

Fig. 4. Giant sinkhole that developed catastrophically early in February 1966. It is approximately 125 m in diameter and 50 m in depth and is located within 0.8 km of a residential section of Carletonville.

ary 1966, the largest of all the catastrophic sinkholes formed near the edge of Carletonville a few days after about 15 cm of torrential rainfall. This sinkhole, measuring nearly 125 m in diameter and about 50 m in depth, occurred "without warning" at the lower (north) end of a linear subsidence area which had developed a year earlier (Fig. 4).

The unusual size of the large sinkholes that have formed makes it difficult to visualize an opening within unconsolidated debris of adequate size to accept all the material at the time of collapse. Experimental studies (9) also suggest that a cavity within the debris with a diameter equal to that of the sinkholes that have formed could not possibly support the overlying lithostatic load. I suggest that large sinkholes—and possibly many smaller ones—involve the development of multiple arches between rock pinnacles, close enough to one another so that, as they grow larger, they may suddenly coalesce, thereby increasing the span of the arch beyond its ability to support the load. Experimental studies have shown this not only to be feasible but also to provide the most likely explanation for giant sinkholes (10). This would help to explain both the large amount of existing void into which debris could collapse and the rapidity with which the debris moves downward through multiple openings (Fig. 3). After collapse of several of the large sinkholes, continued downward movement of the debris near the base of the sinkhole has been observed.

Withdrawal of fluids from pore space in unconsolidated debris may cause surface subsidence, no matter what the composition of the underlying bedrock. However, catastrophic sinkhole development occurs only if the under-

lying rocks are carbonates. In addition, bedrock configuration must be highly irregular, typified by pinnacle weathering. If the unconsolidated debris is thick and if there is great lowering of the groundwater surface, then one may expect the development of multiple openings near the base of the mass of desiccated debris. Gradual enlargement of these openings by roof spalling may finally result in coalescence or, at least, a cavity of such size that rapid upward propagation of roof spalls will result in catastrophic collapse of the surface.

Within the Oberholzer compartment on the Far West Rand, South Africa, extensive observations of sinkhole development support the suggestion that sinkholes begin to form when the groundwater surface has been lowered 30 to 60 m. The eight giant sinkholes formed after the groundwater surface had been lowered approximately 160 m. As the area of the cone of depression of the lowering groundwater surface has expanded, the occurrence of sinkholes has increased and has moved outward from the center of the cone.

RICHARD M. FOOSE

Department of Geology, Amherst College, Amherst, Massachusetts 01002

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8. J. Irving (Consulting Engineer, Blyvooruitzicht Mining Co., Johannesburg, South Africa), personal communication.
9. Personal discussion with K. Knight (Department of Civil Engineering, University of Witwatersrand, Johannesburg) and observation of his model experiments which used wet and dry sand.
10. At my suggestion, Dr. Knight prepared model experiments that showed the development of multiple arches with consequent giant sinkholes.
11. I thank the staffs of the Orange Free State and Transvaal Chamber of Mines, Goldfields of South Africa, Ltd., Anglo-American Corp., and Rand Mines, Ltd. Thanks are also due to J. Irving and J. L. Prince for their help in the field investigations.

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Echinoderm Calcite: Single Crystal or Polycrystalline Aggregate

Abstract. *Electron microscopy of natural and broken surfaces of echinoid skeletal plates reveals that the interior portions have the morphology of a single crystal, whereas the exterior is a polycrystalline aggregate with preferred orientation. These data help to resolve earlier contradictory x-ray and optical evidence.*

The skeletal elements of most echinoderms consist of a fenestrate lattice-work of calcite, and in the living animal this stereom is interpenetrated with mesodermal tissue. The calcite frequently contains significant $MgCO_3$ in solid solution. Each skeletal element behaves as a single crystal in polarized light. An important exception to this are the spine-bearing tubercles in some species of echinoids. Raup (1) has summarized much of this information concerning the nature of the echinoderm endoskeleton; and, on the basis of his own optical studies, he has demonstrated that the *c*-axes show a preferred orientation in the individual skeletal elements that is related to the morphology of the echinoid. The optical homogeneity combined with their porous and irregular geometry has often raised the question of whether the skeletal elements are single crystals or highly oriented polycrystalline aggregates. The evidence is contradictory, although the majority of it points to the single-crystal structure. All optical work in polarized light suggests a single crystal, whereas the x-ray data are in disagreement. West (2) using an echinoid spine concluded from Laue patterns that each spine was a single crystal. Donnay (3) reached a similar conclusion. However, Garrido and Blanco (4) and Nissen (5) interpreted their data to mean that each skeletal element is constructed of tiny crystallites in almost-perfect parallel orientation. Currey and Nichols (6), working with scanning electron microscopy, appear to have confirmed the data of those favoring a single crystal. From conventional electron microscopy, I now offer evidence that helps to resolve this conflict.

Adult specimens of the regular echinoid *Strongylocentrotus droebachiensis* (Müller) from Eastport, Maine, were provided for study by P. M. Kier and T. F. Phelan, of the U.S. National Museum. The dried skeletons were placed in Clorox and vacuum-soaked

overnight to insure removal of the organic material from within the fenestrated stereom. Except for the pale green pigment trapped within the calcite, most of the organic material is removed by this treatment. Single-stage platinum-carbon replicas were prepared from both the natural and fractured surfaces of plates, tubercles, and spines, and then examined in the electron microscope.

The surface of the calcite within the interior of the stereom is shown in Fig. 1. Also shown are a fractured column and adjacent latticework opening. The natural surface is rather smoothly finished except for small scattered particles of unknown origin. The fractured column and a larger fractured area in Fig. 2 reveal a conchoidal appearance. This structure is similar to some areas of freshly broken optical-grade calcite and is the same as that shown by Currey and Nichols (6).

The outer-plate surface (surface with spines and tubercles) has an entirely different appearance. There is evidence of a polycrystalline structure (Figs. 3 and 4). Needles or laths of calcite approximately 0.2μ wide occur in preferred orientation. The ends of these features, occurring at the same height, appear as a fine, bumpy surface (Fig. 3). The edges of the laths merge together, forming a smooth surface with rows of parallel elongated pits or microstriae (Fig. 4). This striated structure is observed from place to place on all outer and lateral surfaces of the plates, and it also appears on the spines where the microstriae run parallel to the long axes (Fig. 5).

The notable exceptions to the optical uniformity of the echinoid plate are the tubercles on which the spines are attached. In thin-section and in crossed-polarized light their polycrystalline character is obvious (Fig. 6). Only the mamelon and part of the boss differ in crystallographic orientation. The bulk of the tubercle is crystallographically continuous with the plate itself (not shown). The structure of the surface of the tubercle (Fig. 7) is the same as that described for the exterior of the plate, differing only in that groups of laths and microstriae are arranged in mosaic fashion. The boundaries between grains are characteristic of the tubercle mamelon surface.

Two distinct morphologies occur in a single echinoid plate: (i) the conchoidal fracture and smooth surface as in a single crystal and (ii) the highly

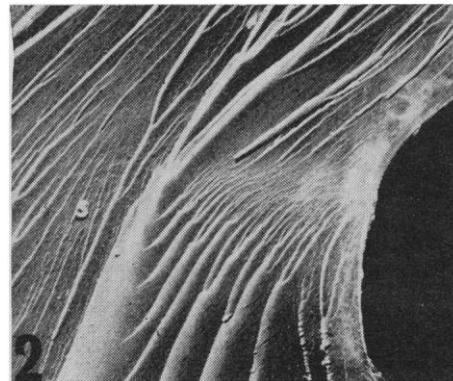
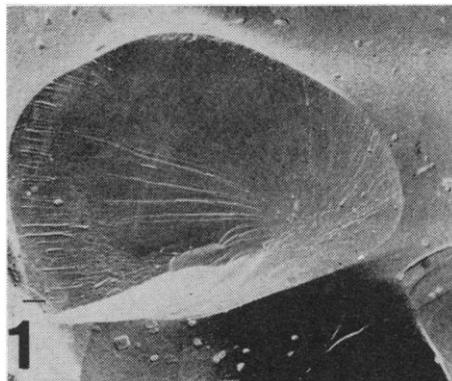


Fig. 1. Fractured stereom column and smooth surface of echinoid plate interior ($\times 5000$). Fig. 2. Conchoidal fracture of plate with adjacent latticework opening ($\times 5000$).

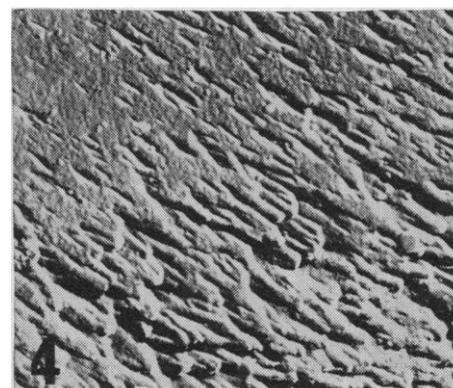
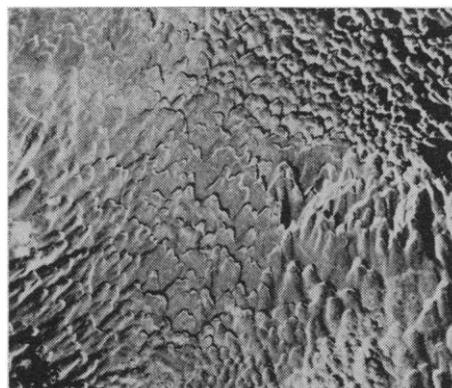


Fig. 3. Oriented needles and laths of plate exterior ($\times 10,000$). Fig. 4. Plate exterior with oriented laths and microstriae ($\times 15,000$).

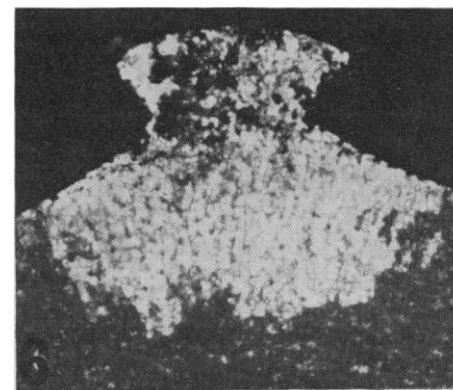


Fig. 5. Microstriae parallel to long axis of echinoid spine ($\times 8500$). Fig. 6. Polycrystalline tubercle in crossed-polarized light ($\times 100$).

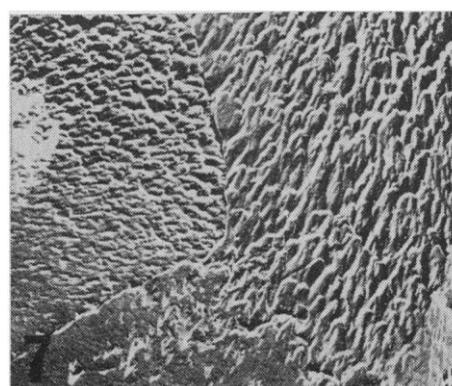


Fig. 7. Mosaic grains made of laths and needles at surface of tubercle ($\times 10,000$). Fig. 8. Coexisting needles and conchoidal fracture of echinoid plate ($\times 10,000$).

preferred orientation of crystallites as in a polycrystalline aggregate. In some photographs, these two types coexist (Fig. 8). This may be indicative of the method of development of the echinoderm skeletal element—a process of oriented polycrystalline growth on, or in, an organic matrix followed by maturation which appears to involve recrystallization by continual fusion and coalescence.

Such an hypothesis allows for the absence of crystal faces within the mature stereom. A recrystallization in the solid state produces a pseudomorph, the shape of which is determined by the polycrystalline aggregate and the original organic framework. Most of the skeletal stereom occurs in the mature state, and, except for the tubercles, only the actively growing surfaces are polycrystalline. Thus, the echinoderm skeletal elements are neither sin-

gle crystals nor polycrystalline aggregates, but they are a combination of both. These data help to reconcile the contradictions in some of the studies of earlier workers.

KENNETH M. TOWE

*Department of Paleobiology,
Smithsonian Institution,
Washington, D.C. 20560*

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Immunoglobulin Structure: Variation in Amino Acid Sequence and Length of Human Lambda Light Chains

Abstract. Variation and conservation in the primary structure of human lambda light chains is revealed by complete amino acid sequence of three Bence Jones proteins. These proteins differ in amino acid sequence in from 38 to 48 positions; they are of unequal length in the amino-terminal half of the chain but have identical sequence in the last 105 amino acids.

Development of the concept that the light chains of immunoglobulins have a variable NH₂-terminal half and an essentially invariant COOH-terminal half began in 1963 with the suggestion of Putnam *et al.* (1) based on peptide maps that "All Bence Jones proteins of the same antigenic type share a fixed portion of their sequence and also have a mutable part." The concept was explicitly proposed on the basis of partial analysis of the amino acid sequence of κ -type Bence Jones proteins (2) and has received strong support as a result of complete amino acid sequence analysis of one κ -type (3) and one λ -type (4, 5) Bence Jones protein of man, and of extensive sequence analysis of two κ -type Bence Jones proteins from the mouse (6). We have now verified this concept for human λ -chains by amino acid sequence analysis of two more λ -type Bence Jones proteins reported herein. Further support is offered by the confirmation (7) of the sequence we have reported for the COOH-terminal half of human λ -chains (5).

We now report the complete amino acid sequence of two human λ -type Bence Jones proteins (designated Ha and Bo, respectively) in comparison to the complete sequence we have published for the human λ -type protein designated Sh (5). In Fig. 1, a different numbering system is employed for the Ha and Bo λ -chains since their length is greater than that of the Sh λ -chain. This is given underneath the sequence for protein Ha. However, all citations in the following text refer to the Sh numbering system given over protein Sh in Fig. 1. [This practice is analogous to that introduced by Braunitzer *et al.* (8) for sequence comparison of the α -chains and β -chains of hemoglobin, which also are of unequal length.] With this alignment the amino acid sequence of all human λ -chains is presumably identical, from Gln-109 through the COOH-terminal residue Ser-213 (9) with the exception that in the Oz(+) serological subtype the arginine at position 190 is replaced by lysine (10). Hence, the amino acid sequence is given in Fig. 1 only for the

NH₂-terminal portions of proteins Ha and Bo through the residue denoted Gly-108 in the Sh numbering system.

The conclusion that the sequence of the last 105 residues in human light λ -chains is essentially invariant is based on the following findings. By methods already described (4, 5), we have isolated from proteins Ha and Bo tryptic peptides corresponding to all those in protein Sh, beginning with Ala-112 through the COOH-terminal residue Ser-213. Without exception, all the corresponding tryptic peptides for this region of the three proteins [namely peptides T₁₆, T₂, T₁₈, T₁₂, T₁, T₂₀, T₇, T₁₁, T₁₉, and T₁₄ of Wikler *et al.*, (5)] are identical in amino acid composition, in NH₂-terminal and COOH-terminal groups (in those cases determined), and in such portions of their sequence as we have already completed. These peptides from proteins Ha and Bo also have the same elution position in ion-exchange chromatography and the same electrophoretic mobility at pH 3.7 as the corresponding peptides isolated from the Sh protein. Insofar as the data have been obtained, the same statements apply to the chymotryptic peptides we have isolated from this portion of the three λ -chains. Furthermore, the published sequence of our λ -chain Sh is identical in every detail (5), except for the position of one amide group, with a sequence from Gln-109 through the COOH-terminus Ser-213 for another λ -type Bence Jones protein (X) (7). Thus, residues 109 to 213 are identical in all four λ -type proteins (Sh, Bo, Ha, and X). On the other hand, the position corresponding to Gly-108 in protein Sh is occupied by arginine in proteins Ha and Bo (Fig. 1) and by serine in protein X.

Like κ -chains (3), human λ -chains differ in many positions in the NH₂-terminal portion of the molecule. Exactly half of the 108 positions in the variable part of the lambda-type protein Sh are identical with corresponding positions in the lambda-type proteins Bo and Ha (Table 1). However, the remaining 54 positions in this portion of the λ -chain Sh differ from one or both of the other λ -chains (Fig. 1). The positions of variation are distributed in a seemingly random fashion throughout the first half of the λ -chains; yet, there are two short peptide stretches where the amino acids differ in all three λ -chains (residues 48 to 51 and 91 to 95). Conversely, up to residue 108 there are four tetra-