

A detailed microscopic image of biological tissue, likely a histological section. It features a complex network of dark, wavy, and branching structures, possibly representing blood vessels or connective tissue, set against a lighter, more granular background. The overall color palette is dominated by dark greens, blacks, and light greys, with some yellowish highlights.

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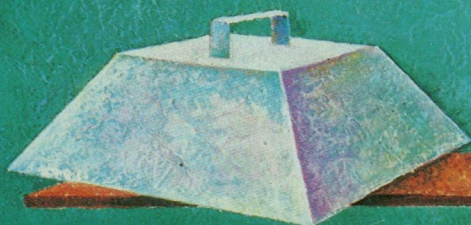
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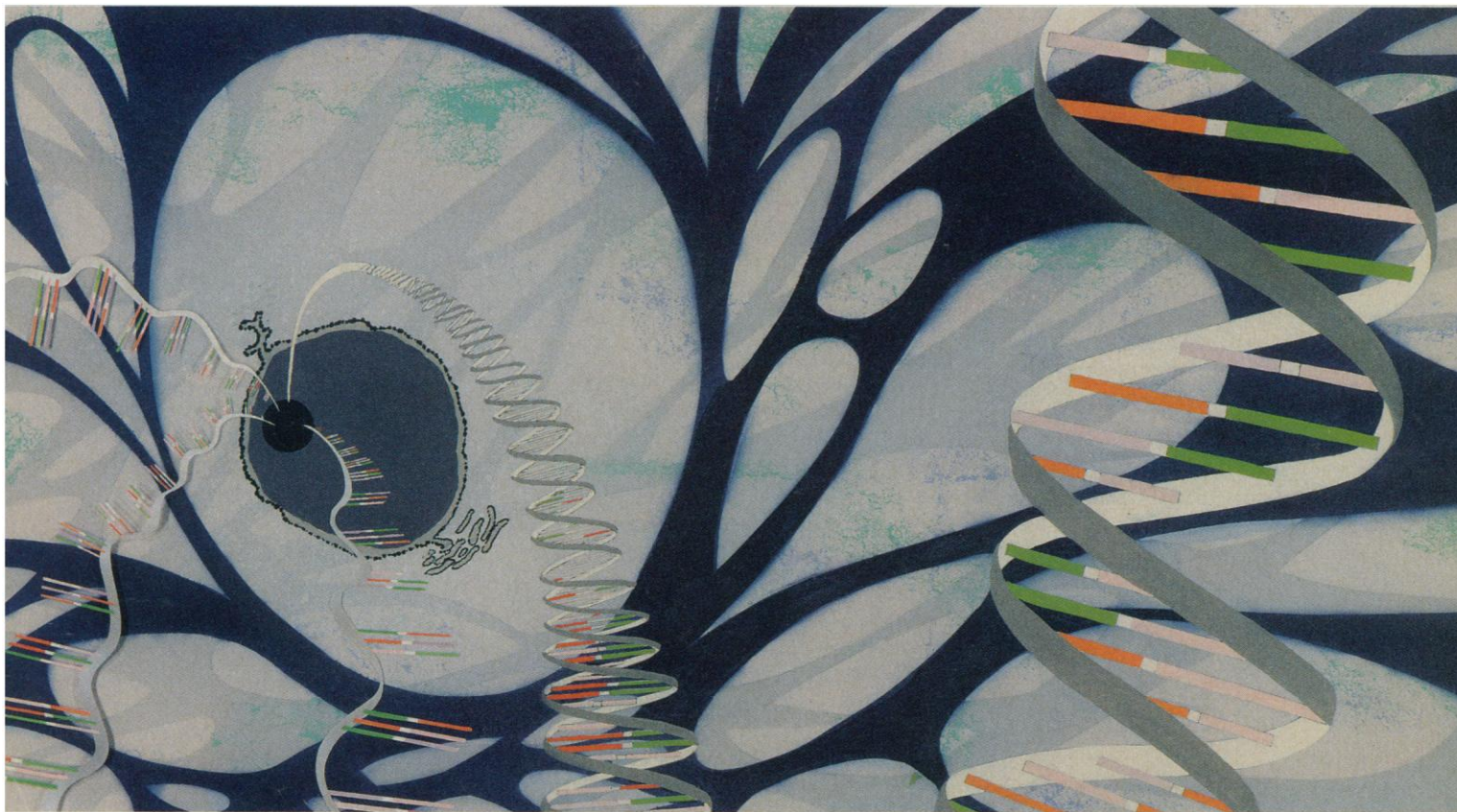


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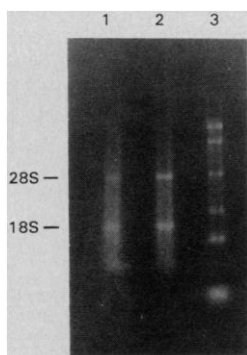
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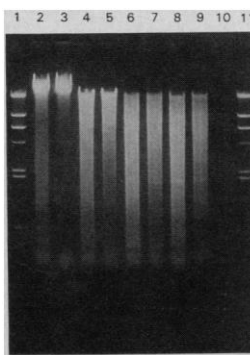
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1479 This Week in *Science*

Editorial

1481 The Dirty Air Act

Letters

1485 Soviet Role in SAGE: G. T. GARVEY ■ Asbestos Policy: A. M. LANGER, R. P. NOLAN, M. ROSS ■ Young Scientists and the Future: D. K. BODKIN; N. SMALHEISER ■ The Cellular Basis of Memory: B. L. McNAUGHTON

News & Comment

1492 Mad Cow Disease: Uncertainty Rules
1494 Female Primatologists Confer—Without Men
1495 Will AIDS Conference Migrate?
1496 NIH Urged to Be a Smart Shopper
1497 Genome Center Grants Chosen
Jittery Hubble Awaits a Cure
1498 *Briefings*: Out of China . . . and Now Back In ■ Feds Hush Up a Bum Bomb Detector ■ Retraining the Cranes ■ Chronic Leaks Plague Shuttle Science ■ Voice Lessons for Psychotics ■ Blocking the Backdaters ■ Human Deathtraps for Mosquitoes ■ FDA Gets a New Boss

Research News

1500 Proton Microbeam Probes the Elements
1502 Signs of the Parkfield Quake?
1503 Cystic Fibrosis Corrected in Lab
Partner Found for the Myc Protein
1504 Millimeter Astronomers Push for New Telescope

Articles

1513 Drug Policy: Striking the Right Balance: A. GOLDSTEIN AND HAROLD KALANT
1522 Magnetic Confinement Fusion: H. P. FURTH
1527 New Methods of Drug Delivery: R. LANGER
1533 Molecular Targets for AIDS Therapy: H. MITSUYA, R. YARCHOAN, S. BRODER

Research Article

1544 Structural Characterization of a Partly Folded Apomyoglobin Intermediate: F. M. HUGHSON, P. E. WRIGHT, R. L. BALDWIN

Reports

1549 Epitaxial and Smooth Films of *a*-Axis YBa₂Cu₃O₇: C. B. EOM, A. F. MARSHALL, S. S. LADERMAN, R. D. JACOWITZ, T. H. GEBALLE

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COVER A partial cross section of a biocompatible polymer system that delivers medications, including proteins, in a controlled manner. This high-powered light micrograph was taken after the polymer matrix had been releasing protein for more than 5 months. See page 1527. [Photomicrography by Rajan Bawa]

- 1552 Direct Interaction of a Ligand for the *erbB2* Oncogene Product with the EGF Receptor and p185^{erbB2}: R. LUPU, R. COLOMER, G. ZUGMAIER, J. SARUP, M. SHEPARD, D. SLAMON, M. E. LIPPMAN
- 1555 Retroviral DNA Integration Directed by HIV Integration Protein in Vitro: F. D. BUSHMAN, T. FUJIWARA, R. CRAIGIE
- 1558 Inhibition of T Cell Receptor Expression and Function in Immature CD4⁺CD8⁺ Cells by CD4: T. NAKAYAMA, C. H. JUNE, T. I. MUNITZ, M. SHEARD, S. A. MCCARTHY, S. O. SHARROW, L. E. SAMELSON, A. SINGER
- 1561 "Pure" Human Hematopoietic Progenitors: Permissive Action of Basic Fibroblast Growth Factor: M. GABBIANELLI, M. SARGIACOMO, E. PELOSI, U. TESTA, G. ISACCHI, C. PESCHLE
- 1564 Protection from Chemotherapy-Induced Alopecia in a Rat Model: A. M. HUSSEIN, J. J. JIMENEZ, C. A. MCCALL, A. A. YUNIS
- 1567 Recovery of Mitogenic Activity of a Growth Factor Mutant with a Nuclear Translocation Sequence: T. IMAMURA, K. ENGLEKA, X. ZHAN, Y. TOKITA, R. FOROUGH, D. ROEDER, A. JACKSON, J. A. M. MAIER, T. HLA, T. MACIAG
- 1570 Extension of the Life-Span of Human Endothelial Cells by an Interleukin-1 α Antisense Oligomer: J. A. M. MAIER, P. VOULALAS, D. ROEDER, T. MACIAG
- 1574 Cell-Adhesive Motif in Region II of Malarial Circumsporozoite Protein: K. A. RICH, F. W. GEORGE IV, J. L. LAW, W. J. MARTIN
- 1577 Molecular Analysis of Acute Promyelocytic Leukemia Breakpoint Cluster Region on Chromosome 17: J. BORROW, A. D. GODDARD, D. SHEER, E. SOLOMON
- 1580 Flip and Flop: A Cell-Specific Functional Switch in Glutamate-Operated Channels of the CNS: B. SOMMER, K. KEINÄNEN, T. A. VERDOORN, W. WISDEN, N. BURNASHEV, A. HERB, M. KÖHLER, T. TAKAGI *et al.*
- 1585 Coding Channels in the Taste System of the Rat: T. R. SCOTT AND B. K. GIZA

Book Reviews

- 1588 Cloth and Human Experience, *reviewed by* C. HENDERSON ■ The Household Economy, S. GUGGENHEIM ■ The Moon Illusion, D. R. PROFFITT ■ Books Received

Products & Materials

- 1598 Benchtop Monitoring of DNA Labeling ■ Liquid Chromatographs ■ Portable GC-MS Instruments ■ Nylon Transfer Membranes ■ Micromanipulators ■ Safe Lab Waste Disposal ■ Fast Reusable Cartridges ■ Literature

Author Index to volume 249 is found on pages I-X
Information for Contributors is found on pages XI-XII

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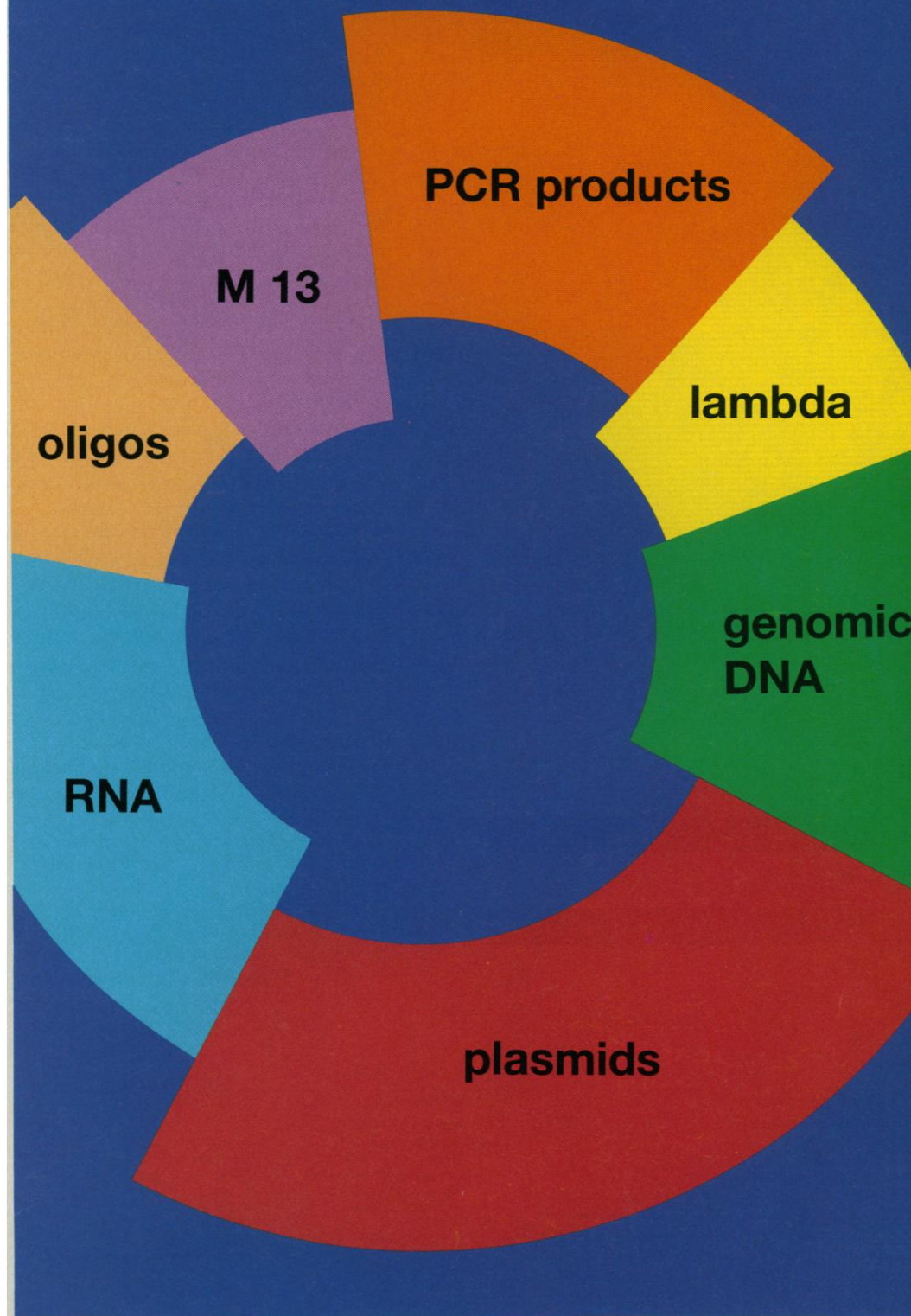
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This Week in SCIENCE

Drug delivery systems

No longer are clinicians limited to prescribing pills, drops, and injections for their patients. Recent advances in biotechnology, chemical engineering, and materials science and a growing understanding of human physiology and immunology have resulted in the development of many new approaches for preparing and delivering therapeutic drugs (cover). Drugs can now be absorbed from skin patches, delivered by external or implanted polymer systems and pumps, or administered embedded in vesicles (including such exotica as magnetic microparticles that can be guided through the body with external magnetic fields). The drugs themselves can be chemically altered in ways that make them more stable and soluble and able to enter previously inaccessible areas of the body. Drug release can be regulated, optimum concentrations can be maintained in the blood stream, and defined areas of the body can be targeted to receive high concentrations of the drug. An overview of these and other new possibilities for diversifying therapies is presented by Langer on page 1527.

Superconducting films

SUCCESS in making multilayer electronic devices from high-temperature superconductors has been elusive because fabrication is difficult to control. Smooth films have been grown with their *c*-axis perpendicular to the surface; for device applications, *a*-axis films would be desirable. Of special interest are superconducting films with a sandwich-type configuration—a Josephson junction consisting of two superconducting materials separated by a thin insulating barrier through which electron pairs might tunnel. Progress in this direction is reported by Eom *et al.* who have grown high-quality thin films of $\text{YBa}_2\text{Cu}_3\text{O}_7$ epitaxially—the atoms of the overlying film crystal and those of the substrate are in perfect alignment—on two different substrates (page 1549). Characterization of film microstructures and surface morphologies indicated that

they grew with the *a*-axis perpendicular to the surface and were atomically smooth, the latter a prerequisite for production of films with sandwich configuration tunnel junctions. Physical features that enhance or impede current flow and other structure-function relations can now be assessed for these new materials.

Alopecia model

HAIR loss (alopecia) is one of the more demoralizing side effects of chemotherapy. It can occur in conjunction with the use of a number of chemotherapeutic drugs, including cytosine arabinoside (ARA-C), doxorubicin (DX), and cyclophosphamide (CTX). An animal model of alopecia has now been developed and an antidote to alopecia may even be in the offing (page 1564): in young rats the bacterial product (and potential chemotherapeutic agent) ImuVert appears to prevent chemotherapy-induced hair losses. Hussein *et al.* report that in a protocol designed to test whether combinations of ImuVert and ARA-C would prevent development of leukemia in rats, alopecia was coincidentally prevented. ImuVert also prevented DX-induced alopecia but not CTX-induced alopecia. The distinction between ARA-C and DX on the one hand and CTX on the other suggests that different mechanisms may be involved in the induction of alopecia by these agents. Tests of whether ImuVert can prevent alopecia in humans may not be too far off: the level of toxicity of ImuVert in humans has already been determined and its efficacy as a chemotherapeutic agent against brain tumors is currently being evaluated.

Cellular lifespans

IN culture, human endothelial cells typically undergo about 60 population doublings and then stop growing and dividing. What accounts for this shift from active proliferation to senescence? One substance that appears to play a part is the protein interleukin-1 α (IL-1 α). Senescent endothelial

cells contain excess amounts of transcript (messenger RNA) for IL-1 α . In contrast, transformed cells, which are immortal, and young cells do not contain detectable amounts of this transcript. Daily exposure of cultured cells to oligodeoxynucleotides that have an antisense sequence—one that is complementary to that of the IL-1 α transcript—blocks the onset of senescence (page 1570). The cells keep on dividing and have the appearance of young endothelial cells. Later, when exposure to antisense molecules is stopped, the cells senesce, which indicates that the effects of antisense molecules are distinct from immortalization. Maier *et al.* propose that the antisense molecules may be repressing the translation of the IL-1 α transcript. The still-unanswered fundamental questions are why IL-1 α transcripts accumulate inside cells at the end of the cell's proliferative phase and exactly what happens when they do.

Flip and flop

STRUCTURAL variations in receptors may help an important neurotransmitter, L-glutamate, produce its diverse effects. L-Glutamate is the major excitatory substance in the mammalian nervous system; for fast neurotransmission, L-glutamate can bind to four types of related receptors. Each of the four receptors has been found to exist in two alternate active forms (page 1580): the so-called “flip” and “flop” forms differ by only a few amino acids. Whether the flip or flop form of the receptor is generated at any given time in any specific cell depends on how messenger RNA molecules are spliced. Sommer *et al.* have characterized the functional differences between the flip and flop forms of the receptors and have identified where in the brain and in which cells flip and/or flop is expressed. They speculate that at different times and places in development and in association with learning one form may be favored for carrying out requisite neurophysiologic functions and that inappropriate production of flip or flop may be at the root of certain neuropathologies.

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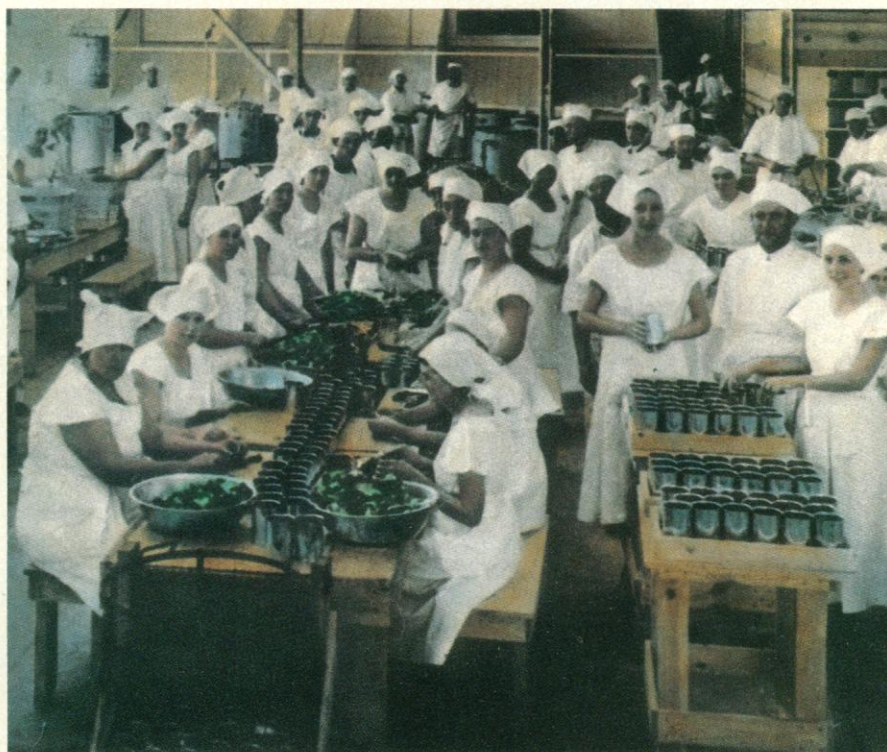
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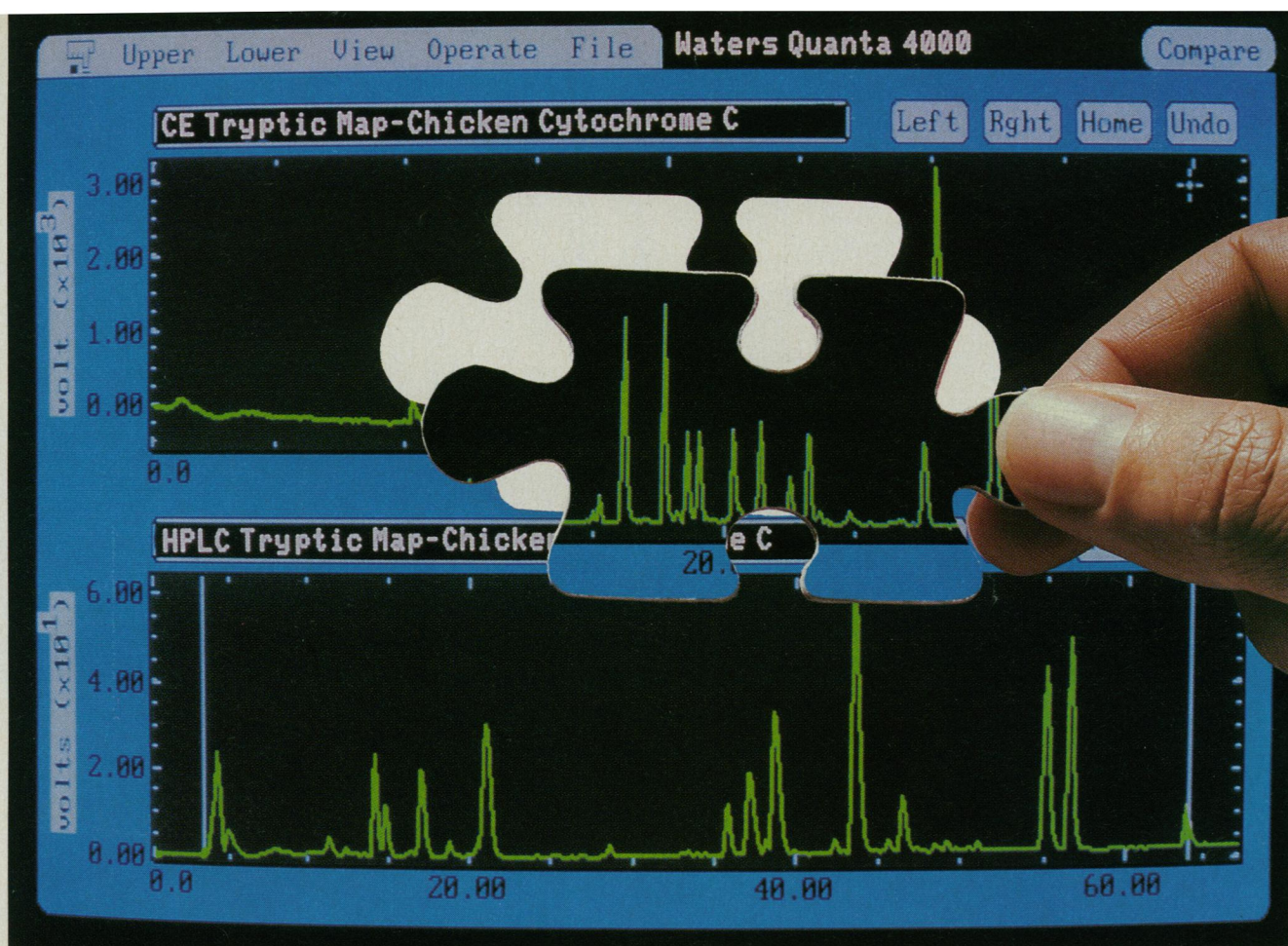
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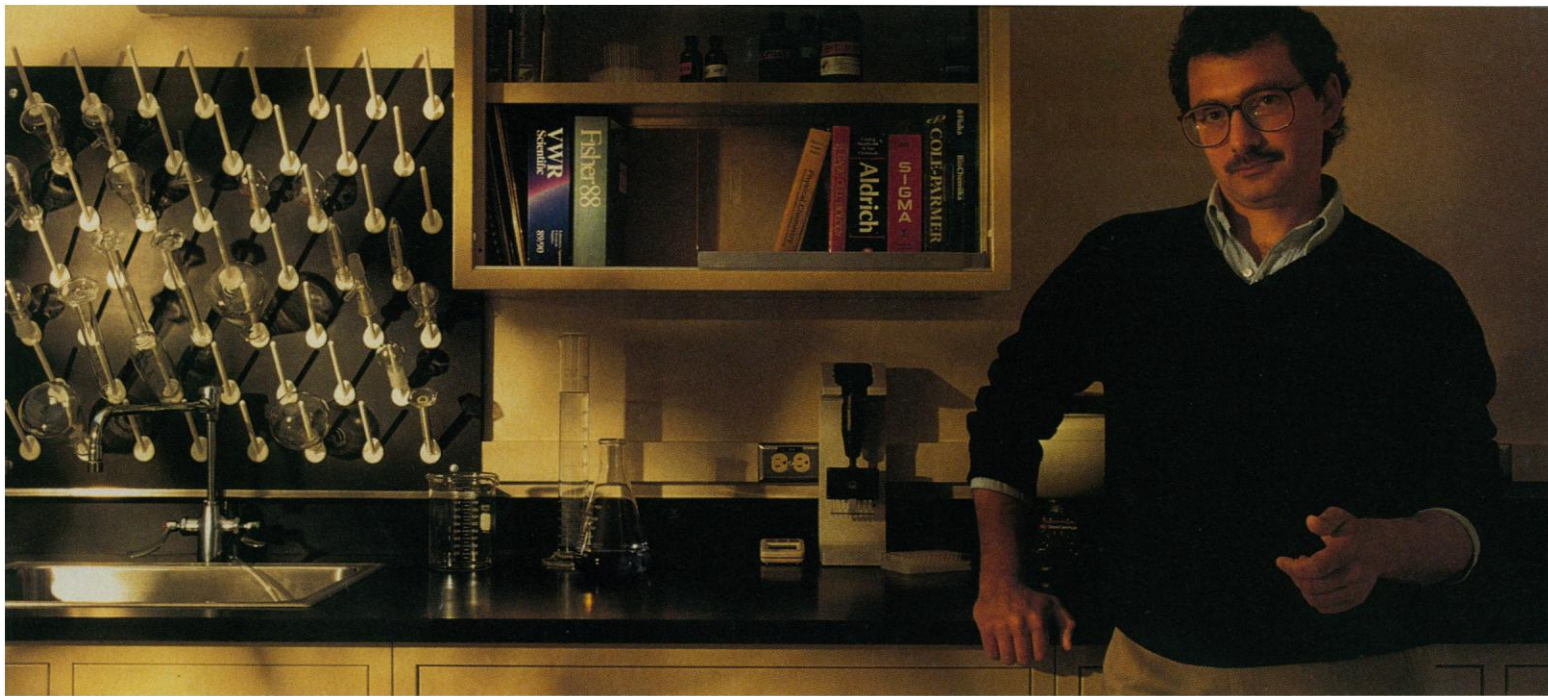
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Background

Some two years ago Bio Medic Data Systems revolutionized laboratory animal identification by introducing an implantable micro-identification device with an encoded number. An interrogation system activates the implantable chip which then transmits its number. In effect: a truly foolproof system akin to adding a unique electronic "universal product code" to each animal.

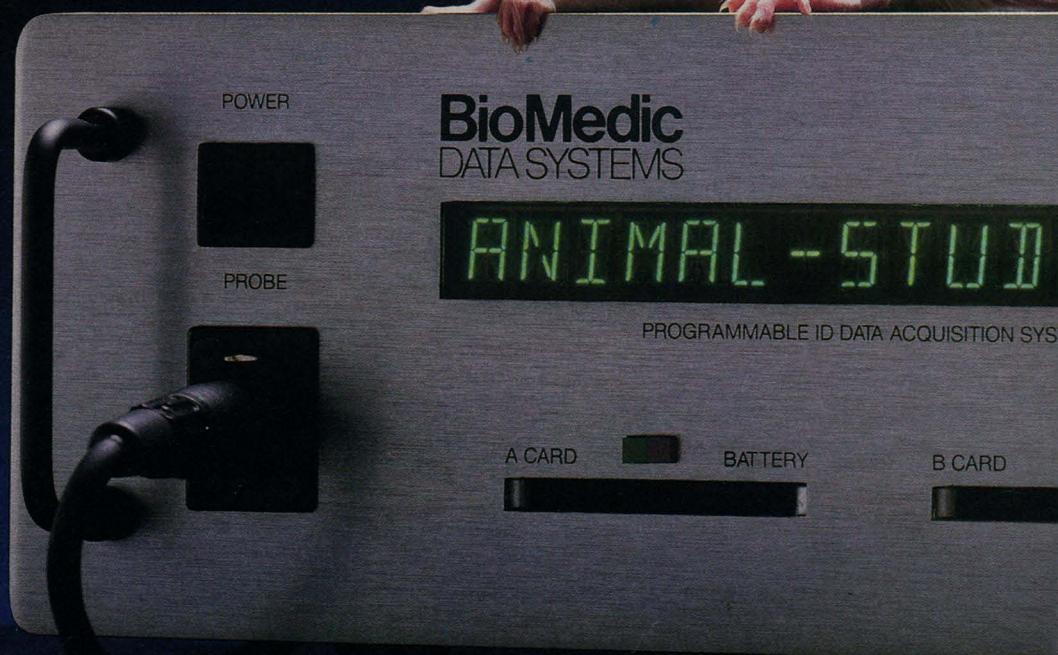
What are the Benefits?

This simple system obsoletes the traditional ear punching or tagging, toe clipping, and tail tattooing. As such, the age-old labor intensive techniques—in terms of the initial identification, the subsequent reading, and the inevitable re-dos—are replaced by a simple, easy, humane and remarkably efficient system. (A dramatic example: 200 animals can be identified in about 45 minutes.)

In addition: the imprecision of the conventional methods is replaced by *positive animal identification*. Animal misidentification or infection can indeed be catastrophic should they delay, impede, or destroy a crucial investigation. This simple foolproof system now converts ear punching or tagging, toe clipping, and tail tattooing into unacceptable risks... and who needs that when a positive animal identification system is now available!



Microchip implant
shown at 8X
magnification



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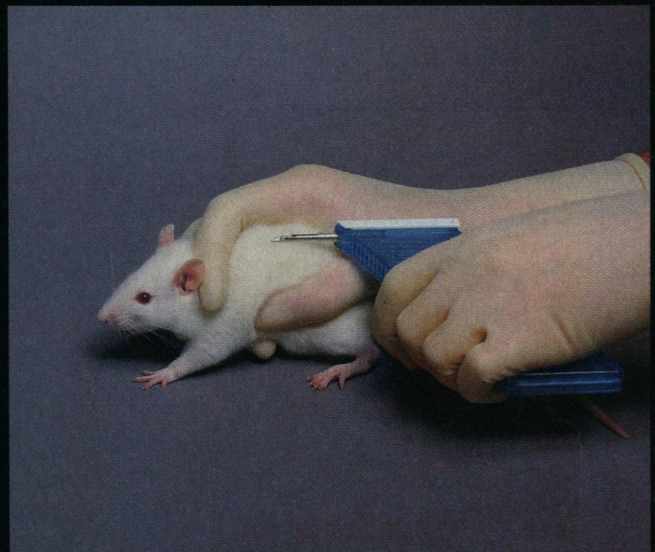
- no significant effects on normal body weights.
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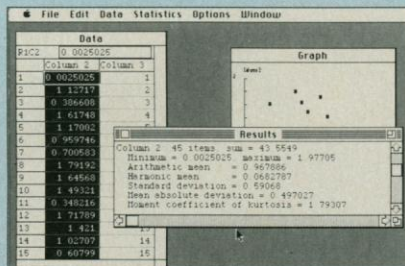
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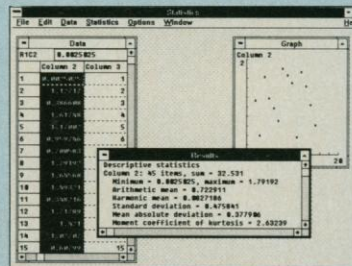


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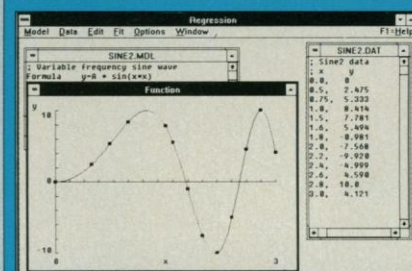
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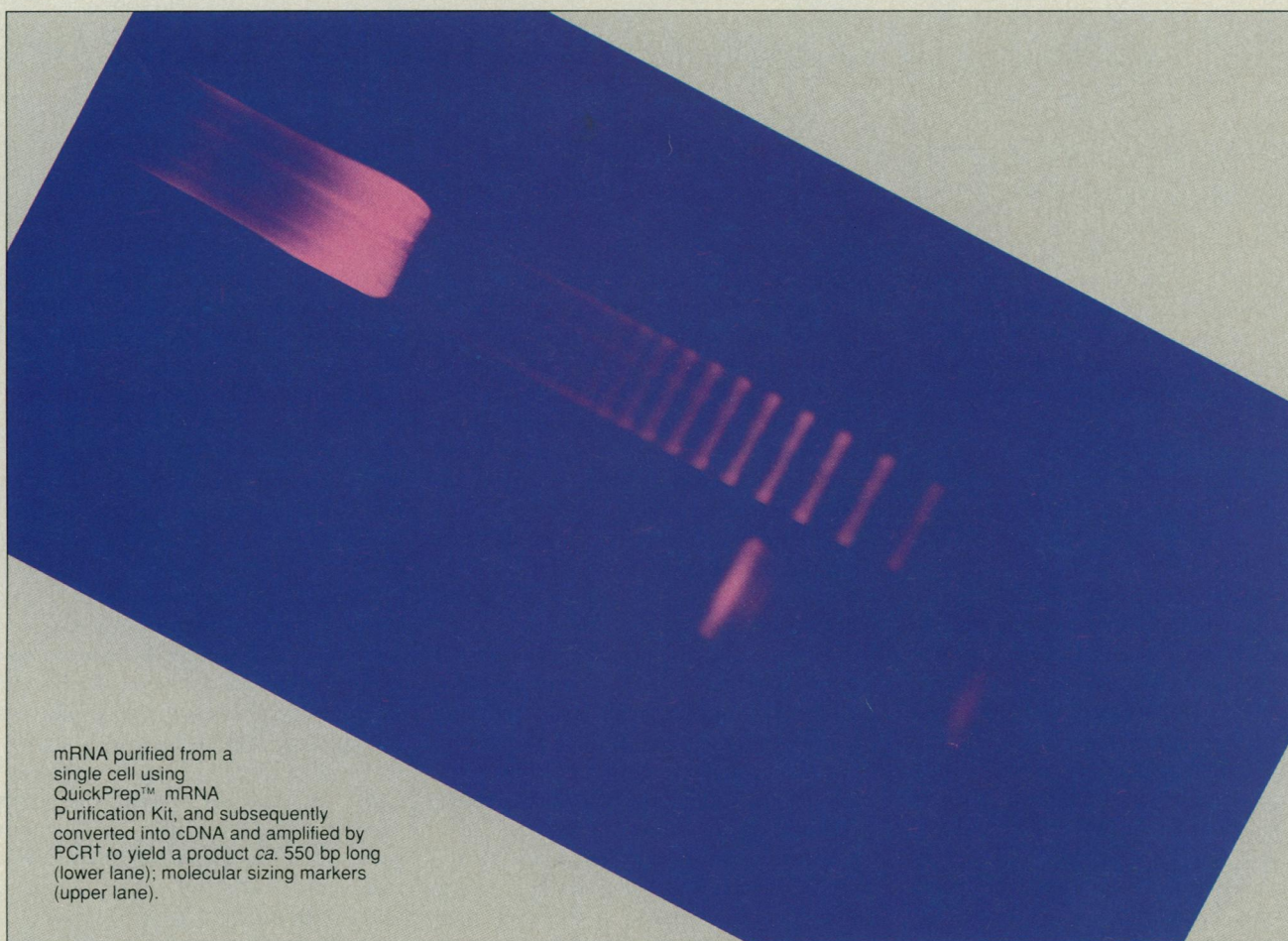
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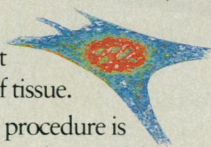
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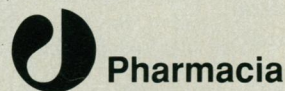
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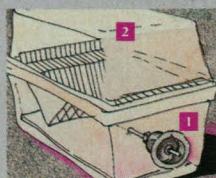
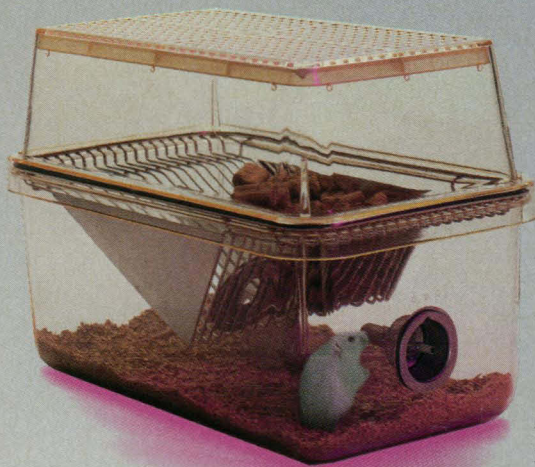
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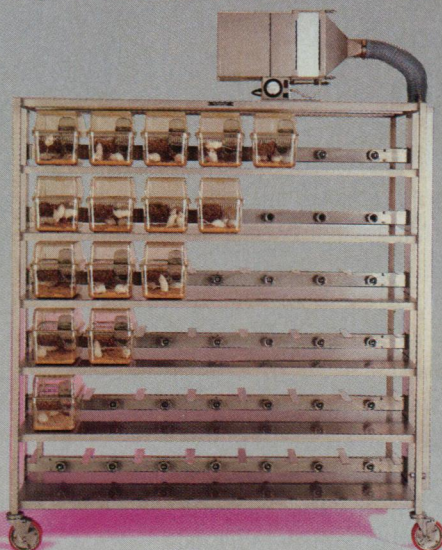


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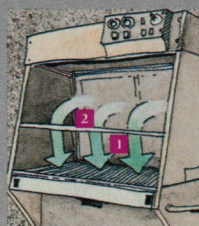
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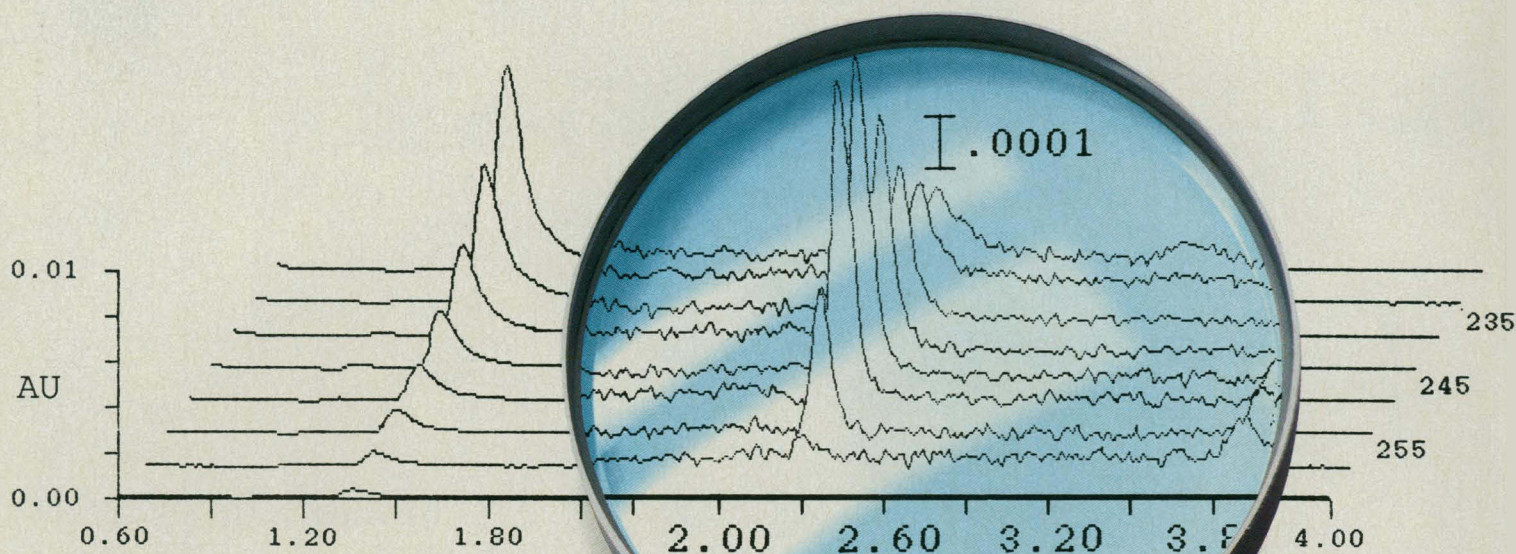
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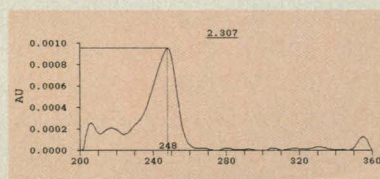
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Grants awarded are expected to be paid out before the end of June 1991.

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The Neurosciences

Challenges for the '90s

A 3-Day Seminar at the AAAS
Annual Meeting in Washington, DC

Seminar dates: 16 – 18 February 1991

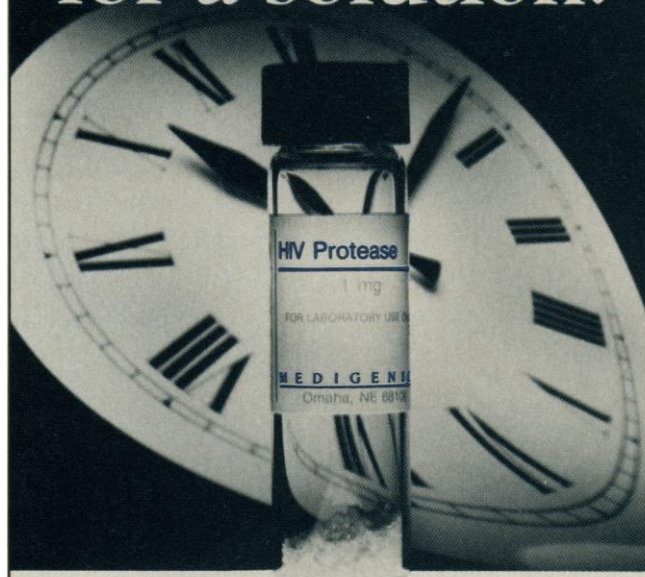
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Session topics (*presiders in parentheses*): Stimulus Transcription Coupling in Neuronal Cells (*James I. Morgan*) ♦ Structure and Function of Potassium Channels (*Arthur M. Brown*) ♦ Olfaction and Taste (*Gordon M. Shepherd*) ♦ Activity-Dependent Plasticity in Development and Learning (*Carla J. Shatz*) ♦ Cognitive Processes (*Larry R. Squire*) ♦ Molecular Basis of Neurological Disease (*Joseph B. Martin*). The plenary lecture will be delivered by Shosaku Numa of the Kyoto University Faculty of Medicine.

For a complete program and a registration form, see any of the following issues of *Science* magazine: 19 October, 26 October (insert), or 7 December; or write to AAAS Meeting Promotion Dept., Room 815, 1333 H Street, NW, Washington, DC 20005.

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