

The method used for relabeling is essentially a repetition of the original process. After the unreacted fluorescein derivatives have been dialyzed away, the labeled protein is simply shaken overnight again with the proper amount of dried isocyanate. Since the protein solution is already in approximately 1 gram percent concentration, no more saline is added. However, buffer is again added (10 percent of the original volume of the globulin).

In our opinion, the main advantages to be obtained from the use of dried isocyanate in the manner described above are (i) that the isocyanate can be prepared centrally in either commercial or noncommercial laboratories and can then be sent out to smaller research or diagnostic laboratories for actual use, and (ii) that an antiserum can be made to stain more intensely by relabeling, without danger of loss of protein content as a result of denaturation.

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References and Notes

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3. M. Goldman, *J. Exptl. Med.* 105, 557 (1957).
4. J. M. Marshall, Jr., *ibid.* 94, 21 (1951); personal communication from A. H. Coons, who kindly supplied details of unpublished methods he uses.
5. M. Goldman, *J. Exptl. Med.* 105, 549 (1957).
6. The possibility of performing the labeling reaction in the absence of organic solvents was suggested by R. Goldwasser, research fellow, Laboratory Branch, Communicable Disease Center, Atlanta, Georgia.
7. H. Holter and J. M. Marshall, Jr., *Compt. rend. trav. lab. Carlsberg Sér. chim.* 29, 7 (1954).

31 July 1957

Mortality of Aquatic Insects and Fishes Caused by Use of Latex Tubing in Experimental Apparatus

A commonly used brand of latex rubber tubing was employed in the construction of apparatus for experiments on the dissolved oxygen requirements of certain aquatic insects under varying conditions of flow. A unit of the apparatus consisted of a glass tube through which water was continually recirculated by means of a small centrifugal pump. About 10 ft of 1/2-in. latex tubing provided the connections between the tube, a 5-gal bottle in which the dissolved gas content of the water was adjusted, and the pump. The test animals were held in the tube in a capsule of stainless-steel screen. The total volume of water in use at any one time was about 20 lit. During any one experiment there was no ex-

Table 1. Mortality associated with latex tubing in standing water tests.

Test No.	Animal	No.	Test conditions	Animals dead in		
				24 hr	48 hr	72 hr
1a	<i>A. pacifica</i>	10	Latex tubing		3	7
1b	<i>A. pacifica</i>	10	(control)		0	0
2a	<i>A. californica</i>	10	Latex tubing		5	6
2b	<i>A. californica</i>	10	(control)		0	0
3a	<i>L. reticulatus</i>	10	Latex tubing	0	0	5
3b	<i>L. reticulatus</i>	10	(control)	0	0	0
4a	<i>O. tshawytscha</i>	10	Latex tubing	10		
4b	<i>O. tshawytscha</i>	10	(control)	0		
5a	<i>O. tshawytscha</i>	10	Latex tubing	10		
5b	<i>O. tshawytscha</i>	10	(control)	0		
6	<i>A. californica</i>	10	Plastic tubing		0	0

change of the water. Test water temperatures were about 20°C.

In numerous trials it proved to be impossible, with the use of this apparatus, to keep more than a small percentage of the nymphs of *Acroneuria pacifica* and *Acroneuria californica* (Plecoptera) alive and in good condition, even under "control" conditions, for periods of several days. Usually, within 24 hours, some mortality occurred, and all or most of the surviving nymphs were in sluggish condition. Generally, in 2 or 3 days, about half of the nymphs were dead or moribund.

Subsequent testing of individual parts of the apparatus for harmful effects on the nymphs indicated that only the latex tubing could be responsible for the trouble. Confirmatory tests were conducted, in 1-gal glass jars, in which the two species of *Acroneuria*, fingerling king salmon (*Oncorhynchus tshawytscha*), and immature guppies (*Lebistes reticulatus*) were used. In these experiments a piece of latex tubing several feet long was coiled in the test jars filled with water. The animals were then introduced into the test water, which was kept well aerated and in continuous circulation by rising air bubbles. Appropriate controls were used. As Table 1 shows, no losses occurred among the control animals, but in all cases there was considerable mortality among the test animals, of all species, in the jars that contained the tubing. A piece of plastic tubing was used in a similar jar test. No deaths occurred, and the stonefly nymphs used appeared to be normally active after 3 days.

When about 1500 ml of water per minute flowed continuously and without recirculation or reuse through 10 ft of the latex tubing and into a glass tube containing stonefly nymphs, no difficulty was experienced in keeping the animals alive and in good condition for several days.

As a result of these experiments, the original apparatus was reconstructed and the plastic tubing, which had been found to be apparently harmless, was used; in the course of tests performed thereafter,

with this apparatus, no unexpected mortality of stonefly nymphs occurred.

Although widely used and apparently entirely suitable for most experimental purposes, latex rubber tubing evidently should be used with caution in experiments involving the recirculation of water in closed systems where there is no continuous exchange of water and where species of animals sensitive to the constituents of the tubing are utilized. Even though mortality might not result from such use of the tubing, the results of sensitive physiological tests might be markedly influenced.

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8 July 1957

Synthesis of "On-Off" and "Off" Responses in a Visual-Neural System

The most distinctive feature of the discharge of impulses in the vertebrate optic nerve in response to a light stimulus is the marked activity elicited by changes in the level of illumination. The early records of Adrian and Matthews from the whole optic nerve (1) demonstrate a strong burst of activity when the light is turned on, a continuing discharge at a lower rate as long as the light remains on, and, upon the cessation of light, a renewed burst which gradually subsides.

Hartline (2) has shown that this composite response results from individual fibers whose activity differs markedly: some fibers discharge regularly as long as the light shines; others discharge only briefly when the light is turned on and again when it is turned off, with no activity during steady illumination; still others respond only when the light is turned off. These complex responses, observed in third-order neurones, have been ascribed by Hartline (2) and Granit (3)