

Gently does it. In their AFM experiments, Lantz *et al.* can distinguish between silicon atoms at different surface locations on the basis of their force response curves (**left**). Shown here are four unit cells of the silicon 7×7 surface, with a corner hole at the center (marked with an x). This site serves as a reference to measure background forces. The corner adatom (green) and the center adatom (red) have different stiffnesses, as indicated by the curves of the same color in the graph. This difference is exaggerated for clarity. The tip is shown at the point of closest approach, where the chemical bond (yellow cloud) starts to form.

which is likely if the tip picks up some of the material from the surface, a perfect image showing the periodicity of the sample is obtained, but local defects do not show up. This can be easily understood by taking two washboards and sliding one on top of the other. The resulting up/down motion will be the same even if you remove one of the ridges from one washboard.

"True" atomic resolution was not achieved until 1995, when the 7×7 reconstruction of silicon (a beautiful rearrangement of the surface atoms) was imaged with an AFM (3, 4). It was the atomic-scale imaging of this surface that gave the scanning tunneling microscope (STM), the older sibling of the AFM, instant fame in 1983 (5). Lantz et al. chose this classic surface for their experiment. It is ideal for this first demonstration because the atoms are widely spaced and each has a "lone" atomic orbital sticking out of the surface. All the atoms are silicon, but their stiffness differs nevertheless depending on their position, which determines the way they bond to their neighbors underneath. This type of force spectroscopy has been done on surfaces for some time (6), but this is the first time that the surface is also imaged with atomic resolution in the same experiment (after all, you have to take the blindfold off to check that vour identification of the fruit is correct!).

In a recent related study, Foster *et al.* identified the fluorine atoms in CaF_2 on the basis of the shape of the features in the AFM image, aided by theoretical modeling (7). The observed triangular shape of the

is caused by the contribution to the image from fluorine atoms just below. This result is important because this type of sample, being an insulator, cannot be imaged with STM. Giessibl et al. have also shown that under special conditions, it is possible to image individual orbitals such as those of an atom on the tip (8) This work has elicited criticism (9)and a technically compelling response (10, 11). The idea of squeezing microscopic samples with an AFM tip is also being applied to biological specimens. Mahaffy et al. have

surface fluorine atoms

recently made quantitative viscoelastic measurements on fibroblasts at the submicrometer level and concluded that the response is dominated by the cytoskeleton (12).

The AFM studies discussed here were performed in the so-called noncontact mode, which uses the weak attractive forces between tip and sample for imaging. The area of noncontact AFM with true atomic resolution is starting to blossom (13, 14). Over the short term, we can expect improvements through the use of stiffer can-

PERSPECTIVES: CARBON CYCLE

tilevers and smaller oscillation amplitudes (8). The use of carbon nanotubes as tips (15) promises to improve the weakest link in these experiments. Once we have identified surface atoms and their bonding strengths, we will be able to do more atomic manipulation (16), even at room temperature (17). The use of sophisticated tactile ("haptic") interfaces (18, 19) will allow us to "feel" the atoms in real time and gently push them to desired locations. We may at last be able to take off the boxing glove.

References and Notes

- 1. M.A. Lantz et al., Science 291, 2580 (2001).
- G. Binnig, C. F. Quate, Ch. Gerber, *Phys. Rev. Lett.* 56, 930 (1986).
- 3. F. J. Giessibl, Science 267, 68 (1995).
- S. Kitamura, M. Iwatsuki, Jpn. J. Appl. Phys. 34, L145 (1995).
- G. Binnig, H. Rohrer, Ch. Gerber, E. Weibel, *Phys. Rev. Lett.* 50, 120 (1983).
 S. P. Jarvis, H. Yamada, S.-J. Yamamoto, H. Tokumoto, J.
- S. P. Jarvis, H. Yamada, S.-I. Yamamoto, H. Tokumoto, J. B. Pethica, Nature **384**, 247 (1996).
- A. S. Foster, C. Barth, A. L. Shluger, M. Reichling, *Phys. Rev. Lett.* 86, 2373 (2001).
- F. J. Giessibl, S. Hembacher, H. Bielefeldt, J. Mannhart, Science 289, 422 (2000).
- H. J. Hug et al., Science, 30 March 2001 (www. sciencemag.org/cgi/content/full/291/5513/2509a).
 F. J. Giessibl et al., Science, 30 March 2001 (www.
- F. J. Giessibl et al., Science, 30 March 2001 (www sciencemag.org/cgi/content/full/291/5513/2509a).
- New data supporting Giessibl *et al.*'s response was presented on 14 March 2001 at the Spring meeting of the American Physical Society in Seattle (*Bull. Am. Phys. Soc.* 46, 846 (2001).
- R. E. Mahaffy, C. K. Shih, F. C. MacKintosh, J. Kas, *Phys. Rev. Lett.* 85, 880 (2000)
- S. Morita and M. Tsukada, Eds., Proceedings of the First International Workshop on Non-Contact Atomic Force Microscopy, Appl. Surf. Sci. 140, 1 (1999).
- Force Microscopy, Appl. Surf. Sci. 140, 1 (1999).
 R. Bennewitz, C. Cerber, E. Meyer, Eds., Proceedings of the Second International Workshop on Non-Contact Atomic Force Microscopy, Appl. Surf. Sci. 157, 1 (2000).
- 15. V. Barwich et al., Appl. Surf. Sci. 157, 269 (2000)
- 16. D. M. Eigler, E. K. Schweizer, *Nature* **344**, 524 (1990). 17. T. W. Fishlock, A. Oral, R. G. Egdell, J. B. Pethica, *Na*-
- *ture* **404**, 743 (2000). 18. M. Sincell, *Science* **290**, 1530 (2000).
- M. Sincell, Science 290, 1550 (2000).
 See also www.cs.unc.edu/Research/nano.
- **Discriminating Plants**

F. I. Woodward

The increasing concentration of carbon dioxide (CO_2) in the atmosphere (1) is the surest evidence that humans are changing the global environment. Atmospheric CO_2 would accumulate even faster if the oceans and the terrestrial biosphere did not absorb about half the CO_2 emissions from burning fossil fuels and deforestation. The amounts of CO_2 absorbed by these sinks are difficult to quantify, however, and this is hampering a detailed understanding of the carbon cycle.

Recent studies suggest that plants dis-

criminate in a systematic way against CO_2 containing the stable isotope ¹⁸O, offering an exciting opportunity to differentiate the terrestrial from the oceanic sink (2, 3); the greater the terrestrial sink the lower the ¹⁸O content of atmospheric CO_2 (4). However, Gillon and Yakir (5) suggest on page 2584 of this issue that current understanding of this discrimination may be incomplete, resulting in incorrect sink estimates.

Both transpiration and photosynthesis are involved in the discrimination of plants against CO₂ containing ¹⁸O (C¹⁸O¹⁶O) (see the figure). H₂¹⁶O transpires more rapidly from leaves than the heavier H₂¹⁸O. The site of evaporation therefore becomes enriched in the heavier isotope (3). This

The author is in the Department of Animal and Plant Sciences, University of Sheffield, Sheffield, S10 2TN, UK. E-mail: F.I.Woodward@Sheffield.ac.uk

evaporative ¹⁸O enrichment in water is transferred first to the water in the chloroplast (where photosynthesis takes place) and then, by the enzyme carbonic anhydrase, to dissolved CO_2 in the chloroplast (2). CO_2 entering the leaf thus equilibrates isotopically with chloroplast water.

Plants fall into two groups determined by their photosynthetic pathway. About

85% of plants use the C_3 pathway, with the remainder using the more recently evolved C₄ pathway. A fraction F of the CO₂ diffuses out of the leaf without being fixed photosynthetically but after isotope exchange with chloroplast water. In C₃ plants, F is about 50% of the inward flux of CO₂. In contrast, F is only about 30% in C₄ plants, where the photosynthetic rate is higher and the CO₂ concentration in the chloroplast lower. This outward flux discriminates against C¹⁸O¹⁶O; that is, C¹⁸O¹⁶O is preferentially retained in the plant. Previous studies have assumed complete isotopic equilibrium

SCIENCE'S COMPASS

¹⁸O-enriched CO₂. The ¹⁸O-enriched CO₂ flux from global vegetation was only 78% of that expected for equilibrium with leaf water. This indicates the substantial impact of the incomplete ¹⁸O equilibrium in vegetation dominated by C₄ species. Back calculations from these fluxes assuming isotopic equilibrium would underestimate gross primary production (GPP) by about



Discriminating leafs. The fluxes of $H_2^{18}O$ (purple) and $C^{18}O^{16}O$ (red) into and out of a leaf through a stoma to the sites of evaporation (cell surfaces). Expanded inset: CO_2 exchange in the chloroplast. *CA*, carbonic anhydrase.

between water and CO_2 when calculating this discrimination.

On the basis of measurements of over 50 species, Gillon and Yakir propose that the assumption of complete isotopic equilibrium, due to sufficient carbonic anhydrase activity, requires reconsideration. Their in situ measurements of carbonic anhydrase indicate that isotopic exchange between water and CO₂ is virtually complete for C_3 and C_4 dicotyledonous species (a large class of flowering plants comprising many forest trees, potatoes, beans, and cabbages, as well as roses and clematis). These data are in keeping with previous research (3, 6), including studies that considered the possibility that isotopic exchange between water and CO₂ was incomplete (nonequilibrium) in C₃ species (2, 7). For grasses, however, Gillon and Yakir find that isotopic exchange is incomplete, at about 75% for C3 grasses and just 38% for C₄ grasses, where carbonic anhydrase activity, CA, is both low and variable between species.

For C₄ species with low chloroplast CO₂, low CA, and therefore a lack of isotopic equilibrium, these new results indicate a smaller impact on atmospheric C¹⁸O¹⁶O than indicated by the original equilibrium theory. Gillon and Yakir have expanded these estimates using data on the distribution of vegetation types, including C₄ species, to estimate the global fluxes of

22%, with particularly marked underestimates of C_4 activity.

The authors' comparisons between nonequilibrium and equilibrium estimates of ¹⁸O-enriched CO_2 fluxes assume that the calculated discrimination against C¹⁸O¹⁶O (Δ_A) is unchanging for the two cases. This can only be achieved by changing the global estimates of the ¹⁸O composition at the site of evaporation in leaves. This is contentious but could explain discrepancies between different methods of estimation. Even when Δ_4 is the same for nonequilibrium and equilibrium conditions, there is still the need to account for the forcing of atmospheric $C^{18}O^{16}O$ concentration by the outward flux of CO_2 from the leaf. This requires that the gradient for C¹⁸O¹⁶O exchange is considered from the chloroplast surface rather than the chloroplast itself, as suggested previously by Gillon and Yakir (6). This would lead to an 18% underestimate of GPP calculated from changes in atmospheric C¹⁸O¹⁶O and, assuming equilibrium, slightly less than the 22% calculated by Gillon and Yakir but still a substantial error.

One of the key points raised by Gillon and Yakir is that grasses, and C₄ grasses in particular, show the greatest disequilibrium between ¹⁸O in water and CO₂. C₄ dicotyledons do not show this effect, indicating that nonequilibrium is not simply associated with the C₄ pathway of photosynthesis. Water supply to grass leaves occurs linearly and sequentially from base to tip; the 18 O composition of source water should become enriched by evaporation along this pathway. It has been shown for C₃ and C₄ grasses (8) that the longer the leaf, the greater the enrichment of the source water and the greater the deviation of the 18 O concentration of the enriched water calculated from a constant source water compo-

sition. Longer leaves (typifying C_4 species) therefore have a greater enrichment. Perhaps the large scatter and high disequilibrium values for C_4 grasses estimated by Gillon and Yakir result, at least in part, from variations in leaf length and the necessity to account for the linear evaporative enrichment of ¹⁸O. Helliker and Ehleringer (9) have also found low and very variable *CA* in C₄ grasses, in keeping with Gillon and Yakir's results.

Precise quantification of 18 O discrimination by plants and ecosystems (10) is still not possible, but the work of Gillon and Yakir has brought us one step closer to this important goal. Ultimately, this

approach will provide information about the carbon cycle unmatched by any other current method.

References and Notes

- C. D. Keeling, T. P. Whorf, in *Trends: A Compendium of Data on Global Change* (Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, U.S. Department of Energy, Oak Ridge, TN, 2000).
- 2. G. D. Farquhar et al., Nature 363, 439 (1993).
- 3. P. Ciais et al., J. Geophys. Res. 102, 5873 (1997).
- 4. The ¹⁸O content of atmospheric CO₂ is most strongly influenced by the different ¹⁸O contents of water in the oceans and in precipitation. This leads to a large latitudinal gradient in the ¹⁸O content of CO₂ but with little effect due to marine productivity. By contrast, leaves of terrestrial plants annually influence about one-third of the ¹⁸O content of atmospheric CO₂. In addition, the rate of exchange in the ¹⁸O content of CO₂ is about four times higher for terrestrial plants than for all oceanic exchange (2).
- J. Gillon, D. Yakir, *Science* **291**, 2584 (2001); published online 8 March 2001 (10.1126/science.1056374).
- 6. _____, Plant Physiol. 123, 201 (2000).
- 7. L. B. Flanagan *et al.*, *Aust. J. Plant Physiol.* **21**, 221 (1994).
- B. R. Helliker, J. R. Ehleringer, Proc. Natl. Acad. Sci. U.S.A. 97, 7894 (2000).
- 9. _____, personal communication.
- 10. Unpublished work (C. Still, B. R. Helliker, M. Ribas-Carbo, S. Verma, J. A. Berry, unpublished data) in a tall-grass prairie, dominated by C₄ grasses, but with C₃ grasses active during spring, has indicated that differentiating ¹⁸O discrimination by C₃ and C₄ grasses could only be achieved by flux corrections that assume a lack of isotopic equilibrium. Respiratory fluxes from the soil are particularly substantial, however, complicating differentiation between the sources of fluxes in the field.

Published online 8 March 2001;

10.1126/science.1059472 Include this information when citing this paper.