Children with positive DHT responses to tuberculin had serum cytokine concentrations suggestive of predominant T_H1 responses, in contrast to the T_H^2 profiles seen in children with negative DHTs.

These results are an important extension of observations in the 1960s and 1970s that there is a reciprocal relation between inflammatory and humoral responses to vaccination regimes (7, 8). This reciprocal relation has also been attributed to preferential activation of $T_{\rm H}1$ or $T_{\rm H}2$ subsets of T cells and is consistent with the genetic predisposition to T_H1 or $T_{\rm H}^2$ responses of different strains of mice.

Central to the relevance of the results is the hypothesis that the immune system can be manipulated to manifest a persistent T_H1 or T_{H2} response. If this is the case, vaccination to induce T_H1 responses may be effective against asthma and other allergic disorders (9). In mice, overwhelming Schistosoma mansoni infection induces T_H^2 responses. The infection concomitantly down-regulates the $T_{\rm H} 1$ response to other antigens and delays the clearance of vaccinia virus (10). However, in humans with filariasis, who show T_H2-biased cytokine profiles, the ability to respond to Mtb proteins is not lost (11). Children with eczema, another atopic condition, occasionally undergo spontaneous remission after severe bacterial or viral infections (12), although usually temporarily. Both of these observations suggest that alterations in the $T_{\rm H}2/T_{\rm H}1$ balance may become important only in the presence of continued overwhelming infection.

Also confusing the T_H1/T_H2 theory of asthma are the findings that helminth and other parasite infection may protect against allergic diseases, despite up-regulation of T_{H2} responses. This type of infestation is invoked to explain the low prevalence of asthma in rural Africa and the Venezuelan slums (13, 14). Helminth infection produces high levels of polyclonal IgE that, possibly by saturating the number of binding sites for IgE on mast and other effector cells of allergy, prevent activation of these cells by the relatively trivial exposures to allergens.

Thus, the results of Shirakawa et al. invite the speculation that the decline in childhood tuberculosis infection in Japan is causal in the recent asthma epidemic. However, the incidence of other infections may also be declining, so the case for tuberculosis requires further study. Nevertheless, the new results emphasize the complexity of the environmental contribution to asthma and remind us that identification of the relevant factors may ultimately resolve this epidemic.

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(4). But how signaling through $G\beta\gamma$ is termi-

nated has not been as clear. A report in Cell by Gilman and co-workers (5) and two others in Nature (6) identifying two members of

the RGS family, GAIP and RGS4, as GAPs for members of the $G\alpha$, family and other recent papers shed light on this issue.

identified in yeast, Caenorhabditis elegans,

Members of the RGS family have been

SIGNAL TRANSDUCTION

There Are GAPS and There Are GAPS

Ravi lyengar

GDP

αβγ

Heterotrimeric

G protein

Although their main function is to regulate other proteins, guanine nucleotide-binding proteins (G proteins) are also guanosine triphosphatases (GTPases), cleaving guanosine triphosphate (GTP) to form guanosine diphosphate (GDP). Because of this activity, they oscillate between GTP- and GDP-bound states, and thus regulate diverse processes such as protein synthesis, cytoskeleton assembly, vesicle transport, and signal transduction. The superfamily comprises both small monomeric and large multimeric G proteins, but for all members, the release of bound GDP and the binding of GTP are highly regulated processes (1). The GTPase activity of small G proteins, such as EF-Tu and Ras, is stimulated by associated proteins called GAPs (GTPase activating proteins) (2). But GAPs for most large G proteins had not been described until recent work identified members of the regulators of G protein-signaling (RGS) family as GAPs for this subfamily.

The large heterotrimeric G proteins involved in signal transduction have α subunits, which are related to small G proteins, and $\beta\gamma$ subunits that exist as a single complex. Both $G\alpha$ and $G\beta\gamma$ can independently transmit signals (3). Signal termination for both $G\alpha$ and $G\beta\gamma$ subunits likely occurs through GTP hydrolysis. How the GTPase terminates signaling through $G\alpha$ subunits is easily understood given the observation that GDP-G α subunits have much lower affinities for effectors than do GTP-G α complexes

Activated

receptor

R

GDP

(RGS)(GAP)

Activating

GTPase

Protein

GDP

α



between GBy and GDP interaction with $G\alpha$ subunits



α

GTP

βγ +

Effector

The author is in the Department of Pharmacology, Mount Sinai School of Medicine, New York, NY 10029 USA. E-mail: iyengar@msvax. mssm.edu

Perspectives

(11) indicates that $G\beta\gamma$ has a higher affinity for GDP-G α than for GTP-G α . If G $\beta\gamma$ also has a higher affinity for GDP-G $\!\alpha$ than for effectors such as K⁺ channels, adenylyl cyclase, and phospholipase C, then RGS, by stimulating the GTPase catalytic activity and the accumulation of GDP-G α , could shift the steady-state level toward the inactive heterotrimeric state. Alternatively, if the affinity of G $\beta\gamma$ for GDP-G α and effectors is in the same range, a much greater concentration of $G\alpha$ in comparison to the effector could push $G\beta\gamma$ to the heterotrimeric state. Either way, RGS would be essential to build up the concentration of GDP-G α after activation of the G protein, and the relative stoichiometry of effectors to $G\alpha$ is likely to be important for $G\beta\gamma$ -mediated signaling.

This mechanism may be operative for G_i. Here the $G\alpha$ subunit would define the specificity of receptor interaction and in conjunction with RGS define the lifetime of the active GBy complex. Different G α_i isoforms have different rates of GDP release (12), and if the different RGS isoforms stimulate the GTPase rates to different extents, then a wide range of timing can easily be obtained. Such facile temporal regulation would be particularly valuable for $G\beta\gamma$ regulation of K⁺ and Ca²⁺ channels. RGS stimulation of $G\alpha_{i1}$ GTPase could explain the discrepancy between the 20-fold faster deactivation rate of the muscarinic K⁺ channel upon removal of the agonist, as compared to the basal GTP hydrolysis rate of purified G_i (13). The type of regulation described above presumes that activation results in subunit dissociation and that all of the free $G\beta\gamma$ can regulate effector, although this has not been proven.

Experiments in E. Ross's laboratory have shown that $G\beta\gamma$ inhibits GAP stimulation of the GTPase-G α (14), suggesting that G $\beta\gamma$ and RGS may interact with overlapping regions of $G\alpha$. Additionally, the RGS (GAP) proteins interact with GDP-G α with much lower affinity than they do with the transition-state complex (15). These properties would facilitate GTP hydrolysis on $G\alpha$ followed by dissociation of RGS from $G\alpha$ and the reassociation of $G\alpha$ and $G\beta\gamma$, further supporting the notion that RGS regulated GTPase is the major mechanism for turning off $G\beta\gamma$ signaling. Clearly, for G_i/G_0 coupled systems we have moved from a three- to a four-component system (see figure).

In contrast, GAPs for monomeric G proteins function as On-Off switches because the basal GTPase activity of the monomeric G proteins is very low. RGS is poised to be an effective modulator of transmembrane signaling through G protein pathways and can serve as a putative locus for interactions between signaling pathways.

Comparison of the work by Gilman and co-workers (5, 15) with that from the

Wittinghofer group (6, 17) suggests that GAPs regulate the activity of small and large G proteins by different mechanisms. GDP- $G\alpha$ can bind AlF_4 —small G proteins, such as Ras, cannot, but GDP-Ras protein can bind AlF₄ when associated with its GAP protein (16). Crystallographic studies on $G\alpha$ subunits indicate that the AlF₄-GDP-G α complex represents the transition state of the $S_N 2$ reaction that occurs during GTP hydrolysis (18). Thus for Ras, GAP is thought to contribute residues crucial for formation of the transition state (16). Prominent in this context is an Arg (Arg¹⁷⁸ in $G\alpha_{11}$) present in $G\alpha$ subunits but not in Ras or other small G proteins. The crystal structure of the active domain of the p120GAP and a manual docking model of Ras and the GAP active domain suggest that an Arg from GAP could be used for GTP hydrolysis by Ras (17). Thus, for small G proteins the main function of GAP would be to move the Ras-GTP complex to a transition state for nucleotide hydrolysis. It is unlikely that this is the primary mechanism by which RGS stimulates the GTPase activity of $G\alpha$, since $G\alpha$ by itself can form a transition-state complex (18). However, RGS has the highest affinity for $AlF_4^-GDP\text{-}G\alpha$ (15) indicating that it preferentially binds to the transition-state complex and thus promotes hydrolysis. The precise mechanism by which RGS promotes GTP hydrolysis by $G\alpha$ remains to be determined.

Just when we thought that the basic G protein–signaling system had been well defined, nature has provided us with a surprise. There are probably more surprises to come.

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One of the strengths of the World Wide Web is its capability for disseminating graphics, and few sciences are as rich in images as astronomy. The National Center for Supercomputing Applications at the University of Illinois now has 4000 astronomical images in its Digital Image Library. The Web page allows searching of the database by sky position, name of object, waveband, or bibliographic reference. The images are presented in a variety of formats, including some in the Virtual Reality Markup Language (VRML), which allows rotation and manipulation of data.

The body visible

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Edited by David Voss

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