

# Kin Selection, Social Structure, Gene Flow, and the Evolution of Chimpanzees

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Hypotheses about chimpanzee social behavior, phylogeography, and evolution were evaluated by noninvasive genotyping of free-ranging individuals from 20 African sites. Degrees of relatedness among individuals in one community were inferred from allele-sharing at eight nuclear simple sequence repeat (SSR) loci. Males are related on the order of half-siblings, and homozygosity is significantly increased at several SSR loci compared to Hardy-Weinberg expectations. These data support the kin-selection hypothesis for the evolution of cooperation among males. Sequence variation patterns at two mitochondrial loci indicate historically high long-distance gene flow and clarify the relationships among three allopatric subspecies. The unexpectedly large genetic distance between the western subspecies, *Pan troglodytes verus*, and the other two subspecies suggests a divergence time of about 1.58 million years. This result, if confirmed at nuclear loci and supported by eco-behavioral data, implies that *P. t. verus* should be elevated to full species rank.

Almost nothing is known about genetic variation of the chimpanzee, *Pan troglodytes*, in nature. Hypotheses concerning chimpanzee sociobiology and evolution have gone untested for many years because of difficulties surrounding tissue acquisition for genetic analysis. We here demonstrate a method of noninvasive genotyping based on DNA amplified from hair shed in night nests or plucked from captive apes (1). Using eight hypervariable simple sequence repeat (SSR) nuclear loci and two mitochondrial (mtDNA) sequences, we demonstrate how multilocus data can be used to provide critical information concerning kin selection, mating structure, individual reproductive success, inclusive fitness, population structure, gene flow, phylogeography, and phylogenetic relationships. This technical advance also permits a reexamination of the relevance of chimpanzee behavioral ecology (2, 3) to the elucidation of the evolution of human behavioral and genetic patterns. Although this report focuses on one species of primate, these methods are immediately applicable to many oth-

ers of concern to ecologists and evolutionary and conservation biologists.

Chimpanzees are geographically widespread and eco-behaviorally diverse. Despite up to 34 years of field study at a few sites, little is known about the phylogeny of the chimpanzee subspecies, their times of divergence, historical population sizes, or patterns of gene flow. Phylogenetic reconstructions are hampered by a complete lack of a fossil record, and data on long-distance dispersal and genetic variation. This study of genetic variation of chimpanzee populations across Africa describes phylogeographic patterns, begins to test hypotheses for the evolution of chimpanzees' social structure, and sets the foundation for comparative studies among communities within and between subspecies.

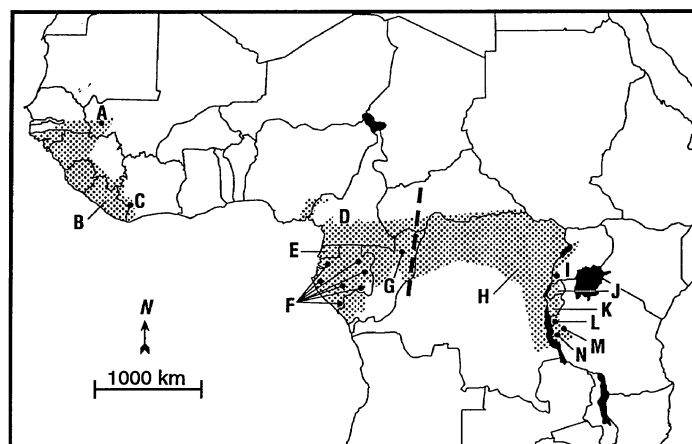
The Kasakela community in the Gombe National Park, Tanzania (Fig. 1, site L), is the longest studied wild chimpanzee population (2). At Gombe, we sampled all ( $N = 43$ )

individuals in the Kasakela social community (1, 4). Adult chimpanzees make nests (in trees) of leaves and branches, and, with the exception of mother-infant pairs, typically sleep individually in a fresh nest each night. Hairs from Gombe were collected by following known individuals, watching them build nests, and then returning at dawn to search the abandoned nest. Genetic characterization of this community is the first step in understanding the diversity and similarities among chimpanzee communities, populations, and subspecies. Our study is necessarily limited by the logistical difficulties of intensive sample collection across Africa, but ongoing and planned studies at other sites will elucidate the constants and variables in chimpanzee population genetic structure.

DNA was amplified by the polymerase chain reaction (PCR) from hair samples collected from animals at 20 sites across Africa (Fig. 1). In all, we genotyped 67 individuals at eight variable nuclear SSR loci (8) and determined DNA sequence variation for many individuals at two informative mtDNA loci (5). The highly polymorphic di- and tri- and tetra-nucleotide SSR loci differ in the number of times the core sequence is repeated (typically 10 to 30 times) (5–7) and such simple sequence length polymorphism (SSLP) was scored on autoradiographs of polyacrylamide gels (8). Individual multilocus genotypes were used to establish pedigree relationships and characterize a population's relative genetic variability and behaviorally important substructuring. The mtDNA sequences were amplified and double-stranded products were directly sequenced to detect phylogenetically informative genetic variation. A 178-bp (base pair) segment of the cytochrome b (cyt b) region of 34 chimpanzees and one human was used to examine the deeper branches of chimpanzee phylogeny, and a 345-bp segment of the more variable control region of 66 chimpanzees, two bonobos (*Pan paniscus*), and one human was used to characterize phyloge-

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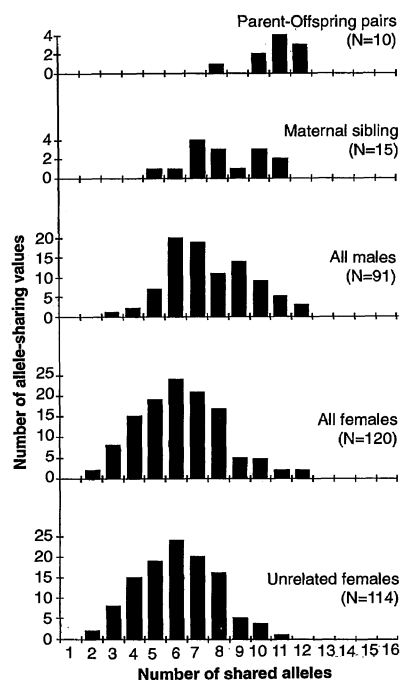
**Fig. 1.** Map of equatorial Africa, with sample collection sites or countries indicated by letters A to N, and approximate modern distributions of chimpanzees indicated by shading (2, 12). The approximate position of the poorly defined subspecies boundaries in central Africa is indicated by the dashed line.

graphic patterns, population genetic structure, and historical gene flow patterns both locally and regionally (9).

We present and discuss our results in a hierarchical fashion beginning with individuals and proceeding to populations, subspecies, and species. We have previously demonstrated the utility of hypervariable SSR variation for unambiguous paternity exclusion and the establishment of formal pedigree relationships among captive chimpanzees (5). In the free-ranging Kasakela community, with 19 matriline in 1991, we have established probable paternity in the cases of *Faustino* and *Sherehe* (whose mothers conceived during episodes of promiscuous mating with nine or ten local males) (8). This noninvasive multilocus SSR genotyping method provides results that are genetically interpretable, replicable, and comparable across gels, samples, laboratories, and closely related taxa. With the addition of more loci and individuals sampled (such as tissues of deceased individuals), it should be possible to further reconstruct the Gombe pedigree and test hypotheses about chimpanzee behavior on the basis of more than 30 years of field observations.

### Social Structure and Kin Selection

Hypotheses that kin selection has influenced the evolution of chimpanzee social



**Fig. 2.** Allele sharing distributions for all eight scored loci in subgroups of the Gombe Kasakela community. *N*, the number of pairwise allele-sharing values. The "unrelated female" group excludes maternal siblings and parent-offspring pairs. This analysis was used for the data on individuals of the Gombe community because information on kinship and gender were available only for this population.

structure predict that degrees of relatedness among males, who are philopatric, and females, who usually disperse at adolescence, should be significantly different (10). To test this, we compared the distributions of allele sharing at all scored loci between pairs of males and pairs of females from the Gombe community. The mean number of alleles per locus ( $\pm$  standard error) shared between males ( $0.854 \pm 0.203$ ) is significantly higher (*t* test,  $P < 0.05$ ) than that between females ( $0.696 \pm 0.240$ ), indicat-

ing that males are indeed more related to one another than are females. To examine the degree of relatedness indicated by allele-sharing values (summed over all eight loci; Fig. 2) within each sex, we first performed a permutation test for resampling of allele-sharing values among males, females, known parent-offspring pairs, maternal sibs (assumed to be half-sibs), and maternally unrelated females. The results suggest that males are related at the level of half-sibs because the allele-sharing values fell below

**Table 1.** Allele frequencies (%) and standard errors for eight loci in the Gombe and non-Gombe sample sets. Allele frequencies were calculated for all individuals in the Gombe community (Total), as well as for all presumed unrelated individuals (Unrelated); (n), the number of chromosomes sampled. Two individuals (Mitumba 1 and 2, from the adjacent community at Gombe) were not included in the calculations because of their unknown genetic relationship to the Gombe Kasakela community. Since all loci are codominant, the allele frequencies were computed by the gene count method (45), and their standard errors were calculated as the square root of the variance of the corresponding binomial proportions (46).

Allele	Population			Allele	Population		
	Gombe		Non-Gombe		Gombe		Non-Gombe
	Unrelated	Total			Unrelated	Total	
<i>Mfd-3</i>				<i>FABP</i>			
1			3.8 $\pm$ 2.7	1			3.7 $\pm$ 2.6
3			1.9 $\pm$ 1.9	3	22.5 $\pm$ 6.6	13.9 $\pm$ 4.1	14.8 $\pm$ 4.8
4			13.5 $\pm$ 4.7	4	50.0 $\pm$ 7.9	50.0 $\pm$ 5.9	33.3 $\pm$ 6.4
5	60.0 $\pm$ 7.7	54.2 $\pm$ 5.9	28.8 $\pm$ 6.3	5	7.5 $\pm$ 4.2	5.6 $\pm$ 2.7	16.7 $\pm$ 5.1
6	10.0 $\pm$ 4.7	6.9 $\pm$ 3.0	23.1 $\pm$ 5.8	6	7.5 $\pm$ 4.2	6.9 $\pm$ 3.0	20.4 $\pm$ 5.5
7			7.7 $\pm$ 3.7	7	12.5 $\pm$ 5.2	23.7 $\pm$ 5.0	11.1 $\pm$ 4.3
8		1.4 $\pm$ 1.4	3.8 $\pm$ 2.7	(n)	40	72	54
9	25.0 $\pm$ 6.8	34.7 $\pm$ 5.6	7.7 $\pm$ 3.7	<i>Mfd-32</i>			
10	5.0 $\pm$ 3.4	2.8 $\pm$ 1.9	1.9 $\pm$ 1.9	1		1.4 $\pm$ 1.4	2.0 $\pm$ 2.0
11			3.8 $\pm$ 2.7	4	5.3 $\pm$ 3.6	5.7 $\pm$ 2.8	8.0 $\pm$ 3.8
12			1.9 $\pm$ 1.9	5	21.1 $\pm$ 6.6	28.6 $\pm$ 5.4	2.0 $\pm$ 2.0
16			1.9 $\pm$ 1.9	6	18.4 $\pm$ 6.3	11.4 $\pm$ 3.8	4.0 $\pm$ 2.8
(n)	40	72	52	8			6.0 $\pm$ 3.4
<i>Mfd-18</i>				9	55.3 $\pm$ 8.1	51.4 $\pm$ 6.0	12.0 $\pm$ 5.0
1	27.5 $\pm$ 7.1	25.0 $\pm$ 5.1	21.4 $\pm$ 5.5	10			24.0 $\pm$ 6.0
4	30.0 $\pm$ 7.2	30.3 $\pm$ 5.4	17.9 $\pm$ 5.1	11			12.0 $\pm$ 5.0
5	2.5 $\pm$ 2.5	1.4 $\pm$ 1.4		12		1.4 $\pm$ 1.4	18.0 $\pm$ 5.4
6	12.5 $\pm$ 5.3	12.5 $\pm$ 3.9	19.6 $\pm$ 5.3	13			6.0 $\pm$ 3.4
7	2.5 $\pm$ 2.5	1.4 $\pm$ 1.4	16.1 $\pm$ 4.9	15			4.0 $\pm$ 2.8
8			5.4 $\pm$ 3.0	16			2.0 $\pm$ 2.0
9			1.8 $\pm$ 1.8	(n)	38	70	50
10	12.5 $\pm$ 5.3	22.2 $\pm$ 4.9	8.9 $\pm$ 3.8	<i>Pla2a</i>			
11	12.5 $\pm$ 5.3	6.9 $\pm$ 3.0	5.4 $\pm$ 3.0	1			3.8 $\pm$ 2.7
12			3.6 $\pm$ 2.5	3			15.4 $\pm$ 5.0
(n)	40	72	56	4			23.1 $\pm$ 5.8
<i>Mfd-23</i>				5	2.6 $\pm$ 2.6	2.9 $\pm$ 2.0	5.8 $\pm$ 3.2
1	16.7 $\pm$ 6.2	17.6 $\pm$ 4.6	18.5 $\pm$ 5.3	6	13.2 $\pm$ 5.5	12.9 $\pm$ 4.0	13.5 $\pm$ 4.7
3			3.7 $\pm$ 2.6	7	60.5 $\pm$ 7.9	64.3 $\pm$ 5.7	11.5 $\pm$ 4.4
4	16.7 $\pm$ 6.2	17.6 $\pm$ 4.6	1.9 $\pm$ 1.8	8	2.6 $\pm$ 2.6	1.4 $\pm$ 1.4	17.3 $\pm$ 5.2
5			5.6 $\pm$ 3.1	9	21.1 $\pm$ 6.6	18.6 $\pm$ 4.6	7.7 $\pm$ 3.7
7			5.6 $\pm$ 3.1	11			1.9 $\pm$ 1.9
9			20.4 $\pm$ 5.5	(n)	38	70	52
10			7.4 $\pm$ 3.6	<i>LL</i>			
12			7.4 $\pm$ 3.6	1			3.8 $\pm$ 2.7
13			7.4 $\pm$ 3.6	2			7.7 $\pm$ 3.7
14	2.8 $\pm$ 2.7	1.5 $\pm$ 1.5	7.4 $\pm$ 3.6	3	26.3 $\pm$ 7.1	32.9 $\pm$ 5.6	7.7 $\pm$ 3.7
15			3.7 $\pm$ 2.6	4	39.5 $\pm$ 7.9	32.9 $\pm$ 5.6	21.2 $\pm$ 5.7
19			5.6 $\pm$ 3.1	5			13.5 $\pm$ 4.7
20	5.6 $\pm$ 3.8	8.8 $\pm$ 3.4	3.7 $\pm$ 2.6	6		1.4 $\pm$ 1.4	5.8 $\pm$ 3.2
21	8.3 $\pm$ 4.6	5.9 $\pm$ 2.9	1.9 $\pm$ 1.8	7	21.1 $\pm$ 6.6	21.4 $\pm$ 4.9	9.6 $\pm$ 4.1
22	22.2 $\pm$ 6.9	26.5 $\pm$ 5.4		8	2.6 $\pm$ 2.6	1.4 $\pm$ 1.4	1.9 $\pm$ 1.9
24	22.2 $\pm$ 6.9	17.6 $\pm$ 4.6		9			11.5 $\pm$ 4.4
26	5.6 $\pm$ 3.8	4.4 $\pm$ 2.5		10			9.6 $\pm$ 4.1
(n)	36	68	54	11		2.9 $\pm$ 2.0	3.8 $\pm$ 2.7
<i>Rena4</i>				12	5.3 $\pm$ 3.6	2.9 $\pm$ 2.0	1.9 $\pm$ 1.9
1			1.9 $\pm$ 1.8	13	5.3 $\pm$ 3.6	4.3 $\pm$ 2.4	1.9 $\pm$ 1.9
2	41.2 $\pm$ 8.4	39.1 $\pm$ 6.1	9.3 $\pm$ 3.9	(n)	38	70	52
3	58.8 $\pm$ 8.4	60.9 $\pm$ 6.1	87.0 $\pm$ 4.6				
4			1.9 $\pm$ 1.8				
(n)	34	64	54				

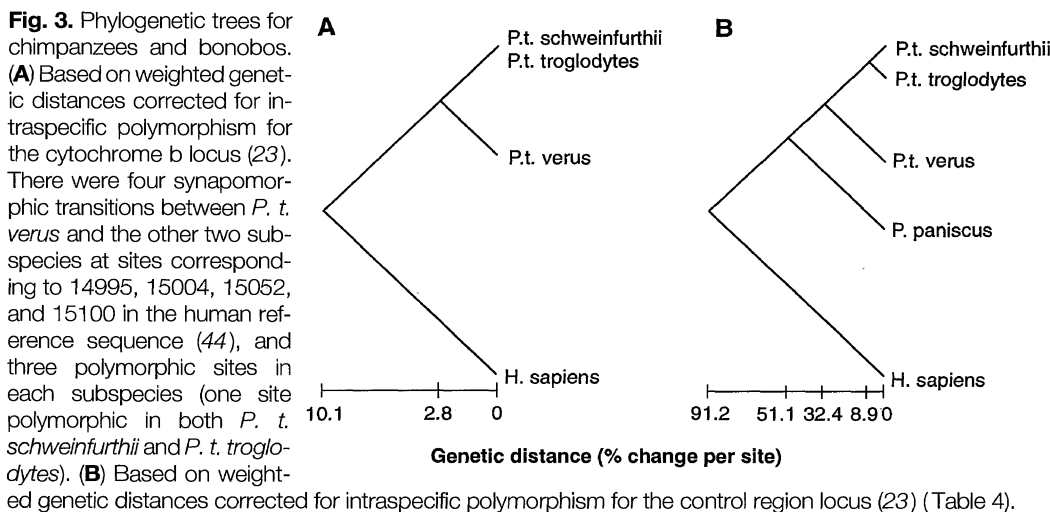
those of half-sibs only 50% of the time (expected if the distributions are the same), whereas female allele-sharing values were lower 65% of the time. Furthermore, male values were lower than female values only 26% of the time. Finally, a Kolmogorov-Smirnov one-tailed test for goodness of fit among distributions of allele-sharing values of non-sib, non-parent-offspring pairs of males compared to all maternal sib values was not significant, indicating that the distributions are not significantly different. In contrast, the corresponding test for female compared to maternal sib values was significant ( $P \leq 0.05$ ).

These genetic data provide formal genetic evidence that the evolution of the sexual differences in social behavior in chimpanzees may be partially explained by kin selection theory. The hypothesis that the more closely related males form a kin group that cooperates to defend a territory, thereby increasing access to females and resources, can now be tested more rigorously and at other sites. The relation of allele-sharing to kinship coefficients can be determined more thoroughly as more data on pedigree relationships accrue, allowing quantitative application of kin selection theory to evolu-

tionary studies of wild chimpanzees.

Allele frequencies and their standard errors were calculated for each of the eight loci in the Gombe Kasakela community (separately for unrelated individuals and for all individuals sampled) and the pan-African (non-Gombe) sample (Table 1). Kolmogorov-Smirnov tests for the significance of the difference between allele frequencies in two samples were performed on the Gombe and pan-African samples to determine whether allele frequencies were similar in the local population and continent-wide sample; significant differences ( $P < 0.05$ ) were detected at two loci, *Mfd-23* and *Pla2a*. This indicates that the pan-African sample, at least at two loci, does not adequately represent the population of which the Gombe community is part. Subdivision of the species into three genetically distinct groups (subspecies) and population structuring into social communities (see below) are most likely the reasons for the observed allele distribution differences. Comparisons between the data sets, however, reveal relative patterns that indicate the probable causes of substructuring.

If female dispersal was high enough to maintain a large and effectively panmictic population, we would expect pooled samples from across the species range and from individuals within a single social community to be in Hardy-Weinberg equilibrium (HWE). Deviations from the predictions of panmixia can be caused by several factors including nonrandom mating, distortions in the contributions of gametes to offspring, nonrandom sampling, Wahlund effect, strong selection, and allele mis-scoring [of which the last five are improbable in this case (11)]. Because these eight loci have not shown significant deviations from expectations of selective neutrality in humans (7), they are unlikely to cause distortions in the contributions of gametes to offspring or to deviate from expectations of panmixia because of strong selection in chimpanzees. Likewise, sampling methods were random with regard to these loci. Methods and accuracy of allele scoring have been described in (8). The assumptions of HWE in the Gombe sample are not satisfied according to the likelihood-ratio test at three loci (*Mfd3*, *FABP*, *LL*); the chi-square test corroborated these results in all but one case, that of the unrelated individuals at locus *Mfd3* (Table 2). Because the pan-African SSR samples comprise a collection of individuals from 15 sites across Africa, we would expect a larger number of significant departures from HWE in the pan-African sample if the departures were due only to population substructure (Wahlund's principle). The fact that there are two significant departures from HWE in the pan-African sample (loci *Mfd3* and *Mfd23*; likelihood-



**Table 2.** Comparison of observed heterozygote frequencies (Obs.) with HWE expectations (Exp.) based on (i) total frequency of heterozygotes, in which all heterozygotes were used to calculate the expected allele frequencies for each locus; which were compared to the observed allele frequencies by chi-square analysis, and (ii) the likelihood ratio test criterion ( $-2 \ln L$ ), computed following (47), but the empirical significance was evaluated by permuting the allelic labels across individuals for each locus-population combination by the procedure described in (48). This permutation test is mathematically equivalent to the Monte Carlo method of exact test of HWE (49). All significance tests were conducted by permuting alleles across individuals and with 2000 replications. \* $P < 0.05$ , \*\* $P < 0.01$ , and \*\*\* $P < 0.001$ .

Statistics	Population			Statistics	Population		
	Gombe	Non-Gombe			Gombe	Non-Gombe	
	Unrelated	Total			Unrelated	Total	
<i>Mfd3</i>							
N	20	36	26	N	20	36	27
Obs.	8	15*	19	Obs.	5***	12***	22
Exp. $\pm$ SE	11.6 $\pm$ 2.2	21.2 $\pm$ 3.0	21.9 $\pm$ 1.9	Exp. $\pm$ SE	13.8 $\pm$ 2.1	24.4 $\pm$ 2.8	21.6 $\pm$ 2.1
-2 ln L	21.0**	23.7**	48.5*	-2 ln L	26.8***	31.2***	15.9
<i>Mfd18</i>							
N	20	36	28	N	19	35	26
Obs.	14	27	23	Obs.	10	16	21
Exp. $\pm$ SE	16.1 $\pm$ 1.8	28.3 $\pm$ 2.5	24.0 $\pm$ 1.8	Exp. $\pm$ SE	11.1 $\pm$ 2.1	19.0 $\pm$ 2.9	22.5 $\pm$ 1.7
-2 ln L	16.7	18.4	33.9	-2 ln L	7.0	11.0	37.8
<i>Mfd23</i>							
N	18	34	27	N	17	32	27
Obs.	11**	25	19**	Obs.	6	11	7
Exp. $\pm$ SE	15.4 $\pm$ 1.5	28.4 $\pm$ 2.2	24.4 $\pm$ 1.5	Exp. $\pm$ SE	8.5 $\pm$ 2.1	15.5 $\pm$ 2.8	6.4 $\pm$ 2.2
-2 ln L	25.3	23.0	70.1**	-2 ln L	1.3	2.5	1.0
<i>Mfd32</i>							
N	19	35	25	N	19	35	26
Obs.	12	25	23	Obs.	9**	18**	22
Exp. $\pm$ SE	12.0 $\pm$ 2.1	22.6 $\pm$ 2.8	22.0 $\pm$ 1.6	Exp. $\pm$ SE	14.1 $\pm$ 1.9	26.1 $\pm$ 2.6	23.5 $\pm$ 1.5
-2 ln L	3.2	9.0	40.2	-2 ln L	24.1**	34.7**	54.1
<i>FABP</i>							
<i>Pla2a</i>							
<i>Rena4</i>							
<i>LL</i>							

ratio test) compared to three in the Gombe sample suggests that they may be caused by local inbreeding or to a greater degree of relatedness in this social community, resulting from either single-sex dispersal among small communities or recent changes in dispersal behavior at Gombe, rather than prolonged genetic isolation of the population. In addition, all significant deviations in the Gombe sample are toward deficiencies in the number of heterozygotes, as expected if inbreeding or high male-male relatedness were the cause.

Nonrandom association of alleles from genetically unlinked loci (8) (deviations from gametic phase equilibrium) may be interpreted as evidence for either population substructure or selection. To examine this we used a test for independent segregation of alleles (Table 3). Because there were significant differences between observed and expected heterozygosities, the chosen test included both values for all loci. The analyses showed that observed variances of the number of heterozygous loci per individual were always within the 95% confidence limits, provided that there was independent segregation of alleles at different loci. Thus, despite the greater relatedness and homozygosity in the Kasakela community, significant cosegregation of alleles at these loci was not observed, and extensive inbreeding is not indicated.

The results showing patterns of deviation from the expectations of HWE, in-

creased homozygosity, and no deviations from gametic phase equilibrium, together with the significant differences in the levels of relatedness among males compared to that among females, strongly support the hypothesis that premating single-sex dispersal is the primary source of genetic substructuring in this community, rather than recent changes in dispersal behavior of Gombe females or genetic isolation of the Gombe community leading to recent increases in inbreeding levels. The tests do not, however, exclude these recent changes as contributing factors, so that continued genetic monitoring of this community and comparative studies are recommended. Evidence of similar genetic patterns in other communities would further support the hypothesis that kin selection has been a strong force in the evolution of chimpanzee social structure.

Gene Flow and Phylogenetics

Systematists recognize two species in the genus *Pan*, the bonobo or pygmy chimpanzee (*P. paniscus*) of Zaire and the chimpanzee (*P. troglodytes*). The latter has an unusually wide distribution across equatorial Africa and in the past has been variously assigned to as many as 17 species and 34 subspecies. The three well-known subspecies are: western masked or pale-faced *P. t. verus*, central black-faced *P. t. troglodytes*, and eastern long-haired *P. t. schweinfurthii*

(12). Vernacular names and minor cranio-metric variation notwithstanding, these geographically defined allopatric subspecies cannot be distinguished morphologically in captivity (13), so most previous genetic studies were limited because they involved captive animals of unknown geographic origin and taxonomic identity (14, 15). We therefore conducted a genetic survey of mitochondrial variation throughout the range of *P. troglodytes*, widely sampling within and among the geographic ranges of the three subspecies.

The 178-bp cyt b sequence was highly conserved within subspecies and no fixed differences were found between *P. t. troglodytes* and *P. t. schweinfurthii*. In contrast, a 2.8% genetic distance was found to clearly separate western *P. t. verus* from the other two subspecies (Fig. 3A). This short segment was chosen simply because we could look for differences among subspecies while using a fairly well-characterized gene locus. To look at patterns of intrasubspecific variation, however, and more clearly resolve the relationships between subspecies, we used sequences of a longer segment of the control region. The 345-bp control region sequence was far more variable both within and among subspecies, having 124 (35.5%) variable positions, including two single base deletions. Within *P. t. schweinfurthii*, there were 25 haplotypes (distinctive sequences; missing data were excluded) among 37 individuals, and 6 haplotypes

**Table 3.** Test of association of alleles among loci from the distribution of heterozygous loci. Only 57 individuals tested for all eight loci are included in this test. The expected distributions of the number of heterozygous loci and the theoretical mean, variance, and 95% confidence interval (CI) of variance are reported. In method (1) the observed proportions of heterozygotes for

each locus (Table 2) were used for computations; in method (2) the unbiased estimates of heterozygosities (50) were used. Tests for nonrandom association of alleles among loci from the distribution of heterozygous loci were performed using the method described (51), following the theory of Brown *et al.* (52). An alternative method (53) provides equivalent results.

Number of heterozygous loci	Population								
	Gombe						Non-Gombe		
	Unrelated			Total					
	Exp.		Obs.	Exp.		Obs.	Exp.		Obs.
	1	2		1	2		1	2	
0	0	0.0	0	0	0.0	0	0.0	0.0	0
1	0	0.4	0	0	0.5	0	0.0	0.0	0
2	1	1.6	1	1	2.2	0	0.1	0.0	0
3	2	3.5	4	4	5.8	0	0.4	0.1	0
4	2	4.3	7	7	8.6	5	2.0	0.9	5
5	7	3.3	11	7	7.7	3	5.5	3.7	3
6	3	1.5	5	5	4.0	7	8.4	8.4	7
7	0	0.4	2	2	1.1	5	6.2	8.9	5
8	0	0.0	0	0	0.1	4	1.4	2.1	4
Total	15.0	15.0	30	30.0	30.1	24	24.0	24.1	
Mean	4.600	3.946	4.700	4.265	5.302	6.000	5.904	6.295	
Variance	1.400	1.835	1.459	1.775	1.684	1.913	1.254	0.989	
95% CI of variance									
Lower		0.593		0.922	0.871		0.545	0.400	
Upper		3.077		2.629	2.497		1.963	1.577	

were shared among 2 to 6 individuals. Within *P. t. troglodytes*, there were 17 haplotypes among 18 individuals, and within *P. t. verus*, all 11 sequences were different from each other; no haplotypes were shared between subspecies. Several synapomorphies (shared derived characters) were fixed or nearly fixed in each subspecies (Fig. 4). Despite the large variation within subspecies, cladistic analysis shows that 10 fixed (and 2 slightly polymorphic) synapomorphies link the eastern and central subspecies to the exclusion of the western subspecies, which in turn exhibits 11 fixed and 1 polymorphic (in one individual) synapomorphies that

bind all *P. t. verus* to the exclusion of the others (Fig. 4). Using the neighbor joining method, a phylogenetic tree was constructed from the pairwise distances (Fig. 5; summarized in Fig. 3B). Alternative parsimony and distance cluster analyses also yielded trees with the same subspecies clades and very similar subclusters within subspecies (16, 17).

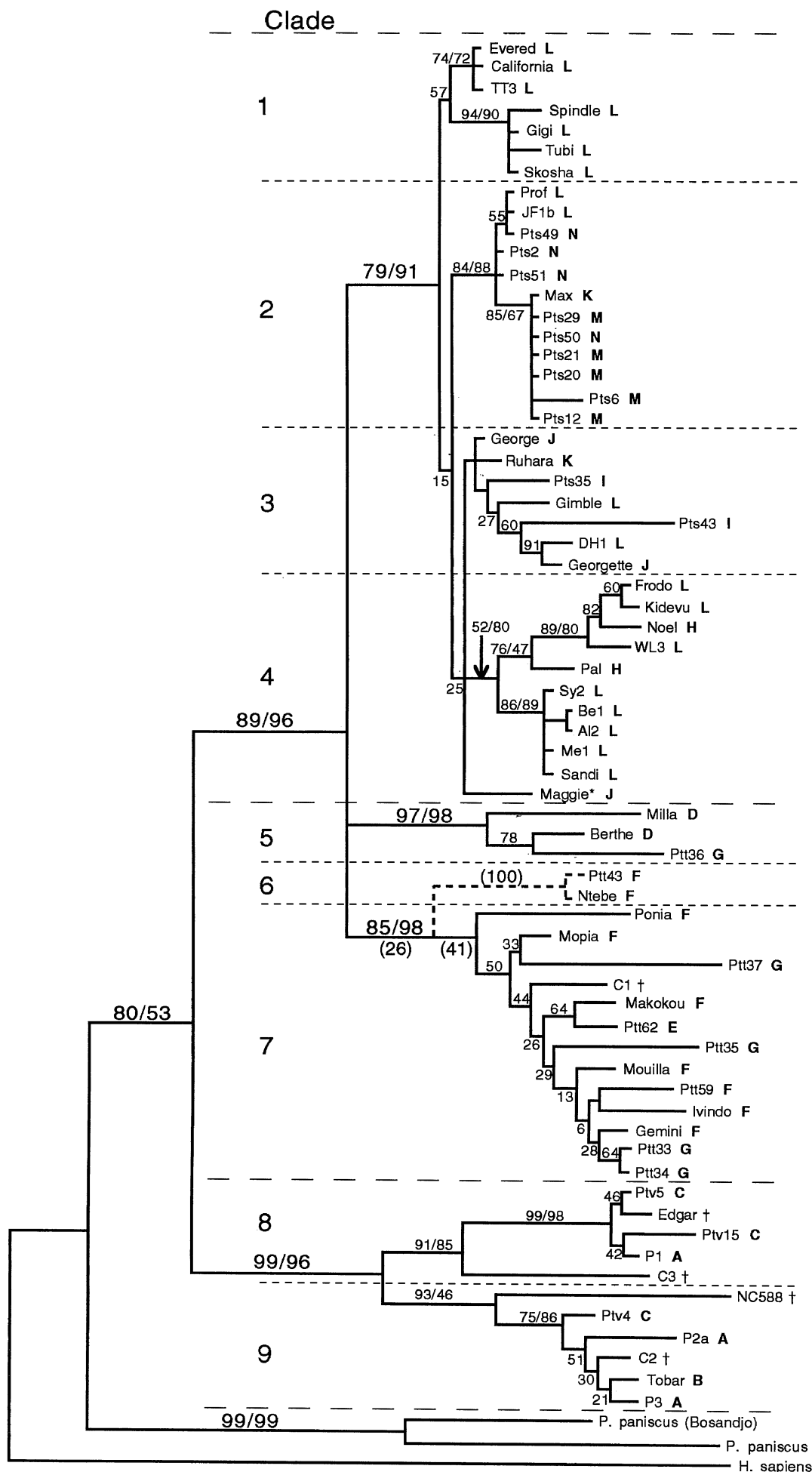
Of 24 adults in the Kasakela social community at Gombe in July 1991, 19 individuals known or suspected to be maternally unrelated were sampled for sequencing; we found 15 different haplotypes in the control region. This surprisingly high within-community genetic variation is attributable to

gene flow based on female dispersal behavior in this species (18). The sequence similarities of some individuals from Gombe to individuals from a geographic range covering over 600 km suggests that historical gene flow has been significant. In the last 50 years, however, the 32-km<sup>2</sup> Gombe National Park, now occupied by 150 to 160 chimpanzees, has become completely isolated from other habitats by deforestation (19). In the absence of gene flow with other populations, genetic erosion due to genetic drift and inbreeding may begin to significantly increase the probability of extirpation of this and other populations experiencing genetic isolation and habitat frag-

		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80	81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	100	101	102	103	104	105	106	107	108	109	110	111	112	113	114	115	116	117	118	119	120	121	122	123	124	125	126	127	128	129	130	131	132	133	134	135	136	137	138	139	140	141	142	143	144	145	146	147	148	149	150	151	152	153	154	155	156	157	158	159	160	161	162	163	164	165	166	167	168	169	170	171	172	173	174	175	176	177	178	179	180	181	182	183	184	185	186	187	188	189	190	191	192	193	194	195	196	197	198	199	200	201	202	203	204	205	206	207	208	209	210	211	212	213	214	215	216	217	218	219	220	221	222	223	224	225	226	227	228	229	230	231	232	233	234	235	236	237	238	239	240	241	242	243	244	245	246	247	248	249	250	251	252	253	254	255	256	257	258	259	260	261	262	263	264	265	266	267	268	269	270	271	272	273	274	275	276	277	278	279	280	281	282	283	284	285	286	287	288	289	290	291	292	293	294	295	296	297	298	299	300	301	302	303	304	305	306	307	308	309	310	311	312	313	314	315	316	317	318	319	320	321	322	323	324	325	326	327	328	329	330	331	332	333	334	335	336	337	338	339	340	341	342	343	344	345	346	347	348	349	350	351	352	353	354	355	356	357	358	359	360	361	362	363	364	365	366	367	368	369	370	371	372	373	374	375	376	377	378	379	380	381	382	383	384	385	386	387	388	389	390	391	392	393	394	395	396	397	398	399	400	401	402	403	404	405	406	407	408	409	410	411	412	413	414	415	416	417	418	419	420	421	422	423	424	425	426	427	428	429	430	431	432	433	434	435	436	437	438	439	440	441	442	443	444	445	446	447	448	449	450	451	452	453	454	455	456	457	458	459	460	461	462	463	464	465	466	467	468	469	470	471	472	473	474	475	476	477	478	479	480	481	482	483	484	485	486	487	488	489	490	491	492	493	494	495	496	497	498	499	500	501	502	503	504	505	506	507	508	509	510	511	512	513	514	515	516	517	518	519	520	521	522	523	524	525	526	527	528	529	530	531	532	533	534	535	536	537	538	539	540	541	542	543	544	545	546	547	548	549	550	551	552	553	554	555	556	557	558	559	560	561	562	563	564	565	566	567	568	569	570	571	572	573	574	575	576	577	578	579	580	581	582	583	584	585	586	587	588	589	590	591	592	593	594	595	596	597	598	599	600	601	602	603	604	605	606	607	608	609	610	611	612	613	614	615	616	617	618	619	620	621	622	623	624	625	626	627	628	629	630	631	632	633	634	635	636	637	638	639	640	641	642	643	644	645	646	647	648	649	650	651	652	653	654	655	656	657	658	659	660	661	662	663	664	665	666	667	668	669	670	671	672	673	674	675	676	677	678	679	680	681	682	683	684	685	686	687	688	689	690	691	692	693	694	695	696	697	698	699	700	701	702	703	704	705	706	707	708	709	710	711	712	713	714	715	716	717	718	719	720	721	722	723	724	725	726	727	728	729	730	731	732	733	734	735	736	737	738	739	740	741	742	743	744	745	746	747	748	749	750	751	752	753	754	755	756	757	758	759	760	761	762	763	764	765	766	767	768	769	770	771	772	773	774	775	776	777	778	779	780	781	782	783	784	785	786	787	788	789	790	791	792	793	794	795	796	797	798	799	800	801	802	803	804	805	806	807	808	809	810	811	812	813	814	815	816	817	818	819	820	821	822	823	824	825	826	827	828	829	830	831	832	833	834	835	836	837	838	839	840	841	842	843	844	845	846	847	848	849	850	851	852	853	854	855	856	857	858	859	860	861	862	863	864	865	866	867	868	869	870	871	872	873	874	875	876	877	878	879	880	881	882	883	884	885	886	887	888	889	890	891	892	893	894	895	896	897	898	899	900	901	902	903	904	905	906	907	908	909	910	911	912	913	914	915	916	917	918	919	920	921	922	923	924	925	926	927	928	929	930	931	932	933	934	935	936	937	938	939	940	941	942	943	944	945	946	947	948	949	950	951	952	953	954	955	956	957	958	959	960	961	962	963	964	965	966	967	968	969	970	971	972	973	974	975	976	977	978	979	980	981	982	983	984	985	986	987	988	989	990	991	992	993	994	995	996	997	998	999	1000
Evered	GOMBE	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..	..</																																																																																																																																																																																																																																																																

Fig. 4. Individual chimpanzee by control region nucleotide site matrix for nucleotide positions variable in two or more *P. troglodytes* individuals. Nucleotide sites have been rearranged to cluster markers for each clade (clades and

subspecies separations match those in Fig. 5, described in text); sites 1 to 345 refer to the human reference sequence (44), with 1 corresponding to position 16047; ? = missing data; reference = chimpanzee consensus sequence.



**Fig. 5.** Phylogenetic tree constructed using the neighbor joining method based on genetic distances estimated from control region sequences. Genetic distances were computed using the standard Kimura two-parameter model in MEGA software (16). Sample origins are indicated by letters A to N after sample identities, and correspond to letters on the map (Fig. 1). Clades referred to in the text are numbered 1 to 9; clades 1 to 4 are *P. t. schweinfurthii*; 5 to 7 are *P. t. troglodytes*; 8 and 9 are *P. t. verus*. The Ntebe-Ptt43 sequences are substantially different from other sequences of that subspecies, and were found in two Gabonese individuals separated by 400 km. Their branch position on the tree is indicated by a dashed line, and inclusion of these sequences in bootstrap analyses yielded the values in parentheses for clades 6 and 7 (others were not significantly affected). The reliability of each interior branch was tested by 500 bootstrap replications and a standard error test in the MEGA software package. Where single numbers occur on branches, they represent the bootstrap confidence levels (BCL); standard error confidence probabilities (CP) are shown on some branches in the format BCL/CP. Sites containing missing data and alignment gaps were not used in the bootstrap and standard error tests. \*Maggie was included in clade 4 for simplicity because the branch was not significantly supported. †Samples of unknown geographic origin.

mentation (20). In that our data on local variation in nuclear genes (and field observations) provide no evidence for inbreeding depression in this community yet, we note that habitat loss and exposure to human-borne diseases are still more serious threats. The extent to which population isolation has or will affect other aspects of community structure and behavior remains to be investigated.

In the eastern subspecies, *P. t. schweinfurthii*, the samples include 32 individuals of precisely known geographic origin and 7 of known country of origin. Both parsimony and neighbor joining (Fig. 5) methods created trees with four primary clades (clades 1 to 4) within this subspecies, each with identical individual composition. There is some geographic differentiation; clade 3 contains primarily samples from the northernmost sites (Uganda, Rwanda, and Burundi) and clade 2 includes all nine apes from the two southernmost sites (Mahale and Tongwe). Widespread historical gene flow is evidenced, however, by inclusion of the geographically intermediate Gombe genotypes in all four clades. The absence of any of the southernmost samples in "northern" clade 3, and of northernmost samples in "southern" clade 2, indicates possible isolation by distance beyond about 600 km.

Control region variation provides evidence for similar high historical gene flow patterns in the other two subspecies. Within the central subspecies, *P. t. troglodytes*, we can show three clades (clades 5 to 7)



**Table 4.** Variation among individuals within and between subspecies for the control region sequence. On and below diagonal: mean weighted genetic distances corrected for intrasubspecific polymorphism (23) and mean num-

ber of transitions (ts) and transversions (tv), not corrected for missing data (ts/tv); above diagonal: time since divergence (myr + 95% CI).

Taxon	All Pt (%)	Pts (%)	Ptt (%)	Ptv (%)	Pp (%)	Human (%)
All <i>P. troglodytes</i>	20.19	NA	NA	NA	2.5 (2.0–3.0)†	4.7 (4.2–5.2)*
<i>P. t. schweinfurthii</i>	NA	4.0 (7/0.9)	0.44 (.21–1.64)	1.58 (.75–5.96)	NA	NA
<i>P. t. troglodytes</i>	NA	8.9 (17.5/3.2)	9.0 (13.4/2)	1.58 (.75–5.96)	NA	NA
<i>P. t. verus</i>	NA	32.4† (27/7)	32.4† (29/9.8)	6.3 (14.7/1.3)	NA	NA
<i>P. paniscus</i>	51.1 (29.8/14.7)	57.4 (NA)	62.3 (NA)	54.3 (NA)	26.6 (29.7/15)	NA
Human	91.2 (34.8/20.9)	99.3 (NA)	100.7 (NA)	94.3 (NA)	85.2 (38.5/20)	2.9‡ (NA)

\*Data from (26).

†Mean weighted distance to *P. t. schweinfurthii* and *P. t. troglodytes* combined, corrected for intrasubspecific polymorphism.

‡Data from (24).

involving 18 individuals from known localities in Gabon, Equatorial Guinea, Cameroon, and Congo (Fig. 5). Clade 7 comprises individuals from Gabon, Equatorial Guinea, and Congo which cluster together without any detectable geographic substructure over 800 km. Clade 5 was observed in the northern part of the subspecies range, but sympatric with members of clade 7. Clade 6 includes two individuals separated by approximately 400 km within Gabon. Additional variation in this subspecies is to be expected because our present geographic sampling in the north and east of its range is inadequate.

The third subspecies, *P. t. verus*, from western Africa is the most vulnerable today (21) and has the smallest geographic range. Samples from chimpanzees from Mali and Ivory Coast, separated by up to 900 km, shared nearly identical sequences from the two widely divergent mtDNA haplotype groups (clades 8 and 9) indicating, again, a historical pattern of long-distance gene flow. The trees based on these control region data (Figs 3B and 5) indicate that the earliest divergence was between the western African *P. troglodytes* clades and the lineage leading to the other two subspecies. The branch (and corrected genetic distances) (Table 4) connecting *P. t. verus* to the other two subspecies is relatively deep, suggesting a long separation between these two groups. In addition, the other two subspecies show substantially less divergence and less intrasubspecific polymorphism that has arisen since subspecies divergence; the relatively deep split within *P. t. troglodytes* may represent ancestral polymorphism (as shown in Fig. 5, clade 5), and accounts for the higher variation within *P. t. troglodytes* than within the western subspecies. The presence of this ancestral polymorphism, as well as relatively less variation within clades of *P. t. troglodytes* than within *P. t. verus*, supports a longer isolation of *P. t. verus* as a subspecies.

These patterns of substantial genetic variability indicate that populations have been sufficiently large since divergence of each subspecies to allow accumulation and maintenance of genetic variation. The observation that the variation is dispersed

throughout the range of each subspecies further indicates that there has been substantial historical gene flow. These data are not consistent with a recent expansion of any subspecies populations, or with a rapid expansion followed by population isolation. Studies of larger samples from other populations within each subspecies range should allow quantitative evaluation of historical gene flow rates.

Comparisons of these cyt b and control region sequences provide insight into the evolutionary relations and ages of these taxa (22). At the cyt b locus, the corrected genetic distance (23–25) between *Homo* and *Pan* is 10.1%. A  $4.7 \pm 0.5$  million year [myr  $\pm$  95% confidence interval (CI)] estimated divergence time between humans and chimpanzees (26, 27), yields a substitution rate of  $2.15\% \text{ myr}^{-1}$  (95% CI range, 0 to  $6.37\% \text{ myr}^{-1}$ ). This calibration gives a divergence time based on cyt b between *P. t. verus* and the two other subspecies (corrected  $d = 2.8\%$ ) of about 1.3 myr (95% CI range: 0.44 to  $>2.5$  myr) (28). Calibration by means of an earlier human-ape divergence [for example, 8 myr (29)], would yield a correspondingly earlier split within *P. troglodytes* (2.2 myr, roughly contemporaneous with the origin of the genus *Homo*).

To calibrate the control region data, we used estimates of divergence time between *P. paniscus* and *P. troglodytes* of  $2.5 \pm 0.5$  myr ago, based on previous calibrations of this and other, more conserved, regions of the mitochondrial genome (26, 30, 31). According to Nei (28, 32), the substitution rate is then  $20.4\%$  (5.43 to  $42.96\%$ )  $\text{myr}^{-1}$  and the derived divergence times between the subspecies are given in Table 4. The calculated control region based divergence time between *P. t. verus* and the two other subspecies is 1.58 myr. *Pan paniscus* is seen to lie closer to the *Pan-Homo* split as postulated by Zihlman and others (33). Because gene trees may not reflect correct species trees and because standard errors associated with relatively short DNA sequences are large, further discussion of the bearing of these results on the Pliocene-Pleistocene speciation of these apes should await phylogenetic analysis of more loci (34).

These data indicate that at the sequences examined *P. t. verus* is more differentiated from the other two subspecies than are some other full species of mammals (35). The average genetic distance between the western subspecies and the other two is two-thirds the genetic distance between the two chimpanzee species *P. paniscus* and *P. troglodytes*. This degree of differentiation within *P. troglodytes* is greater than that reported for the cytochrome oxidase II sequence (36). The probable isolation of *P. t. verus* for about 1.6 myr is greater than the estimated mean duration of late Cenozoic mammal species (37). In our opinion, if this result is confirmed at three or more unlinked nuclear loci (34), and supported by eco-behavioral data, *P. t. verus* merits elevation to full species rank: *P. verus* (Schwartz) 1934.

## Implications

Our demonstration that free-ranging chimpanzees can be genotyped noninvasively has resulted in four significant findings. First, our observation that, within a community, males are more closely related to one another than are females, and that this difference can account for genetic substructuring of the population, constitutes a substantial preliminary test of a sociobiological hypothesis concerning the evolution of cooperative behavior in males. Second, our discovery that dispersal, as evidenced by maternally transmitted genotypes, is detectable over distances of 600 to 900 km within each subspecies provides evidence that supports the hypothesis that high amounts of gene flow may account for the low amounts of morphological differentiation in chimpanzees (38). Third, our discovery that the West African *P. t. verus* is actually a well-differentiated and independently evolving taxon has important implications for both evolutionary and conservation biology. Although exact climatic or ecological (or both) patterns of change in equatorial Africa over the past 2 million years are unclear, repeated cycles of drying possibly associated with hypothermal periods occurred and periodically reduced forests to western, central, and eastern refugia. Extant chimpanzee taxa may have differentiated from

one another in such forest fragments (39). Reconstructing the history of the genus *Pan* and of our common ancestor will have to allow for the previously underappreciated variation within chimpanzees. This may help to resolve the gorilla-chimpanzee-human "trichotomy" question (40) by improving the resolution of synapomorphies over homoplasy (the phylogenetic "signal"). Fourth, our discovery of considerable genetic variation within chimpanzee populations across broad geographic ranges provides us with a way of quantifying genetic diversity in these endangered species and monitoring genetic erosion in both wild and captive populations.

Conservation efforts in the wild and in captivity need to account for the genetic diversity in the genus *Pan* and its partitioning into several well-differentiated evolutionarily significant units (41). Although captive chimpanzees have traditionally been sorted into and managed as two species, the three subspecies of *P. troglodytes* have, for example, been afforded no special significance under current NIH colony management policies. Our results suggest that treating the >2000 chimpanzees in biomedical facilities in the United States as genetically equivalent requires reassessment. Inappropriately matched individuals are undesirable in both experimental and breeding situations, and some previous results based on genetically dissimilar apes may require reinterpretation (42). However, chimpanzee social, emotional, and intellectual similarities to humans render management from a purely genetic standpoint unethical; the behavioral or social (or both) needs of captive individuals must also be considered. We do not advocate breaking up mismatched but long-bonded pairs.

Finally, management of increasingly fragmented populations in Africa may require occasional translocation of individuals for their own protection and to preserve naturally occurring variability in the wild. Such inter-ventive management is fraught with hazards including disease introduction and the behavioral problems of translocated wild individuals and reintroduced captive-born or raised individuals (43). Our results suggest that the possibility of outbreeding depression must also be considered. Our appreciation of the natural diversity within these species may have important implications for their future evolution and for their use as models in biomedical, paleoanthropological, and evolutionary studies of humans.

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9. The section of the cyt b gene amplified corresponds to bases 14816 through 15174 of the human mitochondrial sequence (44); the sequenced portion corresponds to human positions 14964 to 15141. The primers are identified by a coded name, followed by the human nucleotide reference number of the 3' end and L or H to indicate light or heavy strand: *Gytb1* (L14841) 5'-CCATCCAACATCTCAGCATGATGAAA-3' and *Cytb2* (H15149) 5'-GCCCTCA-GAATGATATTTGCTCTCA-3' (15; without restriction sites). The section of the control region amplified corresponds to bases 16019 through 16428 of the human sequence (44), *D-88* (L16041) 5'-CTCTGT-TCTTTTCATGGGGAAGC-3' and *D-441* (H16407) 5'-CGGGATATTGATTTACCGGAGG-3' (K. Garner, Zool. Soc. San Diego, personal communication). Sequencing was done from both strands of the control region PCR product as described [J. C. Garza and D. S. Woodruff, *Mol. Phylogenet. Evol.* **1**, 202 (1992)]. Sequencing of the cyt b region was only performed with the heavy strand (*Cytb2*) primer for most samples. Cyt b sequences: GenBank L35346–L35379; control region sequences: GenBank L35380–L35442. Five additional published sequences were included in the analyses: three *P. troglodytes* (C1, C2, C3), one *P. paniscus* (plus another sequenced by us, GenBank L35443), and one human [D. R. Foran, J. E. Hixson, W. M. Brown, *Nucleic Acids Res.* **16**, 5841 (1988); T. D. Kocher and A. C. Wilson, in *Evolution of Life*, S. Osawa and T. Honjo, Eds. (Springer, Tokyo, 1991), pp. 391–413; A. Di Rienzo and A. C. Wilson, *Proc. Natl. Acad. Sci. U.S.A.* **88**, 1597 (1991)]. All sequences were aligned by hand. All chimpanzee control regions had a 1-bp deletion at human position 16077 (44). Chimpanzee sequences differed from one another by deletions of 1 or 2 bp at sites 127 and 128 (Fig. 4); two individuals also had deletions at site 250 (Fig. 4).
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- region sequences (24). We calculated control region *R*'s from all of the chimpanzee data and from each subspecies separately by counting transversions and transitions on the minimum length trees found by parsimony analysis (24) ( $R \pm 95\%$  CI; all *P. troglodytes*  $8.5\% \pm 3.52$ , *P. t. schweinfurthii*  $8.6 \pm 6.84$ , *P. t. troglodytes*  $10.1 \pm 7.06$ , *P. t. verus*  $18.0 \pm 21.36$ ); we found that, although they are quite variable, these values are not significantly different from Vigilant *et al.*'s estimate. Divergence rates were determined (24, 28). Further, the estimate of  $R = 15$  is probably low [J. Wakeley, *Mol. Biol. Evol.* **11**, 436 (1994); M. Ruvolet *et al.*, *ibid.* **10**, 1115 (1993); C. Wills, *Evolution*, in press] and results in a more recent estimate of divergence than if higher *R*'s were used in distance calculations.
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