

Aromatic Interactions

The distribution of dihedral angles between two aromatic residues in globular proteins is shown in figure 1 of the Research Article by S. K. Burley and G. A. Petsko (1). It can provide evidence for the importance of specific interactions between aromatic rings only if it differs significantly from a random distribution. The expected distribution can be considered in terms of the angles between the normals to the two ring planes. Singh and Thornton (2) have pointed out that the distribution of this angle that would arise by chance varies as the sine of the angle and has a mean value of $\sim 57^\circ$. The overall distribution in figure 1 of Burley and Petsko thus closely approximates a random distribution.

However, it is also necessary to consider the spatial displacement of the two aromatic rings, as measured by polar coordinates r , T_0 , and T_ϕ between ring centers [figure 2 in (2)]. A parallel, fully stacked arrangement ($P = 0$) can only occur for T_0 approaching 90° . When the P distribution is plotted for different ranges of this angle [figure 4 in (2)], two arrangements show significant deviations from random $62.5^\circ < T_0 < 90^\circ$ (the fully stacked arrangement) and

$45^\circ < T_0 < 67.5^\circ$ (the moderately stacked arrangement). In the fully stacked arrangement the perpendicular interaction is clearly preferred and is in agreement with recent quantum mechanical calculations that show that the perpendicular interaction is energetically preferred. In the partially stacked arrangement the deviation from random appears to favor values of $P < 60^\circ$, which is also consistent with quantum mechanically derived nonbonded potential calculations.

The cutoff criteria also need to be carefully defined in the analysis of aromatic interactions. Center-to-center distances differ for rings that are in van der Waals contact above one another and those that are side by side (3.4 to 6.8 Å). It is preferable for such non-spherical side chains to define contacting aromatic groups by closest atom-atom distances [allowing for probable errors (< 1 Å) in the x-ray analyses] in order to arrive at an equal probability of contacts around the rings.

In conclusion, we wish to emphasize the importance of taking into consideration the available three-dimensional space and expected "random" distributions for such side chain interactions. A striking preference for

perpendicular packing of aromatic rings is observed for a small subgroup in a special spatial displacement. Elsewhere energetic preferences and constraints imposed by packing of many hydrophobic residues in the protein core may give a slight preference for either the perpendicular or the parallel arrangement.

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Brain "Identifier Sequence"

In their report "Brain 'identifier sequence' is not restricted to brain: Similar abundance in nuclear RNA of other organs" (1), Owens *et al.* incorrectly assert (their reference 11) that the "identifier" (ID) family of rat repeated sequences (2) is completely divergent over the more 3' repeated nucleotides from the 4D-12 rat located sequence (3). We reported that the 4D-12 sequence is

a typical version of the ID sequences published by others (2, 4) and that transcripts of it were abundant in the nuclei of various rat cells, including those from liver and kidney (3). We also pointed out (3) that the ubiquitous nuclear distribution of 4D-12 (ID) transcripts contrasted with the finding of ID transcripts only in brain cytoplasm (2).

Figure 1 shows the alignment between

the ID sequence in the p2A120 clone (2) used by Owens *et al.* (1) and the 4D-12 sequence (2), as well as two other members of the ID family (4). The sequence labeled B is cited by Owens *et al.* as a typical ID sequence (their reference 4). Although these sequences exhibit differences in the lengths and composition of their A-rich right ends, this is a typical feature of repeated interspersed mammalian DNA families (5) such as the ID family, and no region of "complete divergence" among these ID sequences is evident.

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Fig. 1. Alignment among rat ID sequences. Only the ID portion of the p2A120 clone [without its 3' oligo C track used for cloning in the Pst I site of the vector (2)] and of the DNA sequences reported in (4)

(L and B) is shown. All of the sequence of the 4D-12 repeat is shown, but without its 5' G- or 3' C-tails that were used for cloning this sequence (3). The arrow indicates the beginning of the A-rich region, which is typically at the right end of members of interspersed repeated DNA families and which varies both in length and composition (5). The dots indicate where the sequences are identical; the dashes or letters indicate where they are not.

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p2A120 GGGGCTGGGGATTAGCTCAGTGCTAGAGCGCTTACCTAGGAAGCACAAG 50
4D-12  A.....G.....C.....TG.....
L      .....G.....G.....
B      .....G.....G.....

p2A120 GCCCTGGGTTTCGG-TCCCCAGCTCCGAAAAA..... 100
4D-12  .....GA.....AT.AC.GG.....G.CC.....AAAA
L      .....G.....G.....G.CC.....AAAA
B      .....G.....G.....G.CC.....AAAA

L      AAAAAAAAAA
B      AAAAAACCAAAACAAAAACA
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