lyophiled protein for tissue culture studies, we made quantitative investigations of bacterial survival after certain combinations of chemical treatment and lyophiling.

The test medium chosen was Brewers thioglycollate broth⁴ because it supports the growth of anaerobes as well as aerobes.

A local contaminant (gram positive spore-former) was used as a test organism, since previously it had been shown to be more resistant to chemical and thermal treatment (toluene, ether, chloroform, acetone, propylene glycol, various degrees of heat) than any of four standard test organisms,⁵ Staph. aureus, Staph. albus, Ps. pyocyanea, B. subtilis. The organism was always used as a 24-hour culture of a 24-hour culture.

The results of these experiments are summarized in Tables I and II.

TABLE 1

STERILIZING EFFECT OF CHEMICAL ADDED TO THE DRY RESI-DUE AFTER LYOPHILING. NUMBERS EQUAL DUE AFTER LYOPHILING. NUMBER VIABLE ORGANISMS/ML.

		Incubation 20 hours 37° C.	Lyophiled 24 hours	Lyophiled 24 hours plus Tol- uene	Relyo- philed
Experiment Experiment	$\frac{1}{2}$	2,900,000 2,800,000	100,000	130,000	30
STERILIZING	E	T FFECT OF CI MEDIUM BI	ABLE 2 HEMICALS AI FFORE LYOPH	DED TO THE ILING	AQUEOUS

Incubation 20 hours 37° C.	Lyophiled 24 hours	
1,100,000	53,000	
Less than 2,000	0	
Less than 2,000	0	
	Incubation 20 hours 37° C. 1,100,000 Less than 2,000 Less than 2,000	

These results indicate that reasonably clean handling of the proteinaceous material, plus toluene treatment and lyophiling, will result in a sterile product.

The material from which proteins intended for subsequent sterile use are to be prepared should be handled in a clean way to reduce sources of contamination. Sufficient toluene should be added to saturate the original material, which then should be worked up in a cool room as quickly as possible to avoid multiplication of the contaminants present. If the product is not sterile, after lyophiling, it may be dissolved in the amount of distilled water removed, toluene again added and relyophiled. In a planned experiment, using organisms in prime condition and using a selective medium, the organisms became non-culturable.

Preparations of fibrinogen, thrombin and plasma, all from slaughter-house beef blood, and of chick embryo

4 J. H. Brewer, Jour. Bact., 30: 10, 1940.

⁵ M. S. Marshall, J. B. Gunnison, M. P. Luxen, Proc. Soc. Exp. Biol. Med., 43: 672, 1940.

extract handled in the open air in non-sterile containers, have been obtained in the lyophiled state in sterile condition in this manner.

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REMOVING FROZEN PLUNGERS OF GLASS SYRINGES

THE method of removing the plungers of frozen glass syringes suggested by McCoord¹ was formerly used in our laboratory exactly as he describes it. However, the difficulty in some cases due to the leakage of the water around the rubber gasket caused me to devise a somewhat surer method.

The same principle is used as that suggested by McCoord in which a small tuberculin syringe is utilized to give the necessary hydraulic pressure. Instead of connecting the two syringes by a hypodermic needle and rubber gasket, we have made use of a connector fashioned from the hubs of two broken hypodermic needles. These are brazed together so that they can be used to join the two syringes. In order to make certain that the hole in the needles has not been plugged up in the process of brazing they are subsequently drilled out with a small metal drill. It is important that all air be removed from the system before exerting pressure. This can be accomplished in the "frozen" syringe by using a fine neeedle which will pass freely through the nipple.

This method has been used in our laboratory for the past five years and has never failed to release the plungers even though they are sealed by coagulated blood.

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¹ SCIENCE, Vol. 94, p. 170.

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