

Extraction of C₂₁O₅ and C₂₁O₆ Steroids from Aqueous Media

Although a great variety of solvents have been used for the extraction of corticosteroids, it seems, as has been pointed out by Mason (1), that "solvents have been chosen as a matter of personal preference rather than on the basis of any study of their merits and shortcomings." By and large, the chlorinated hydrocarbons have become the favorite solvents after the introduction of ethylene chloride as a selective solvent for cortical hormones by Cartland and Kuizenga (2). Although Reichstein (3) indicated the excellent extraction properties of ethyl acetate for corticosteroids, relatively little use has been made of this solvent until quite recently, when Meyer (4), after testing a variety of solvents for extraction of blood, found that ethyl acetate was particularly favorable, since it extracted smaller amounts of extraneous material and did not form emulsions.

Owing to its relative nonemulsifying properties, as discovered by Meyer, ethyl acetate was subsequently introduced for extraction of corticosteroids from guinea pig and human urine (5-7). In these studies, in which the more polar solvent

ethyl acetate was used, the two C₂₁O₆ steroids 6 β -hydroxycortisol and 2 α -hydroxycortisol have been isolated. In this connection, the finding of corticosteroids with mobilities on paper of C₂₁O₆ steroids in adrenal extracts obtained by the use of ethyl acetate as the extracting solvent is noteworthy (8). The fact that in previous studies, in which methylene chloride was used, these C₂₁O₆ steroids were not observed except in trace amounts (9) has made it pertinent to study the extractability of C₂₁O₆ steroids from aqueous media.

In this report (10) partition coefficients of some C₂₁O₅ and C₂₁O₆ steroids for the two systems chloroform/water and ethyl acetate/water are presented (Table 1). The partition coefficients (expressed as $K = C_{org}/C_{aq.}$) were determined by shaking the solvents containing the steroids (100 to 600 γ) with an equal volume of water (5 ml). Solvents that were not previously saturated with respect to each other were used, since the partition coefficients were required for calculations on batch extraction with new solvent. The partitions were done at room temperature. Determinations were done on aliquots from each phase.

It appears from Table 1 that ethyl acetate is a better extracting solvent than

chloroform for both the C₂₁O₅ and C₂₁O₆ steroids studied. From the partition coefficients of the C₂₁O₆ steroids given, it becomes clear that chloroform (similar data were obtained with methylene chloride) is not a suitable extraction solvent for these steroids. Since in the usual extraction procedure the original extract is further purified by washing with alkali and water to remove acidic interfering material, only small yields of C₂₁O₆ steroids can be obtained owing to the appreciable extraction by the water phase.

In view of the relatively low partition coefficients of some C₂₁O₅ steroids for the CHCl₃/H₂O system, a reevaluation of the significance of earlier quantitative studies in which chlorinated hydrocarbons have been used will be necessary. Thus, the relatively small yields of 3 α , 11 β , 17 α , 21-tetrahydroxypregnan-20-one isolated from human urine following cortisol feeding undoubtedly have been the result of the aqueous washes of the methylene chloride extracts during the extraction and fractionation procedure (9). The present data indicate the necessity for further work on extraction and separation of highly hydrophilic steroids, since increasing the polarity of the solvents decreases the extraction selectivity and presents greater difficulties in isolation.

SHLOMO BURSTEIN*

Worcester Foundation for Experimental Biology, Shrewsbury, Massachusetts, and Laboratory of Pathology, National Cancer Institute, National Institutes of Health, Bethesda, Maryland

Table 1. Partition coefficients of some C₂₁O₅ and C₂₁O₆ steroids.* The determinations were done by ultraviolet spectrophotometry in methanol with the Beckman DU (UV), by the phosphomolybdic acid (P.M.), or the Porter-Silber (P.S.) reactions as indicated.

No.	Steroid	K CHCl ₃ / H ₂ O	K EtOAc/ H ₂ O	Determination method
1.	11 β , 17 α , 21-Trihydroxy-4-pregnene-3,20-dione	6.4	12.2	UV at 240 m μ ; P.M. and P.S.
2.	11 β , 17 α , 21-Trihydroxy-4,6-pregnadiene-3,20-dione	4.8	12.6	UV at 280 m μ
3.	11 β , 17 α , 21-Trihydroxy-1,4-pregnadiene-3,20-dione	3.6	11.2	UV at 242 m μ
4.	3 α , 11 β , 17 α , 21-Tetrahydroxypregnan-20-one†	2.4	11.0	P.M.
5.	11 β , 17 α , 20 β , 21-Tetrahydroxy-4-pregnen-3-one	1.1	1.6	UV at 240 m μ
6.	11 β , 17 α , 20 α , 21-Tetrahydroxy-4-pregnen-3-one	0.9	1.7	UV at 242 m μ
7.	6 β , 17 α , 21-Trihydroxy-4-pregnene-3,11,20-trione	0.3	2.2	UV at 233 m μ
8.	9 α , 11 β , 17 α , 21-Tetrahydroxy-4-pregnene-3,20-dione	0.1	2.0	UV at 242 m μ
9.	2 α , 11 β , 17 α , 21-Tetrahydroxy-4-pregnene-3,20-dione	0.3	1.9	P.S.
10.	6 β , 11 β , 17 α , 21-Tetrahydroxy-4-pregnene-3,20-dione	0.05	0.9	P.S.

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† For unexplained reasons, variable results were obtained with steroid 4 during the chloroform-water partitions with low recoveries from the aqueous phase. The value given in the table represents the average of three determinations in which the total recoveries were above 90 percent.

References and Notes

1. H. L. Mason, *Recent Progr. Hormone Research* 9, 267 (1954).
2. G. F. Cartland and M. H. Kuizenga, *J. Biol. Chem.* 116, 57 (1936).
3. T. Reichstein, *Helv. Chim. Acta* 19, 29 (1936).
4. A. S. Meyer, *J. Biol. Chem.* 203, 469 (1953) and *Recent Progr. Hormone Research* 9, 379 (1954).
5. S. Burstein and R. I. Dorfman, *J. Biol. Chem.* 206, 607 (1954).
6. —, *ibid.* 213, 581 (1955).
7. S. Burstein, R. I. Dorfman, E. M. Nadel, *ibid.* 213, 597 (1955) and *Arch. Biochem. and Biophys.* 53, 307 (1954); E. M. Nadel, S. Burstein, R. I. Dorfman, *Arch. Biochem. and Biophys.* 61, 144 (1956); S. Burstein, *J. Am. Chem. Soc.* 78, 1769 (1956); E. M. Nadel, S. Burstein, R. I. Dorfman, *Proc. Am. Assoc. Cancer Research* 2, 37 (1955).
8. I. E. Bush, *Biochem. J.* 50, 370 (1952); R. Haynes, K. Savard, R. I. Dorfman, *J. Biol. Chem.* 207, 925 (1954).
9. S. Burstein, K. Savard, R. I. Dorfman, *Endocrinology* 53, 88 (1953); S. Burstein, unpublished observations.
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* Present address: College of Physicians and Surgeons, Columbia University, New York, N.Y.

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