

because it is adaptive to change 5-HT's effect on the excitability of LG only in response to a persistent change in the animal's social status. In this case, these slow changes in 5-HT's effect could function as part of the "memory" of recent agonist interactions: Too short a memory might needlessly expose the animal to the risks of aggressive interactions that occur between strangers.

We conclude that social experience can modulate neural circuit function by controlling the effect of a neuromodulator on the response of an identified neuron. Presumably this type of neural plasticity mediates the animal's social adaptation by producing experience- and context-dependent changes in the relative excitability of neural circuits. These excitability changes would translate into corresponding changes in the relative frequencies with which different behaviors are expressed. Animals differ in temperament both in their groups and individually over time; it is likely that the type of neuronal and neural circuit changes reported here underlie this sort of behavioral plasticity.

REFERENCES AND NOTES

1. E. F. Coccaro, *Int. Clin. Psychopharmacol.* **7**, 3 (1992); V. M. Linnoila and M. Virkkunen, *J. Clin. Psychiatry* **53**(suppl.), 46 (1992); S. Winberg, G. E. Nilsson, K. H. Olsen, *J. Comp. Physiol. A* **170**, 93 (1992); H. D. Steklis, M. J. Raleigh, A. Kling, K. Tachiki, *Am. J. Primatol.* **11**, 133 (1986); O. Cases *et al.*, *Science* **268**, 1763 (1995).
2. M. S. Livingstone, R. M. Harris-Warrick, E. A. Kravitz, *Science* **208**, 76 (1980).
3. E. A. Kravitz, *ibid.* **241**, 1775 (1988).
4. D. Glanzman and F. B. Krasne, *J. Neurosci.* **3**, 2263 (1983).
5. F. B. Krasne, *J. Exp. Biol.* **50**, 29 (1969); J. J. Wine and F. B. Krasne, in *The Biology of Crustacea*, D. C. Sandeman and H. L. Atwood, Eds. (Academic Press, New York, 1982), vol. 4, pp. 241–292.
6. E. T. Vu, S. C. Lee, F. B. Krasne, *J. Neurosci.* **13**, 4379 (1993).
7. D. Real and G. Czernasty, *Brain Res.* **521**, 203 (1990).
8. Juvenile (1 to 3 cm) crayfish (*Procambarus clarkii*) were hatched and raised in the laboratory, and adult (8 to 10 cm) crayfish were obtained commercially. Both were isolated for longer than 1 month. Socially dominant and subordinate animals were created by pairing previously isolated crayfish for 12 days or longer. Juvenile pairs were placed in mesh cages 5 cm in diameter in groups of eight in 20-gallon aquaria. Adult pairs were placed in individual small aquaria.
9. Crayfish of known dominance status were chilled to immobility, and the abdomen was removed and dissected dorsally to expose the ventral nerve cord. The abdomen was pinned out in a petri dish and maintained in crayfish saline [A. Van Harreveld, *Proc. Soc. Exp. Biol. Med.* **34**, 428 (1936)]. The LG axon was identified visually in the terminal connective and penetrated in the proximal portion of the axon (Fig. 1). Shocks (0.3 ms) between 3 V and 7.5 V were applied to the ipsilateral third and fourth nerves of the terminal ganglion at 90-s intervals and evoked both sub- and superthreshold LG EPSPs. The stimulus series was repeated four times. The preparation was then perfused with serotonin (100 μ M in 7 preparations at the outset of these experiments, and 50 μ M for the remaining 93 preparations) or with a 5-HT agonist (50 μ M, 67 preparations), all at 1.2 ml/min. After 45 min of perfusion, responses to four more stimulus series were recorded during continued perfusion. The effects on the LG response produced by the two concentrations of serotonin were indistinguishable. A saline wash followed; responses to a final set of stimuli were recorded after a 1-hour wash. Although the electrophysiology was performed with knowledge of the animals' dominance status, the effects of serotonin on the responses of all three types of animals were unmistakable and immediately distinguishable: The responses of isolate and dominant animals were always greater than those of controls, whereas those of subordinates were always less.
10. S.-R. Yeh, R. A. Fricke, D. H. Edwards, data not shown.
11. D. H. Edwards, W. J. Heitler, E. M. Leise, R. A. Fricke, *J. Neurosci.* **11**, 2117 (1991); R. S. Zucker, *J. Neurophysiol.* **35**, 599 (1972).
12. 5-HT increased EPSPs in the reisolated subordinates (Fig. 4, filled inverted triangles in the left panel) $57.6 \pm 16.5\%$ (mean $\pm$ SEM) above those of controls, whereas EPSPs in the reisolated dominants (Fig. 4, right panel) were increased $42.1 \pm 14.0\%$ above those of controls, and EPSPs in isolates (Fig. 4, left columns of both panels) were $63.3 \pm 8.8\%$ above those of controls.
13. 5-HT, agonist reduced EPSPs in the reisolated subordinates (Fig. 4, open triangles in left panel) $3.7 \pm 8.1\%$ below those of controls, whereas EPSPs in the reisolated dominants (Fig. 4, open triangles in right panel) were increased $3.7 \pm 7.4\%$ above those of controls, and EPSPs in the isolates (Fig. 4, open triangles in left columns of both panels) were increased $8.4 \pm 10.3\%$ above those of controls.
14. R. V. Bovbjerg, *Physiol. Zool.* **26**, 173 (1953).
15. S. R. Yeh, R. A. Fricke, D. H. Edwards, unpublished results.
16. R. A. Glennon, *J. Med. Chem.* **30**, 1 (1987); P. P. A. Humphrey, P. Haratig, D. Hoyer, *Trends Pharmacol. Sci.* **14**, 233 (1993); B. E. Leonard, *Int. Clin. Psychopharmacol.* **7**, 13 (1992).
17. B. Zhang and R. Harris-Warrick, *J. Exp. Biol.* **190**, 55 (1994).
18. We thank M. Hörner, F. B. Krasne, and E. A. Kravitz for their thoughtful comments on this paper. Supported by NIH research grant RO1 NS26457 and NSF research grant IBN 94-23486.

26 July 1995; accepted 10 November 1995

Zinc-Induced Collapse of Augmented Inhibition by GABA in a Temporal Lobe Epilepsy Model

Eberhard H. Buhl, Thomas S. Otis, Istvan Mody*

In the kindling model of temporal lobe epilepsy, several physiological indicators of inhibition by γ -aminobutyric acid (GABA) in the hippocampal dentate gyrus are consistent with an augmented, rather than a diminished, inhibition. In brain slices obtained from epileptic (kindled) rats, the excitatory drive onto inhibitory interneurons was increased and was paralleled by a reduction in the presynaptic autoinhibition of GABA release. This augmented inhibition was sensitive to zinc most likely after a molecular reorganization of GABA_A receptor subunits. Consequently, during seizures, inhibition by GABA may be diminished by the zinc released from aberrantly sprouted mossy fiber terminals of granule cells, which are found in many experimental models of epilepsy and in human temporal lobe epilepsy.

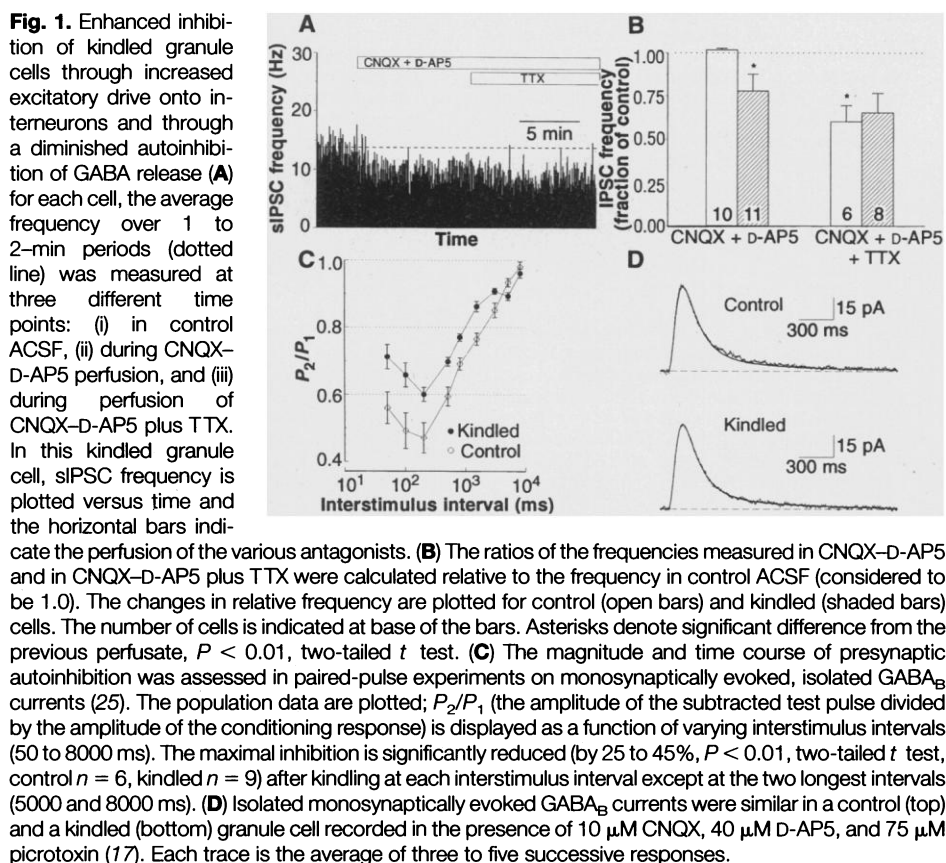
Synaptic inhibition in the mammalian forebrain is primarily mediated by GABA acting at its various receptors (1, 2). It is not known by how much the balance between excitation and inhibition has to be offset for pathological changes to occur. A reduced synaptic inhibition will favor hyperexcitability, a condition long associated with epilepsy (3). Although in vitro studies of acute epilepsies have often relied on the experimental impairment of inhibition by GABA (3), the fate of inhibition by GABA in chronic epilepsy models and particularly in human epilepsies remains unclear (3–6). A distinctive sprouting of mossy fibers in the dentate gyrus is shared among human temporal lobe epilepsy (TLE) and several experimental epilepsy models (7–10). The aberrantly sprouted mossy fibers form recurrent

excitatory synapses with other granule cells but may also contribute to the synaptic drive onto inhibitory interneurons (11). The kindling model of TLE, in which seizures are induced by initially subthreshold electrical stimuli delivered daily to limbic areas over several days or weeks, replicates many anatomical and pathological features, including the sprouting of mossy fibers as seen in human TLE (12). The model is characterized by an enhanced functional inhibition by GABA in the dentate gyrus (6, 13, 14), but there is no estimate of the excitatory drive onto the inhibitory interneurons.

We measured the degree of glutamatergic excitatory drive onto specific interneurons (15, 16) responsible for generating spontaneous inhibitory postsynaptic currents (sIPSCs) in dentate gyrus granule cells after kindling by recording inhibitory currents in the whole-cell configuration (17) at the reversal potential (0 to +5 mV) of excitatory synaptic events (18). At this membrane potential, sIPSCs could be detected selectively (Fig. 1A), whereas the excitatory drive onto the interneurons remained intact and could be assessed by the

E. H. Buhl, MRC Anatomical Neuropharmacology Unit, Oxford University, Mansfield Road, Oxford OX1 3TH, UK. T. S. Otis, Department of Neurophysiology, University of Wisconsin Medical School, Medical Sciences Center, 1300 University Avenue, Madison, WI 53706-1532, USA. I. Mody, Department of Neurology RNRC 3-131, UCLA School of Medicine, 710 Westwood Plaza, Los Angeles, CA 90095-1769, USA.

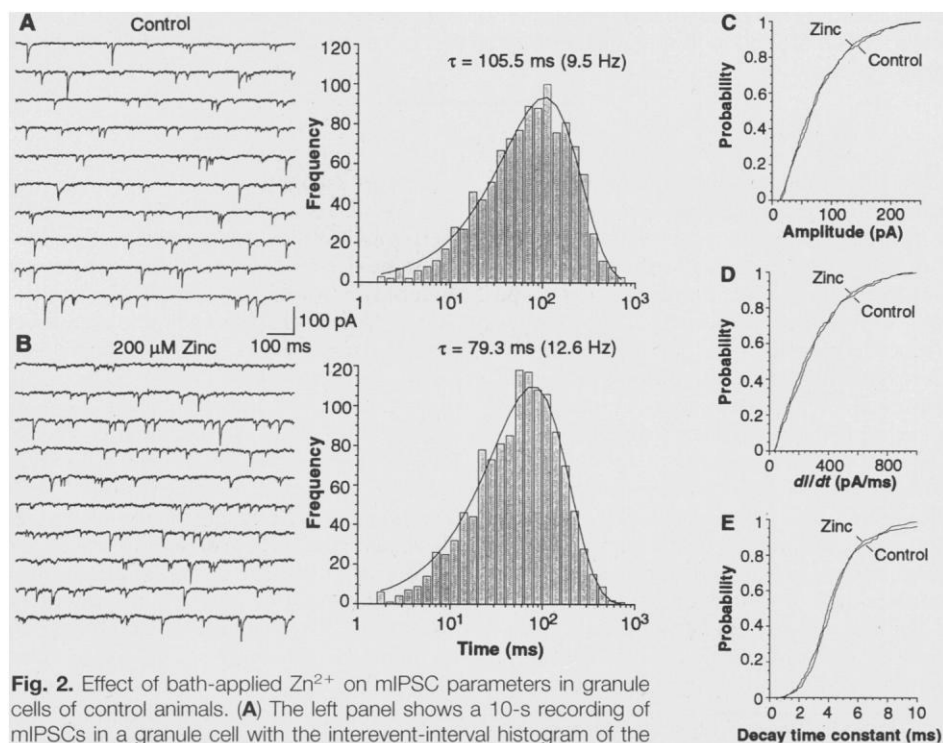
*To whom correspondence should be addressed.



decrease in event frequency caused by perfusing the ionotropic glutamate receptor antagonists 6-cyano-7-nitro-quinoxaline-2,3-dione (CNQX, 10 μ M) and D-2-amino-5-phosphonovalerate (D-AP5, 40 μ M). Such treatment causes a reduction of less than 10% in the frequency of sIPSCs (18, 19). Consistent with a weak basal spontaneous excitatory drive onto interneurons in control slices (Fig. 1B), the average ratio of sIPSC frequencies observed after and before the CNQX + D-AP5 perfusion was 1.02 ± 0.01 ($n = 10$ slices). In contrast, after kindling, perfusion of CNQX + D-AP5 caused a significantly more-pronounced reduction (0.78 ± 0.1 ; $n = 11$ slices; $P < 0.01$, Student's t test) in the frequency of inhibitory events (Fig. 1, A and B). This finding can best be explained by an enhanced excitatory drive onto the interneurons responsible for generating spontaneous events in kindled granule cells. The anatomical substrate for this enhanced excitatory drive may be the sprouted mossy fibers that invade infra- and supragranular regions known to be abundant in interneuron processes (20).

Going a step beyond the excitatory drive onto the interneurons in the epileptic dentate gyrus, we also wanted to know what fraction of the spontaneous GABA release was related to invasion of the inhibitory terminals by action potentials (2, 18, 19, 21). In control cells, the frequency of miniature IPSCs (mIPSCs) recorded after perfusion of the Na⁺ channel blocker tetrodotoxin [TTX, 1 μ M (18, 19, 21)] was $60 \pm 9\%$ ($n = 6$ slices) of that recorded in the presence of CNQX + D-AP5 alone. However, in kindled preparations, once the excitatory amino acid receptor antagonists lowered sIPSC frequency, it could not be reduced further by perfusing TTX onto the slices (Fig. 1, A and B). Hence, in the absence of an excitatory drive onto the interneurons, most of the sIPSCs in kindled granule cells seem to result from a release of GABA that is independent of action potentials.

The release of GABA from inhibitory terminals is also under the control of presynaptic GABA_B autoreceptors (22). These receptors can inhibit GABA release by as much as 40 to 60%, producing an activity-dependent disinhibition (23). An augmented presynaptic autoinhibition of GABA release, especially upon sustained firing of interneurons, could reduce dramatically the efficacy of inhibition during seizures (23). In kindled dentate gyri, we observed the opposite: The paired-pulse inhibition of monosynaptically evoked IPSCs, a paradigm used to measure activation of GABA_B autoreceptors (24, 25), was significantly reduced (Fig. 1C). Reduced autoinhibition in epilepsy may guarantee a steady release of



GABA with repeated presynaptic activity, particularly during the sustained activity accompanying seizures of GABA-transmitting terminals. Contrasted with the diminished activation of presynaptic GABA_B receptors after kindling, stimulus-evoked IPSCs mediated by GABA_B receptors (25) were comparable in control and kindled granule cells (Fig. 1D).

These physiological findings appear to be consistent with enhanced inhibition mediated by GABA_A receptors in the epileptic—that is, presumably hyperexcitable—dentate gyrus. With an augmented inhibition, how can the balance of excitability ultimately tip in favor of excitation and produce epilepsy? Excitatory neurotransmission mediated by the *N*-methyl-D-aspartate type of glutamate receptors is clearly enhanced after kindling (26), but other key factors may undermine the dampening effect of an augmented inhibitory activity.

We tackled this issue by examining another possible pathophysiological role of the massive and aberrant mossy-fiber sprouting characteristic of the epileptic dentate gyrus. Mossy fibers are loaded with Zn²⁺ that can be released on stimulation and that is estimated to reach local concentrations of a few hundred micromoles per liter (27). Sprouted to supragranular sites, mossy fiber boutons still contain Zn²⁺ (7, 10) that could be released onto perisomatic and proximal dendritic regions of granule cells where little if any Zn²⁺ is being released normally under nonepileptic conditions. These regions are precisely where spontaneous inhibitory events originate on granule cells (16). Because Zn²⁺ inhibits certain types of GABA_A receptors (28), mainly during early development (29), we considered whether it may also inhibit GABA_A receptors of granule cells. First we examined the effect of Zn²⁺ on mIPSCs in control neurons. Perfusion of 200 μM ZnCl₂ had no effect on the amplitude, decay-time constant, or rates of rise of mIPSCs (Fig. 2, *n* = 10). The frequency of mIPSCs was moderately increased by Zn²⁺ (12 ± 5%; *n* = 10; *P* = 0.038, two-tailed *t* test), an effect that lasted beyond the wash, persisting in some cells for as long as 40 min after the return to the control perfusate. These findings reflect known presynaptic effects of Zn²⁺ (30) and are consistent with the presence of Zn²⁺-insensitive GABA_A receptors on control granule cells.

If GABA_A receptors were similar in control and kindled granule cells, then contrary to our original hypothesis, Zn²⁺ release from sprouted mossy fibers could not cause a diminished inhibition in the epileptic dentate gyrus. The hypothesis could still be valid if marked differences were present in the sensitivity of control and “epileptic” GABA_A receptors to Zn²⁺. Trying to un-

ravel possible differences in the Zn²⁺ sensitivity of control and epileptic GABA_A receptors, we examined the effect of Zn²⁺ on mIPSCs recorded in slices obtained from kindled animals. In contrast to its effect in controls, Zn²⁺ produced a striking effect on mIPSCs recorded in kindled granule cells. Perfusion of ZnCl₂ (200 μM) blocked mIPSCs in kindled neurons, resulting in a significant decrease in mIPSC frequency (56 ± 8%; *n* = 18; Fig. 3, A and B). The presynaptic effect of Zn²⁺, commonly seen as a lasting increase in the frequency of mIPSCs in control neurons, was masked in kindled cells and could only be observed after the wash of ZnCl₂ (Fig. 3C). Thus, the reduction of mIPSC frequency by perfusion of ZnCl₂ must have resulted solely from a reversible antagonism of epileptic granule-cell GABA_A receptors by Zn²⁺ (Fig. 3, D through H). This antagonism was charac-

terized by significantly (*P* < 0.01, paired two-tailed *t* test) reduced median amplitudes (by 33 ± 4%, from 57.9 ± 3.3 to 38.3 ± 3.3 pA; *n* = 18; Fig. 3, D and G), rates of rise (by 35 ± 5%, from 261 ± 16 to 168 ± 15 pA/ms; *n* = 18; Fig. 3, E and H), and faster decay-time constants (by 20 ± 5%, from 3.48 ± 0.08 to 2.82 ± 0.22 ms; *n* = 18; Fig. 3, F and H) of the mIPSCs recorded in the presence of Zn²⁺ in kindled neurons. We also simulated the effects of Zn²⁺ on mIPSCs (31). These simulations reflected the possibility that Zn²⁺ may block the epileptic GABA_A receptor channels (32) through a noncompetitive mechanism (33).

The most obvious explanation for the Zn²⁺ sensitivity of kindled synaptic GABA_A receptors would be the possible kindling-induced loss of γ subunits. However, because of the increased benzodiazepine binding after kindling (34), this pos-

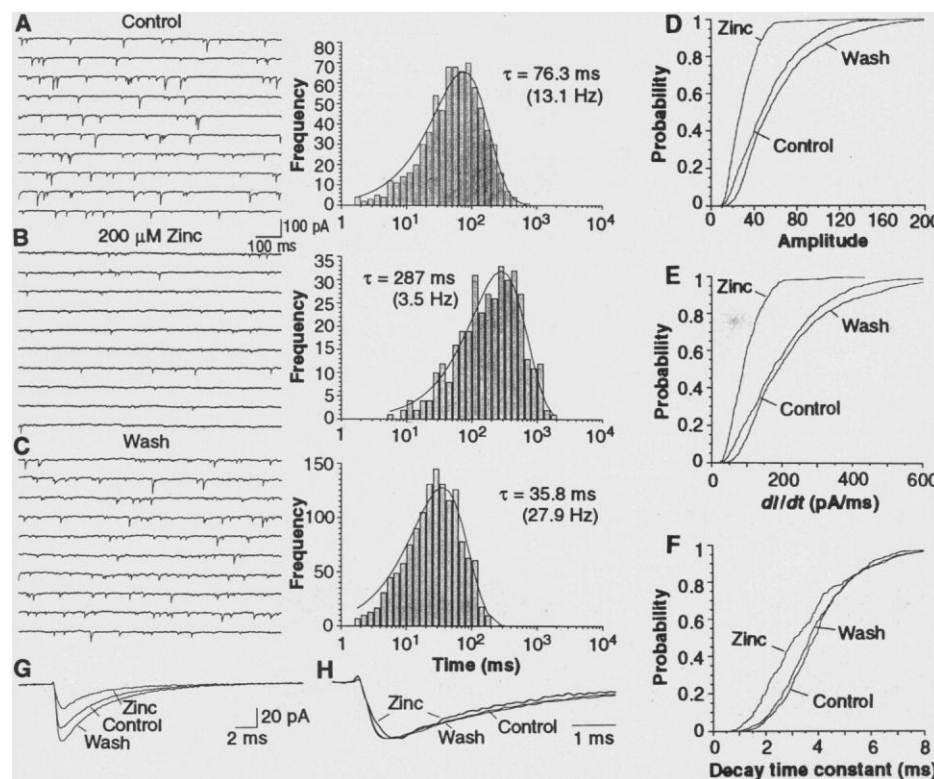


Fig. 3. Block of mIPSCs by bath-applied Zn²⁺ in a kindled granule cell. **(A)** The mean frequency of mIPSCs in control ACSF was 13.1 Hz. **(B)** Perfusion of 200 μM ZnCl₂ resulted in a dramatic reduction (to 27% of control, or 3.5 Hz) of mIPSC frequency. **(C)** The effect of Zn²⁺ was reversible. Moreover, indicating a long-term presynaptic effect comparable to that observed in controls, the frequency of mIPSCs more than doubled after washout and remained elevated for the remainder of the experiment (>30 min). The leftmost panels in **(A)** through **(C)** show 10-s consecutive recordings of mIPSCs in the presence of ionotropic glutamate antagonists. The corresponding panels to the right show the respective interevent-interval histograms fitted with exponential distributions. Cumulative probability plots of mIPSC parameters taken from the same cell illustrated in **(A)** through **(C)** indicated that Zn²⁺ resulted in significant reduction of mIPSC amplitudes **(D)**, rates of rise **(E)**, and decay-time constants **(F)**, indicating a postsynaptic action on GABA_A receptors in the granule cell. The effects on mIPSC parameters were readily reversible after superfusion with control ACSF. **(G)** After events had been sorted according to their rates of rise, those (±15%) scattered around the median were averaged and superimposed. Note the Zn²⁺-induced reversible reduction of the average mIPSC. **(H)** The superimposition of normalized averages on a different time scale shows that Zn²⁺ perfusion reversibly reduced the mIPSC rate of rise and increased the rate of decay, consistent with an action of Zn²⁺ at synaptic GABA_A channels.

sibility seems unlikely. Nevertheless, we tested the sensitivity of mIPSCs recorded in kindled preparations to the benzodiazepine agonist zolpidem. As in controls (18, 31), perfusion of zolpidem (10 μ M) produced a significant lengthening ($327 \pm 43\%$) of mIPSC decay-time constants in kindled granule cells ($n = 6$), consistent with the presence of functional benzodiazepine receptors after kindling. The absence of γ subunits, which are critical for benzodiazepine sensitivity (28), is commonly associated with the blocking effect exerted by Zn^{2+} on GABA_A receptors during early ontogeny (29). Therefore, the preserved benzodiazepine sensitivity after kindling must reflect the continued presence of γ subunits in epileptic GABA_A receptor channels (35); the Zn^{2+} sensitivity of these receptors must have arisen despite the functional γ subunits. Other subunits may also regulate the Zn^{2+} sensitivity of GABA_A receptors: Some benzodiazepine-sensitive receptors are inhibited by Zn^{2+} (33), possibly through certain α subunits or the δ subunit (36).

In summary, two additional components of inhibition seem to compensate for hyperexcitability in the epileptic dentate gyrus: (i) an increased excitatory drive onto inhibitory interneurons, and (ii) a decreased autoinhibition of GABA release. Yet this enhanced inhibition by GABA ultimately collapses in the kindled hippocampus, and Zn^{2+} may be pivotal in this breakdown. During massive neuronal activity, Zn^{2+} released from sprouted mossy fibers around granule cell bodies and proximal dendrites (37) could cause a significant impairment in the function of epileptic GABA_A receptors that are sensitive to Zn^{2+} . This Zn^{2+} -induced reduction of inhibition by GABA may promote the spread of epileptic activity. Furthermore, if Zn^{2+} -sensitive GABA_A receptors were present in the normal adult brain, the activity-dependent Zn^{2+} release from neighboring excitatory boutons (27) may explain the intense predisposition of certain brain structures to epilepsy (3).

REFERENCES AND NOTES

- R. L. Macdonald and R. W. Olsen, *Annu. Rev. Neurosci.* **17**, 569 (1994).
- I. Mody, Y. De Koninck, T. S. Otis, I. Soltesz, *Trends Neurosci.* **17**, 517 (1994).
- P. A. Schwartzkroin, Ed., *Epilepsy: Models, Mechanisms, and Concepts* (Cambridge Univ. Press, Cambridge, 1993).
- R. S. Sloviter, *Science* **235**, 73 (1987); M. Isokawa, G. Avanzini, D. M. Finch, T. L. Babb, M. F. Levesque, *Epilepsy Res.* **9**, 242 (1991); M. Avoli, *ibid.* **10**, 33 (1991); R. S. Sloviter, *Hippocampus* **4**, 250 (1994); K. Uruno, M. J. O'Connor, L. M. Masukawa, *ibid.*, p. 583.
- R. S. Sloviter, *Hippocampus* **1**, 41 (1991); J. W. Bekenstein and E. W. Lothman, *Science* **259**, 97 (1993).
- S. Otis, Y. De Koninck, I. Mody, *Proc. Natl. Acad. Sci. U.S.A.* **91**, 7698 (1994).
- T. Sutula, H. Xiao-Xian, J. Cavazos, G. Scott, *Science* **239**, 1147 (1988).
- J. Cronin and F. E. Dudek, *Brain Res.* **474**, 181 (1988); J. E. Cavazos, G. Golarai, T. P. Sutula, *J. Neurosci.* **11**, 2795 (1991); M. Isokawa, M. F. Levesque, T. L. Babb, J. J. Engel, *ibid.* **13**, 1511 (1993).
- T. P. Sutula, G. Cascino, J. Cavazos, I. Parada, L. Ramirez, *Ann. Neurol.* **26**, 321 (1989).
- T. L. Babb, W. R. Kupfer, J. K. Pretorius, P. H. Crandall, M. F. Levesque, *Neuroscience* **42**, 351 (1991).
- C. E. Ribak and G. M. Peterson, *Hippocampus* **1**, 355 (1991); R. S. Sloviter, *Neurosci. Lett.* **137**, 91 (1992); M. M. Okazaki, D. A. Evenson, J. V. Nadler, *J. Comp. Neurol.* **352**, 515 (1995).
- T. P. Sutula, *Epilepsy Res.* **10**, 62 (1991); J. O. McNamara, *J. Neurosci.* **14**, 3413 (1994). Three of the 24 kindled animals in this study were prepared for Timm's stain. The pattern of supragranular mossy fibers in all of the kindled animals examined was consistent with the level of sprouting described by Sutula *et al.* (7).
- M. N. G. Titulaer, W. E. Ghijsen, W. Kamphuis, T. C. De Rijk, F. H. Lopes da Silva, *J. Neurochem.* **64**, 2615 (1995).
- L. P. Tuff, R. J. Racine, R. Adamec, *Brain Res.* **277**, 79 (1983); M. W. Oliver and J. J. Miller, *Exp. Brain Res.* **57**, 443 (1985); W. Kamphuis, J. A. Gorter, W. J. Wadman, F. H. Lopes da Silva, *Neurosci. Lett.* **141**, 101 (1992).
- E. H. Buhl, K. Halasy, P. Somogyi, *Nature* **368**, 823 (1994).
- I. Soltesz, D. K. Smetters, I. Mody, *Neuron* **14**, 1273 (1995).
- The procedure for surgery and daily kindling of male Wistar rats through the hippocampal commissure was described previously in detail [G. Köhr, Y. De Koninck, I. Mody, *J. Neurosci.* **13**, 3612 (1993)]. Only cells that satisfied our criteria throughout the experiment for a constant low access resistance recording were included in the analysis. A total of 26 control cells from 2 age-matched and 11 sham-implanted controls, and 44 granule cells from 21 kindled animals with at least 15 stage five motor seizures were included. Coronal or horizontal 400- μ m-thick brain slices were prepared from control and kindled rats (24 to 48 hours after the last seizure) as described previously (20). The artificial cerebrospinal fluid (ACSF) contained (in mM) 126 NaCl, 5 KCl, 2 CaCl₂, 2 MgCl₂, 26 NaHCO₃, 10 D-glucose, 0.01 CNQX (Tocris Cookson, Bristol, UK), 0.04 D-AP5 (Tocris Cookson, Bristol, UK), and 0.001 TTX (Calbiochem). The ACSF and the recording chamber were continuously bubbled with a mixture of 95% O₂ and 5% CO₂. Miniature IPSCs were recorded from dentate gyrus granule cells with tight-seal, whole-cell patch clamp pipettes containing 140 mM CsCl, 2 mM MgCl₂, 10 mM HEPES, and occasionally 0.2 mM EGTA or bis(aminophenoxy) ethanetetraacetic acid (BAPTA); CsOH was used to adjust the pH to 7.2. For experiments measuring isolated synaptic GABA_B responses, the pipettes contained (in mM) 135 K-gluconate, 10 HEPES, 2 MgCl₂, and 0.2 tris-guanosine triphosphate (GTP); osmolarity of the filling solutions was between 265 and 275 mosm. All recordings were obtained at $35 \pm 0.5^\circ\text{C}$ and holding potentials of -70 ± 3 mV with either an Axopatch 1D, an Axopatch 200A, or a List EPC-7 amplifier. Access resistances (compensated 60 to 80%) were frequently monitored and were constant during the perfusion of ZnCl₂ and comparable between recordings in control and kindled granule cells. The signals were transmitted through a low-pass filter at 3 kHz (-3 dB, eight-pole Bessel filter, Frequency Devices 9002) and digitized at 20 kHz (DT 2821 A/D interface). Individual events were detected and analyzed off-line [CDR software; J. Dempster, University of Strathclyde, Glasgow, UK; see also (16) and (18)]. All experimental values are given as means $\pm$ SEMs. Significance was set at the 0.01 level.
- T. S. Otis and I. Mody, *Neuroscience* **49**, 13 (1992).
- S. M. Thompson, *Prog. Neurobiol.* **42**, 575 (1994).
- Z. S. Han, E. H. Buhl, Z. Lőrinczi, P. Somogyi, *Eur. J. Neurosci.* **5**, 395 (1993).
- F. A. Edwards, A. Konnerth, B. Sakmann, *J. Physiol. London* **430**, 213 (1990); T. S. Otis, K. J. Staley, I. Mody, *Brain Res.* **545**, 142 (1991). The release of GABA from inhibitory terminals in the CNS persists in the absence of any excitatory drive onto the interneurons, any action potentials, or any Ca²⁺ entry into the terminals [see also (2), (18), and (19)]. This type of GABA release is responsible for generating IPSCs. Their frequency varies from brain region to brain region but it is consistently less than that observed in the presence of action potentials.
- N. G. Bowery *et al.*, *Nature* **283**, 92 (1980).
- C. H. Davies, S. J. Starkey, M. F. Pozza, G. L. Collingridge, *Nature* **349**, 609 (1991); D. D. Mott and D. V. Lewis, *Science* **252**, 1718 (1991); D. D. Mott, C. W. Xie, W. A. Wilson, H. S. Swartzwelder, D. V. Lewis, *J. Neurophysiol.* **69**, 674 (1993); S. M. Thompson, M. Capogna, M. Scanziani, *Trends Neurosci.* **16**, 222 (1993).
- C. H. Davies, S. N. Davies, G. L. Collingridge, *J. Physiol. London* **424**, 513 (1990); N. A. Lambert and W. A. Wilson, *J. Neurophysiol.* **72**, 121 (1994).
- T. S. Otis, Y. De Koninck, I. Mody, *J. Physiol. London* **463**, 391 (1993). The monosynaptic-evoked GABA_B currents were fit with an equation of the form $I(t) = A[1 - \exp(-t/\tau_{n1})]^4 [Aw_{n1}\exp(-t/\tau_{n1}) + Aw_{n2}\exp(-t/\tau_{n2})]$ where $I(t)$ is current as a function of time, w_{n1} and w_{n2} are weighting factors (where $A = Aw_{n1} + Aw_{n2}$), τ_{n1} is the time constant of the activation parameter, and τ_{n1} and τ_{n2} are the time constants of the inactivation parameters. The average values ($\pm$ SEM) for τ_{n1} , τ_{n1} , τ_{n2} , w_{n1} , and w_{n2} of the GABA_B responses were comparable in milliseconds except w_{n1} and w_{n2} : 41.8 ± 2.7 , 152.7 ± 10.8 , 981 ± 9.8 , 0.87 ± 0.02 , and 0.11 ± 0.02 (control, $n = 6$), and 45.6 ± 1.2 , 143.4 ± 7.6 , 722 ± 86.9 , 0.83 ± 0.06 , and 0.17 ± 0.06 (kindled, $n = 9$).
- I. Mody and U. Heinemann, *Nature* **326**, 701 (1987); J. E. Kraus, G. C. Yeh, D. W. Bonhaus, J. V. Nadler, J. O. McNamara, *J. Neurosci.* **14**, 4196 (1994).
- S. Y. Assaf and S. H. Chung, *Nature* **308**, 734 (1984); G. A. Howell, M. G. Welch, C. J. Frederickson, *Nature* **308**, 736 (1984); C. J. Frederickson and D. W. Moncrieff, *Biol. Signals* **3**, 127 (1994).
- A. Draguhn, T. A. Verdorn, M. Ewert, P. H. Seeburg, B. Sakmann, *Neuron* **5**, 781 (1990); T. G. Smart, S. J. Moss, X. Xie, R. L. Huganir, *Br. J. Pharmacol.* **103**, 1837 (1991).
- G. L. Westbrook and M. L. Mayer, *Nature* **328**, 640 (1987); T. G. Smart and A. Constanti, *Br. J. Pharmacol.* **99**, 643 (1990); N. L. Harrison and S. J. Gibbons, *Neuropharmacology* **33**, 935 (1994); T. G. Smart, X. Xie, B. J. Krishek, *Prog. Neurobiol.* **42**, 393 (1994).
- Y. X. Wang and D. M. Quastel, *Pflügers Arch.* **415**, 582 (1990).
- Y. De Koninck and I. Mody, *J. Neurophysiol.* **71**, 1318 (1994); I. Soltesz and I. Mody, *ibid.* **73**, 1763 (1995).
- According to the model, to produce the observed effects of Zn^{2+} on kindled mIPSC kinetics without a concomitant reduction in single-channel conductance (33), the probability of opening (p_{open}) during bursts had to be decreased by 25 to 30%, an effect also observed in steady state single-channel recordings (33). Additionally, the unblocking rate had to be lowered by 71% (from 6666 to 1961 s⁻¹) and the closing rate of the channels had to be enhanced by 80% (from 303 to 543 s⁻¹). The ON rate, a parameter contributing to the rate of rise of mIPSCs (38), had to be reduced by 61% (from 7662 to 3000 s⁻¹).
- P. Legendre and G. L. Westbrook, *Mol. Pharmacol.* **39**, 267 (1991); T. G. Smart, *J. Physiol. London* **447**, 587 (1992).
- F. Valdes, R. M. Dasheiff, F. Birmingham, K. A. Crutcher, J. O. McNamara, *Proc. Natl. Acad. Sci. U.S.A.* **79**, 193 (1982).
- M. Clark, G. S. Massenburg, S. R. B. Weiss, R. M. Post, *Mol. Brain Res.* **26**, 309 (1994); W. Kamphuis, T. C. De Rijk, F. H. Lopes da Silva, *Neurosci. Lett.* **174**, 5 (1994); M. Kokaia *et al.*, *Mol. Brain Res.* **23**, 323 (1994).
- N. C. Saxena and R. L. Macdonald, *J. Neurosci.* **14**, 7077 (1994); G. White and D. A. Gurley, *Neuroreport* **6**, 461 (1995).
- In a few preliminary experiments, we tetanically (10 Hz/10 s) stimulated the mossy fibers in the CA3

region of kindled slices in the presence of blockers of excitatory amino acid receptors with the aim to release endogenous Zn^{2+} onto the granule cells. In one of three experiments, we successfully reproduced the effects of perfused Zn^{2+} on sIPSCs, presumably through the release of Zn^{2+} from sprouted mossy fibers. Repetitive stimuli delivered to the same location in control slices had no effect on sIPSCs ($n = 6$). In slices, Zn^{2+} release experiments are difficult

to control because even low-frequency stimuli used to test evoked responses can inadvertently release the bulk of Zn^{2+} from the mossy fibers. In the absence of any exogenous Zn^{2+} added to the ACSF, the lost Zn^{2+} cannot be replenished (C. J. Frederickson, personal communication).

38. D. J. Maconochie, J. M. Zempel, J. H. Steinbach, *Neuron* **12**, 61 (1994).
39. We thank R. W. Olsen and P. Somogyi for critical

comments on the manuscript, C. J. Frederickson for technical advice, J. Gruneich for technical assistance, I. Parada for the Timm's staining, and S. Vietla for carrying out some of the zolpidem experiments. This research was supported by National Institute of Neurological Disorders and Stroke grants NS 12151 and NS 30549 (I.M.).

28 September 1995; accepted 30 November 1995

TECHNICAL COMMENTS

Analog Computational Power

Response: Peter Shor (1) and Richard Y. Kain (2) recently commented on my report "Computation Beyond the Turing Limit" (3). Shor questions the nature of the advice used in analog computation, or equivalently, the real weights in the neural networks model. He suggests that they must either be programmed, or be random, or be physical constants, and notes problems in all these cases.

First, by definition, the constants are not necessarily programmable, because they are general, not only computable, real numbers.

Second, the constants are inherently different from random numbers. Rather, they compose the real characteristics of a system. To exemplify this, consider the logistic map

$$x_{n+1} = ax_n(1 - x_n)$$

(4) (the state of the system at time n is represented by x_n). Here a minute change in the constant a may result in a qualitative change in the motion, such as period doubling or a transition from periodic to chaotic motion. Independent of this, let me illuminate random processes. Computer scientists often model a random process as a coin that has a success probability (to fall on "heads") of $1/2$. Probabilistic Turing machines that use a coin with (exactly) $1/2$ success probability compute the class BPP (which, as Shor says, is believed to be no stronger than what deterministic Turing machines compute efficiently). However, I have shown (5) that if the success probability of the coin is a real number, the resulting class is again super-Turing! (The

networks in this case compute the class BPP/log, which is included in P/poly.) The exact probability of the coin is not known to the Turing machine (nor the underlying process) that utilizes it and can only be approximated by a chain of flips; yet, it still adds power to the classical model.

Third, the weights in neural network models can be thought of as modeling the physical characteristics of a specific system. Shor's comment that the current measurements of physical constants are poor is crucial if one wants to build a general analog computer directly from the description; the design of such a computer is an open problem. However, this problem is immaterial for the mathematical modeling of an analog computation of nature.

In a natural analog computation process, one starts from initial conditions that constitute the (finitely describable) input, and the system evolves according to the specific equations of motion to its final position, which constitutes the output. The evolution is controlled by the exact equations of motion with the exact physical constants. The analog physical system "solves" the equations of motion exactly. For example, planetary motion is used to measure time with very high precision although we know the gravitational constant G only to two digits. The planets, of course, revolve according to the exact value of G , irrespective of its measurement by humans.

Although the networks are defined with unbounded precision, up to the q th step of the computation, only the first $O(q)$ bits in

both weights and activation values of the neurons (and the first $\log q$ bits that describe the stochastic process) influence the result (6). This property of neural networks is identical to that of chaotic systems, suggesting that neural networks are indeed natural models of analog physical dynamics.

In his comment, Kain does not mention the importance of constraints in computation as established by Karp (7) and others. The imposition of constraints is one of the main developments that revolutionized the classical theory of computation from discrete mathematics into the modern complexity theory of realistic machines. Readers interested in pursuing some of the issues raised by Kain (for example, the difference between oracle and advice machines, as well as complexity) are referred to (8) for their precise description. To summarize, both under resource constraints (complexity) and in their absence (computability), my model exceeds the Turing power, and thus may be referred to as a "super-Turing" one.

Hava T. Siegelmann

*Technion Israel Institute of Technology,
Technion City, Haifa 32000, Israel*

REFERENCES

1. P. Shor, *Science* **271**, 92 (1996).
2. R. Y. Kain, *ibid.*, p. 92.
3. H. T. Siegelmann, *ibid.* **268**, 545 (1995).
4. M. J. Feigenbaum, *J. Stat. Phys.* **19**, 25 (1978).
5. H. T. Siegelmann, *Lecture Notes in Computer Science* **820**, 23 (1994).
6. H. T. Siegelmann and E. D. Sontag, *Theor. Comput. Sci.* **131**, 331 (1994).
7. R. M. Karp, in *Complexity of Computation Computations*, J. W. Miller and R. E. Thatcher, Eds. (Plenum, New York, 1972), pp. 85–104.
8. J. L. Balcazar, J. Diaz, J. Gabarro, *Structural Complexity*, vols. 11 and 12, of *EATCS Monogr.* (Springer-Verlag, New York, 1988 and 1990).

10 July 1995; revised 25 October 1995; revised 22 November 1995; accepted 27 November 1995