by each Cu within the $[Cu_3O_2]$ core. Upon inspection of the crystal structure of the fully reduced form of ascorbate oxidase (11), the three trigonally ligated Cu(I) centers (average Cu-Cu distance, 4.5 Å) appear geometrically predisposed toward accommodation of O2 and formation of a $[Cu_3O_2]$ cluster. However, no current spectroscopic studies of the metastable oxygen intermediates of multicopper oxidases and their derivatives support the existence of an intensely absorbing oxo-Cu(III) chromophore, and no unusually short Cu-O bond distances such as those observed in 2 are indicated (12, 13, 32). In accordance with these studies, however, the facile reaction of three Cu(I) monomers with O_2 to form the mixed-valence $bis(\mu_3-oxo)[Cu(II)Cu(II)$ Cu(III)] species 2 does suggest that O_2 bond cleavage at trinuclear Cu sites requires full $4e^-$ reduction of O₂. In the case of native laccase, the fourth electron is provided by the remote "blue" Cu center, whereas in 2, the extra electron must be obtained at the cost of further oxidation of one of the Cu sites.

REFERENCES AND NOTES

- K. D. Karlin'and Z. Tyeklár, Eds., Bioinorganic Chemistry of Copper (Chapman & Hall, New York, 1993).
- 2. N. Kitajima and Y. Moro-oka, *Chem. Rev.* **94**, 737 (1994).
- E. I. Solomon, M. J. Baldwin, M. D. Lowery, *ibid.* 92, 521 (1992); E. I. Solomon and M. D. Lowery, *Science* 259, 1575 (1993).
- K. D. Karlin and S. Fox, in *Active Oxygen in Biochemistry*, J. S. Valentine, C. S. Foote, A. Greenberg, J. F. Liebman, Eds. (Chapman & Hall, Glasgow, 1995), vol. 3, pp. 188–231.
- K. Fujisawa, M. Tanaka, Y. Moro-oka, N. Kitajima, J. Am. Chem. Soc. 116, 12079 (1994); M. Harata, K. Jitsukawa, H. Masuda, H. Einaga, *ibid.*, p. 10817.
- K. A. Magnus, H. Ton-That, J. E. Carpenter, *Chem. Rev.* 94, 727 (1994).
- T. N. Sorrell, W. E. Allen, P. S. White, *Inorg. Chem.* 34, 952 (1995); W. E. Lynch, D. M. Kurtz, S. K. Wang, R. A. Scott, *J. Am. Chem. Soc.* 116, 11030 (1994); S. Mahapatra, J. A. Halfen, E. C. Wilkinson, L. Que, W. B. Tolman, *ibid.*, p. 9785.
- D. J. Spira-Solomon, M. D. Allendorf, E. I. Solomon, J. Am. Chem. Soc. 108, 5318 (1986).
- 9. A. Messerschmidt *et al.*, *J. Mol. Biol.* **224**, 179 (1992).
- M. D. Allendorf, D. J. Spira, E. I. Solomon, *Proc. Natl. Acad. Sci. U.S.A.* **82**, 3063 (1985); L. Ryden and I. Björk, *Biochemistry* **15**, 3411 (1976); I. Zaitseva *et al.*, *J. Biol. Inorg. Chem.* **1**, 15 (1996).
- 11. A. Messerschmidt, H. Luecke, R. Huber, *J. Mol. Biol.* **230**, 997 (1993).
- 12. J. L. Cole, P. A. Clark, E. I. Solomon, J. Am. Chem. Soc. **112**, 9534 (1990).
- 13. J. L. Cole, G. O. Tan, E. K. Yang, K. O. Hodgson, E. I. Solomon, *ibid.*, p. 2243.
- 14. The (1*R*,2*R*)-cyclohexanediamine backbone was chosen both for its preorganized nature and its chirality. In its energetically preferred conformation with the two amine substituents equatorially positioned, this ligand is preorganized for binding a single metal. The enantiomeric purity of the ligand significantly reduces the probability of forming diastereomeric complexes.

15. Although 1 has not been structurally characterized, its ¹H NMR spectrum in the diamine ligand region is nearly identical to that of the structurally characterized trigonal planar complex [LCu(PPh₃)](CF₃SO₃), which is formed upon addition of PPh₃ to a solution of 1. The *N*-perethylated analog of 1, [(L')Cu(CH₃CN)](CF₃SO₃) [L' = *N*,*N*,*N*',*N*'-tetraethyl-trans-(1*R*,2*R*)-cyclohex anediamine], with bound CH₃CN has also been structurally characterized as a trigonal planar species (*18*).
 16. Reported concentrations and molar absorptivites

- 16. Heported concentrations and molar absorptivites are uncorrected for the thermal contraction of CH₂Cl₂ at below-ambient temperatures, consistent with other reports.
- 17. The product of oxygenation depends on the concentration of 1. Reaction of solutions at or below 1 mM in 1 generates a different species **X** with extremely intense electronic and vibrational transitions [per Cu atom: molar absorptivity $\varepsilon = 10,000 \text{ M}^{-1} \text{ cm}^{-1}$ at 392 nm; resonance Raman features at 607 and 583 cm⁻¹ for ¹⁶O₂- and ¹⁸O₂-derived samples, respectively (CH₂Cl₂ solution, 407-nm excitation)]. The close spectroscopic resemblance of **X** to the structurally characterized dimer [(Bn₃TACN)₂Cu₂O₂]-(SbF₆)₂ recently reported [J. A. Halfen *et al.*, *Science* **271**, 1397 (1996)] suggests that it is a similar 2:1 Cu:O₂ complex (Bn₃TACN = 1,4,7-tribenzyl-1,4,7-triazacyclononane).
- 18. Supporting information is available from the author or at the Science Web site http://www.sciencemag.org/ science/feature/beyond/#cole. Included are synthetic procedures and spectroscopic characterization data for all new compounds and x-ray structural information, including tables of crystal collection data, positional and thermal parameters, and interatomic distances and angles.
- 19. Isolated yield, 60%. The x-ray crystal data is available (18).
- 20. Crystal data for [**2**]·4 CH₂Cl₂: brown rhombic blocks from cold (-40°C) CH₂Cl₂-ether; monoclinic C2 (no. 5), *a* = 28.0300(1) Å, *b* = 16.8004(3) Å, *c* = 15.3760(2) Å, β = 119.158(1)°, *V* = 6323.2(1) Å³, and *Z* = 4; 14,745 reflections were collected and appropriately averaged (18), 9779 of which were unique (150 K, 3° < 20 < 46°); 7124 reflections [] F_o]>4 σ (F_o)] yield *R* = 7.4 and R_w = 7.6.
- Although the two clusters are crystallographically unique, they are isostructural to within a rootmean-square (rms) deviation of 0.162 Å (0.093 Å

rms for the N₆Cu₃O₂ core).

- 22. The structure of **2** bears a strong superficial resemblance to that of a previously reported macrocyclic bis(μ_3 -hydroxo)tricopper(II) species; however, this thermally stable cluster exhibits full threefold symmetry, has normal Cu(II)-O and Cu(II)-N distances, and carries an overall charge of 4+ [J. Comarmond, B. Dietrich, J. Lehn, R. Louis, *Chem. Commun.* **1985**, 74 (1985)].
- K. Hesterman and R. Hoppe, Z. Anorg. Allg. Chem. 367, 249 (1969).
- 24. Formulation of 2 as a μ₃-hydroxo-μ₃-oxotricopper(II) species would also be consistent with an overall charge of 3+ but fails to rationalize the short Cu-O bonds exhibited by the unique Cu site.
- 25. D. F. Evans, J. Chem. Soc. 1959, 2003 (1959)
- P. N. Schatz, R. L. Mowery, E. R. Krausz, *Mol. Phys.* 35, 1537 (1978).
- Ferromagnetic interactions in Cu₂O₂ cores have been reported previously [for example, P. Chaudhuri et al., Angew. Chem. Int. Ed. Engl. 24, 57 (1985)].
- 28. M. J. Baldwin et al., J. Am. Chem. Soc. **114**, 10421 (1992).
- M. P. Youngblood and D. W. Margerum, *Inorg. Chem.* **19**, 3068 (1980).
- D. Chang, T. Malinski, A. Ulman, K. Kadish, *ibid*. 23, 817 (1984).
- C. LeVanda, K. Bechgaard, D. O. Cowan, M. D. Rausch, *J. Am. Chem. Soc.* 99, 2964 (1977).
- W. Shin *et al.*, *J. Am. Chem. Soc.* **118**, 3202 (1996);
 P. A. Clark and E. I. Solomon, *ibid.* **114**, 1108 (1992).
- 33. We thank the University of California Mass Spectrometry Facility (Department of Pharmaceutical Chemistry, San Francisco); Z. Hou and V. Mahadevan for experimental assistance; and F. Hollander for use of the Siemens SMART diffractometer at the University of California at Berkeley, Funding provided by NIH grants GM50730 (T.D.P.S.) and DK31450 (E.I.S.) and an NSF predoctoral fellowship (A.P.C.).

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Age and Paleogeographical Origin of Dominican Amber

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The age and depositional history of Dominican amber-bearing deposits have not been well constrained. Resinites of different ages exist in Hispaniola, but all of the main amberiferous deposits in the Dominican Republic (including those famous for yielding biological inclusions) were formed in a single sedimentary basin during the late Early Miocene through early Middle Miocene (15 to 20 million years ago), according to available biostratigraphic and paleogeographic data. There is little evidence for extensive reworking or redeposition, in either time or space. The brevity of the depositional interval (less than 5 million years) provides a temporal benchmark that can be used to calibrate rates of molecular evolution in amber taxa.

In the Dominican Republic, amber (1) occurs in commercially exploitable quantities in two zones (Fig. 1): north of Santiago de los Caballeros (the "northern area") and northeast of Santo Domingo (the "eastern area"). Amber from the northern area has been suggested to be as old as Early Eocene or as young as Early Miocene (2–7); estimates for the eastern area are more diverse, ranging from Cretaceous to Recent (2–4, 6-9). Age spreads of this magnitude are implausible, but to date no resolution of the age of Dominican amber has met with wide acceptance. The resolution offered here is based on a synthesis of available biostratigraphic and paleogeographic data from several parts of Hispaniola (Fig. 2).

In the eastern area, amber-bearing sedi-

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ments occur in the ~100-m-thick Yanigua Formation (Fm), composed of organically rich laminated sand, sandy clay, and some intercalated lignite layers up to 1.5 m thick. Plant debris is found at low frequency throughout. Isolated beds of gravel and calcarenite occur, but true alluvial sediments are absent. Amber pieces are found embedded in lignite and sandy clay. In addition to indicative sedimentary features, the character of the invertebrate and vertebrate fossils from these beds (thinshelled mollusks, foraminifera, and ostracods; crocodiles, sirenians, and turtles) imply that deposition occurred in a near-shore context, probably in coastal lagoons (8, 10) fronting low, densely forested hills (11). Microfossil assemblages (12) and zone definitions (13) indicate a late Early to early Middle Miocene age for this formation.

In the northern area, the amber-bearing unit comprises the upper 300 m of the La Toca Fm, a 1200-m-thick Oligocene to Middle Miocene suite of clastic rocks (14– 16). The amberiferous unit is composed of sandstone with occasional conglomerate that accumulated in a deltaic to deep-water environment. Individual beds—thick, coarse, and tail-graded or massive at their base—grade into amber-containing sandstone with parallel lamination, rarely presenting ripplemarks. Amber fragments from these sands show few surface signs of transport and can reach lengths of 30 to 40 cm. Lignite occurs in the form of thin lamellae within the sandstones; carbonized wood fragments are also common. These rocks grade into flyschoid, deeper water deposits containing detrital amber (17) underlain by thick conglomerate (14, 16). Microfossils (18) in the amber-bearing unit correlate with faunal zones of Early to Middle Miocene age (5, 14–16).

Paleogeographically, the eastern and northern areas were part of the same sedimentary basin that was later disrupted by movements along major faults (Fig. 1). Paleocurrent analysis (19) of amber-bearing rocks of the northern area indicates that the sediment source was located toward the southeast, so the only plausible source of resin input would have been forests surrounding the depositional basin (Fig. 1). In the eastern area, slopewash carried resinites into nearby coastal lagoons, where they were apparently concentrated in lenslike pockets. Resinites in the La Toca Fm were probably slope-washed into river channels cutting the ancestral Cordillera Central, then transported with sand and silt into the deltaic and deep-water environments of the basin. Hydrodynamic experiments (20) indicate that Hymenaea resin and copal float in fast-moving fresh water but sink when the current is slow or negligible. Fresh resin floats in saline water, but copal and amber may float or sink depending on the density of the individual specimen. Therefore, fresh resin and copal entering high-energy marine environments would probably have been widely dispersed.

Outside the major mining areas, amber occurs in small quantities in turbiditic facies of the Early to Middle Miocene Sombrerito Fm (21), south of the Cordillera Central in the area of Plateau Central–San Juán. Trace amounts have also been reported from lagoonal lignite-bearing sediments of the early Middle Miocene Maissade Fm in Haiti (22). These occurrences represent an external temporal control for the age of the amber-bearing deposits north of the Cordillera Central (Figs. 1 and 2).

In combination, these data indicate that the amber-bearing deposits of the Dominican Republic are uniformly late Early to early Middle Miocene in age (15 to 20 million years ago). However, this conclusion does not agree with efforts to date amber through the use of exomethylene resonance signatures visualized by nuclear magnetic resonance spectroscopy (NMRS) (7). In order to derive an age assess-



Fig. 1 (left). Ancestral western Greater Antilles (future Cuba and Hispaniola) in the latter half of the Early Miocene (16 to 18 million years ago). Existing coastlines (interrupted where necessary) provide orientation. SF, Septentrional fault zone; RGF, Río Grande fault zone. (Insert) Present-day Hispaniola, showing location of the main mining districts (northern and eastern areas) and distribution of the latest Eocene through Pliocene rocks (shaded). In the early Neogene, the terranes that comprise Hispaniola were located west of their current positions, closer to present-day southeastern Cuba (note alignment of Altamira and Guantánamo rocks, latest Eocene to Oligocene in age) (14,

15, *32*). Their present separation results from post-Oligocene left-lateral displacement along the SF and RGF [compare with insert and (*15*, *32*)]. The northern and eastern areas formed a protected embayment on the north coast of Hispaniola, wherein sediments could accumulate rapidly. **Fig. 2** (right). Stratigraphic columns of selected Tertiary regions in Hispaniola and eastern Cuba, compiled from various sources (*6*, *8*, *10*, *13*, *15*, *16*, *29*, *31*), with additional information on stratigraphy and age obtained for this report. Formational names are for reference; chronostratigraphic framework after (*33*).

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ment by this method, resonance intensity must first be calibrated against NMRS results for specimens of known age. The only published calibration curve (7) relevant to the dating of Dominican amber is based on two data points: (i) amber from Palo Alto mine (northern area), accepted as Early Miocene because sediments yield microfossils of that age (5), and (ii) a sample of resin from a Recent representative of Hymenaea. Age estimates based on this curve are said (7) to indicate a Late Eocene age for amber recovered from mines at La Toca and Tamboril in the northern area and a Middle Miocene age for specimens from Bayaguana and Cotuí in the eastern area. Microfossil evidence supports a Miocene age for amber-bearing sediments in the mines at Bayaguana (10), but microfossils just as clearly establish that the La Toca mines are also Miocene (18), which contradicts the NMRS results. If amberiferous sediments at La Toca, Palo Alto, and Bayaguana are paleontologically equivalent in age, then the exomethylene decay curve does not produce meaningful results, as has been pointed out by others [for example, (3)].

It has been suggested that amber occurrences in the Dominican Republic may have been emplaced by redeposition, so that they may be older than the sediments that bear them (5, 7, 8). Several considerations make this interpretation unlikely. First, amber has not been reported in rocks older than Miocene age anywhere in the Dominican Republic (Fig. 2); therefore, they cannot be the source of older amber, if such existed. Second, plants and animals preserved in Dominican amber can almost always be allocated to extant groups at low hierarchical levels and often seem to differ little from relatives living in Hispaniola today [see, for example, (2, 3, 5, 5)] 23)]. Third, if Dominican amber derives from a single species of Hymenaea (24), as is strongly indicated by infrared spectroscopy pyrolysis gas chromatography (25), then the resin source was probably highly confined, both geographically and temporally (26). Fourth, the fact that individual pieces of amber may either float or sink in saline waters could have facilitated dispersion during reworking but not the formation of concentrated ore bodies. For all these reasons, it is logical to conclude that Dominican ambers are the same age as the sediments that contain them, that is, it appears that the redeposition of ambers of different ages has not occurred.

The fact that the major occurrences of Dominican amber can be so narrowly constrained in age should open up new possibilities for investigation, such as estimating rates of genetic change from ancient DNA in amber inclusions (27). Genetic material appears to be unusually well preserved in Dominican amber (27, 28), suggesting that it may be an ideal material for this kind of study.

REFERENCES AND NOTES

- Resinites derived from plants differ widely in their chemical composition and physical characteristics (2). Copal and amber are often difficult to distinguish by inspection, but differ in their resistance to heating and organic solvents. Less resistant copal is conventionally interpreted as an unfossilized version of amber, although the relation between age and the complex diagenetic changes that yield true amber is not well understood. Dominican copal from Cotui, allegedly of Holocene age (9), is not discussed in this report because we were unable to examine its original depositional context. It is noteworthy that hard copal, sometimes with biological inclusions, can be recovered in the litter under *Hymenaea* trees today.
- D. A. Grimaldi, *Amber: Window to the Past* (Abrams and American Museum of Natural History, New York, 1996).
- _____, in Amber, Resinites and Fossil Resins, K. B. Anderson and J. C. Crelling, Eds. (ACS Symp. Ser. 617, American Chemical Society, Washington, DC, 1995), p. 203.
- 4. G. O. Poinar and D. C. Cannatella, *Science* **237**, 1215 (1987).
- C. Baroni-Urbani and J. B. Saunders, *Transactions* of the 9th Caribbean Geology Conference, Santo Domingo, Dominican Republic, W. Snow et al., Eds. (1982), vol. 1, p. 213.
- Mapa Geológico de la República Dominicana, Escala 1:250 000, Dirección de Minería, Dominican Republic, and Bundesanstalt für Geowissenschaften und Rohstoffe, Germany (1991).
- 7. J. B. Lambert, J. S. Frye, G. O. Poinar, *Archaeometry* 27, 43 (1985).
- 8. S. B. Brouwer and P. A. Brouwer, in (5), p. 305.
- D. Schlee, *Stuttg. Beitr. Naturkd.* **18**, 63 (1984); R. Burleigh and P. J. Whalley, *J. Nat. Hist.* **17**, 919 (1983).
- 10. W. Van den Bold, Bull. Am. Paleontol. 94, 1 (1988).
- 11. Amber-bearing sediments in the Dominican Republic usually contain lignites (Fig. 2) (8, 29), as do amberiferous localities elsewhere in the world (2). Cenozoic lignite deposits are not rare regionally (Figs. 1 and 2), being known from the Oligocene in Puerto Rico (30), Early to Late Miocene in Hispaniola (Fig. 2) (31), and Early to Middle Miocene in Cuba (13). Lithologically very similar to the amber-bearing Yanigua Fm in Hispaniola are the lignite-bearing Maissade Fm of central Haiti and the Arabos Fm of eastern Cuba (Figs. 1 and 2), which are composed of shales, marls, and sands, containing mixed marine and brackishwater faunas (13, 22). Their amber potential, if any, has never been tested.
- 12. The Yanigua Fm localities we investigated (Colonia San Rafael, Sierra del Agua, Bayaguana, and Yanigua) contain identical microfossil assemblages that correlate with the late Early Miocene Miogypsina-Soritiidae benthic foram zone (13) (M. antillea, Sorites marginalis, and Archaias angulatus) and other forams of Early Miocene through early Middle Miocene age (Ammonia beccarii parkinsoniana, A. b. ornata: Amphistegina sp.; Archaias angulatus; S. marginalis; Elphidium cf. E. advenum, E. cercadensis, E. lens, E. poeyanum, E. puertoricensis, E. sagra; and Quingueloculina poligona). This correspondence is in agreement with the ostracod evidence (10) from mines at Bayaguana and Laguana (Aurila galerita; Bairdia spp.; Cativella sp. aff. C. moriahensis; Cushmanidea howei; Cytherella sp. aff. C. pulchra; Eucytherella sp.; Hemicyprideis agoiadiomensis; H. cubensis (sensu stricto) and H. stephensoni; Loxoconcha runa, L. spinoalata, and L. antillea; Paracypris sp. B and Paracytheridea sp. aff. P. hispida; Paranesidea antillea; Pellucistoma sp.; Perissocytheridea alata; Procythereis? deformis; Pseudopsammocythere ex gr. vicksburgensis; and Uroleberis sp. 1).
- M. A. Iturralde-Vinent, Am. Assoc. Pet. Geol. Bull. 53, 1938 (1969).
- R. de Zoeten and P. Mann, Geol. Soc. Am. Spec. Pap. 262, 265 (1991).
- E. Calais, B. M. de Lepinay, P. Saint-Marc, J. Butterlin, A. Schaaf, *Bull. Soc. Geol. Fr.* **163**, 309 (1992).
- R. de Zoeten, thesis, University of Texas at Austin (1988).

- 17. B. Redmond, in (5), p. 199.
- 18. Microfossils are rare in the amber-bearing unit, but datable assemblages have been recovered from several samples. The rocks at the Palo Alto mine and a second locality nearby correspond to the Catapsydrax dissimilis zone, yielding C. dissimilis, C. unicavus, Globorotalia mayeri, Cassigerinella chipolensis, Globigerinoides trilobus trilobus, G. trilobus inmaturus, Globigerina venezuelana, Siphonina pulchra, Gyroidina cf. G. soldani, and Cibicides mataensis [authors' samples and (5)]. At La Toca, two samples from the flysch underlying the stratigraphic level of the amber vielded the Early Miocene fossils Globigerinoides sp. and Globigerina cf. G. angustiumbilicata and Sphaenolithus heteromorphus. A sample from just above the mine itself (14, 19) vielded an early Middle Miocene assemblage (Discoaster exilis, D. variabilis, and Helicosphaera sellii) corresponding to the Sphaenolithus belemnos zone. Samples from the overlying Villa Trina Fm yielded Middle Miocene to Pliocene microfossils (16, 19).
- 19. J. Dolan *et al.*, *Geol. Soc. Am. Spec. Pap.* **262**, 21 (1991).
- M. A. Iturralde-Vinent, unpublished experiments on fresh resin, copal, and a wide variety of Dominican ambers.
- E. García and F. Harms, Informe del Mapa Geológico de la República Dominicana escala 1:100 000: San Juán (Dirección General de Minería, Santo Domingo, Dominican Republic, 1988); F. J. Harms, Stuttg. Beitr. Naturkd. B 163, 1 (1990).
- F. Maurrasse, *Guide to Field Excursions in Haiti* (Florida Geological Society, Miami, 1982); M. W. Sanderson and T. H. Farr, *Science* **131**, 1313 (1960).
- A. G. Kluge, *Am. Mus. Novit.* **3139**, 1 (1995); R. D. E. MacPhee and D. A. Grimaldi, *Nature* **380**, 489 (1996).
- 24. J. H. Langenheim, Am. Sci. 78, 16 (1990).
- D. A. Grimaldi and A. Shedrinsky, unpublished study; pyrograms on file in Department of Entomology, American Museum of Natural History.
- 26. Prevailing winds, tropical storms, altitude, relative humidity, biological activity, and rapid burial have all been suggested as factors affecting resin production of Hymenaea and its preservation as amber (2, 8, 24). None of these explains why, in the Greater Antilles, large amounts of amber occur only in restricted Miocene deposits of Hispaniola. A possible explanation is that resins were preserved as a result of a favorable combination of factors in a unique paleogeographic scenario (Fig. 1). Rapid burial of resinite-bearing beds by >1000 m of sediments (6, 8, 19) over a period of several million years would surely have had some effect on the diagenetic processes involved in the transformation of copal into the different types of amber (including blue, red, yellow, and transparent). The surface or near-surface exposure of amber-bearing beds is a recent, perhaps Quaternary, phenomenon, because of uplift and exhumation of the Miocene rocks (19).
- R. DeSalle, J. Gatesy, W. Wheeler, D. Grimaldi, *Science* **257**, 1933 (1992); H. N. Poinar, M. Höss, J. L. Bada, S. Pääbo, *ibid.* **272**, 864 (1996).
- R. J. Cano and M. K. Borucki, *ibid.* **268**, 1060 (1995).
 Y. Champetier, M. Madre, J. C. Samama, I. Tabares, in (5), p. 277.
- 30. W. H. Monroe, U.S. Geol. Surv. Prof. Pap. 953, 1 (1980).
- 31. J. B. Saunders, P. Jung, B. Biju-Duval, *Bull. Am. Paleontol.* **89**, 1 (1986).
- 32. M. A. Iturralde-Vinent, J. Pet. Geol. 17, 243 (1994).
- W. A. Berggren, D. V. Kent, C. C. Swisher, M.-P. Aubry, Soc. Econ. Paleontol. Mineral. Spec. Publ. 54, 129 (1995).
- 34. We thank C. Díaz and G. Fernández for microfossil identifications; O. Muñiz for information on the physiology of *Hymenaea*; S. Brouwer for field assistance and much other help; and P. Mann, V. MacPhee, D. Grimaldi, C. Flemming, and two anonymous reviewers for comments on the draft version of this paper. This research was supported by grants from the RARE Center for Tropical Conservation (M.I.-V.), the Office of Grants and Fellowships of the American Museum of Natural History (M.I.-V.), and the National Science Foundation (R.D.E.M.).

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