophys. Res. Commun. 36, 251 (1969); G. Ponchon, A. L.
 Kennan, H. F. DeLuca, J. Clin. Invest. 48, 2032 (1969); J. W. Blunt, Y. Tanaka, H. F. DeLuca, Pro 1503 (1968). Proc. Nat. Acad. Sci. U.S.A. 61,
- D. R. Fraser and E. Kodicek, *Nature* 228, 764 (1970); R. Gray, I. Boyle, H. F. DeLuca, *Science* 172, 1232 (1971); A. W. Norman, R. J. Midgett, J. F. Myrtle, H. G. Nowicki, A. M. Status, Nature 1998 (1998). Biochem. Biophys. Res. Commun. 42, 1082
- 3. M. F. Holick, H. K. Schnoes, H. F. DeLuca M. F. Holick, H. K. Schnoes, H. F. DeLuca, T. Suda, R. J. Cousins, *Biochemistry* 10, 2799 (1971); M. F. Holick, H. K. Schnoes, H. F. DeLuca, *Proc. Nat. Acad. Sci. U.S.A.* 68, 803 (1971); D. E. M. Lawson, D. R. Fraser, E. Kodicek, H. R. Morris, D. H. Williams, *Nature* 230, 228 (1971).
 E. J. Semmler, M. F. Holick, H. K. Schnoes, H. F. DeLuca, *Tetrahedron Lett.* 40, 4147 (1972)
- 4. (1972)
- 5. J. Omdahl, M. Holick, T. Suda, Y. Tanaka, H. F. DeLuca, *Biochemistry* 10, 2935 (1971); M. R. Haussler, D. W. Boyce, E. T. Little-H. F. Deluca, Dichemistry 10, 2935 (1911);
 M. R. Haussler, D. W. Boyce, E. T. Little-dike, H. Rasmussen, Proc. Nat. Acad. Sci. U.S.A. 68, 177 (1971); Y. Tanaka, H. Frank, H. F. Deluca, J. Nutr., in press; J. F. Myrtle and A. W. Norman, Science 171, 79 (1971).
 Y. Tanaka and H. F. Deluca, Arch. Biochem. Biophys. 146, 574 (1971).
 I. T. Boyle, L. Miravet, R. W. Gray, M. F. Holick, H. F. Deluca, Endocrinology 90, 605 (1972); M. F. Holick, M. Garabedian, H. F. Deluca, Science 176, 1146 (1972).
 G. Guroff, H. F. Deluca, H. Steenbock, Amer. J. Physiol. 204, 833 (1963).
 M. F. Holick, M. Garabedian, H. F. Deluca, Biochemistry 11, 2715 (1972).
 C. A. Frolik and H. F. Deluca, J. Clin. Invest. 51, 2900 (1972).
 Supported by PHS grants AM-15512, AM-14881, and GM00236 BCH.
 November 1972: revised 3 January 1973

20 November 1972; revised 3 January 1973