

approximate 30 percent relaxation. The muscle bath medium was changed and serotonin ( $4.5 \times 10^{-8}M$ ) and PDGF ( $33 \times 10^{-12}M$ ) were added. After contraction reached maximal force, acetylcholine ( $5.5 \times 10^{-7}M$ ) was again added and again induced a similar 30 percent decrease in tension.

13. Rat aortic strips were contracted by  $EC_{50}$  doses of either serotonin or  $PGF_{2\alpha}$ , which resulted in approximately 400-mg contractile tension. The muscle bath medium was changed, and after relaxation, an  $EC_{50}$  dose of the above agonists was added simultaneously with PDGF ( $33 \times 10^{-12}M$ ). At maximal contraction, tension generation was approximately 460 mg of force, which represents summation of the effects of both agonists (Figs. 1 and 2).
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24. Partially purified TGF- $\beta$  (prepared from human

sources with estimated 80 percent purity) and cationic FGF (prepared from bovine pituitary with estimated 5 percent purity) were obtained from Biomedical Technologies Inc., Cambridge, MA. Helical strips of rat aorta were contracted as described in Fig. 1. Experimental values are mean  $\pm$  SEM of four observations.  $EC_{50}$  values were estimated as described in Fig. 2. Based on a value of 5 percent for the purity of the cationic FGF fraction, the  $EC_{50}$  for FGF contraction would be 134 pg/ml, which is close to the  $EC_{50}$  value of 60 pg/ml reported for mitosis in VSMC in response to FGF [F. Esch *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **82**, 6507 (1985)].

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## Extended Oxygen Delivery from the Nerve Hemoglobin of *Tellina alternata* (Bivalvia)

DAVID W. KRAUS AND JAMES M. COLACINO

An oxygen-binding hemoglobin localized in the nerves of *Tellina alternata* (Bivalvia) required 30 minutes to unload oxygen when excised nerves were exposed to pure nitrogen. Neural excitability under these conditions could be sustained only until deoxygenation of the hemoglobin was complete. When the oxygen-combining function of the hemoglobin was abolished with carbon monoxide, the neural excitability ceased within a few minutes of oxygen removal, a response identical to that of hemoglobinless homologous nerves of other bivalves. These results demonstrate that aerobic activity can be supported by the oxygen stored on hemoglobin in microscopic tissues for a considerable time under anoxic conditions.

IF PIGMENTS CARRYING  $O_2$  FUNCTION mainly to increase the availability of  $O_2$  to respiring mitochondria, the molecular characteristics of the pigment should be matched to the magnitude and duration of  $O_2$  delivery required by the tissue. As an example, myoglobin rapidly dissociates  $O_2$  (1), exhibits high intracellular mobility (2), and has an  $O_2$  affinity intermediate between those of hemoglobin and cytochrome oxidase (3). These properties enable myoglobin to facilitate  $O_2$  diffusion during steady-state  $O_2$  consumption (4) and to function as a short-term  $O_2$  store (for a few seconds) at the onset of muscular contraction (5). Hemoglobins that slowly dissociate  $O_2$  could not function in facilitated  $O_2$  diffusion (6) or short-term storage. Instead, they could function as a long-term supply of  $O_2$  if matched to a similar demand (7).

A few studies relating hemoglobin characteristics to animal behavior have indirectly demonstrated hemoglobin participation in

extended  $O_2$  supply (8). On the basis of  $O_2$  unloading rates from the hemoglobin, body geometry, and  $O_2$  consumption rates, we proposed that hemoglobin could be used for long-term  $O_2$  storage in a microscopic ani-

mal, *Neodasya* (Gastrotricha) (7). By using continued nervous function as evidence for the presence of  $O_2$  (9), we have found more direct evidence that the oxygen-carrying hemoglobin located in the ganglia and all major nerves of *Tellina alternata*, a deposit-feeding bivalve, is well suited to act as a long-term  $O_2$  storage center. Under anoxic conditions the propagation of externally stimulated action potentials by the nerves continued until deoxygenation of the hemoglobin was complete (~30 minutes). If the hemoglobin was made nonfunctional with CO, the action potentials ceased shortly after ambient  $O_2$  was removed (5 minutes), a response identical to that of hemoglobinless homologous nerves of a sympatric filter-feeding bivalve, *Tagelus plebeius*.

The nerve connecting the cerebral and visceral ganglia (the c-v nerve, which is typically 2 to 4 cm long, 0.012 to 0.016 cm in diameter, and surrounded by a tough, nonremovable collagenous sheath 0.0003 cm thick) was used for this study because it is relatively large and unbranched. The c-v nerve is densely packed with glial cells (red-pigmented in *T. alternata*) and slowly conducting axons (10), 0.1 to 0.3  $\mu m$  in diameter (11). Hemoglobin characteristics in the excised nerve were determined on a microspectrophotometer with a specially constructed gas slide (7). A functional hemoglo-

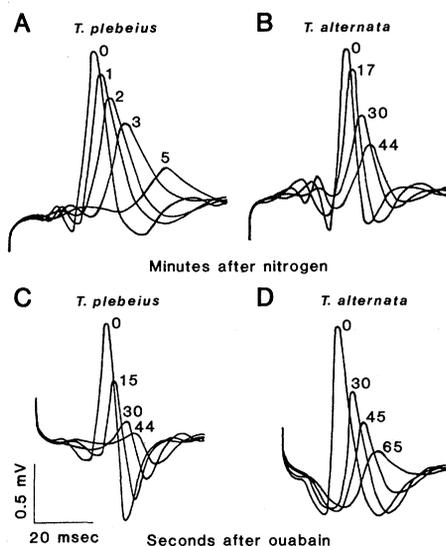


Fig. 1. Time course changes in action potential appearance under different conditions. Action potentials retraced from sequential photographs show a decrease in amplitude and conduction velocity until indistinguishable from baseline. A supramaximal stimulus of 4 to 7 V and 0.5 to 1.0 msec at 2 pulse/sec was used in (A) and (B) and 1 pulse/sec in (C) and (D); stimulus artifact is a vertical line at the left of the trace. The number at each peak indicates the time at which that trace was recorded after the treatment began.

Department of Biological Sciences, Clemson University, Clemson, SC 29634.

bin was identified through the recording of characteristic changes in absorption spectra corresponding to successive deoxygenation and reoxygenation of the tissue with humidified N<sub>2</sub> and air, respectively (12). The affinity for O<sub>2</sub> was calculated with the Hill equation from changes in fractional saturation of the hemoglobin as a function of stepwise increases in O<sub>2</sub> partial pressure (PO<sub>2</sub>). The PO<sub>2</sub> that half-saturated the hemoglobin, P<sub>50</sub>, was 1.1 ± 0.2 mmHg PO<sub>2</sub>, and the measure of heme-heme interaction, the Hill number, was 4.8 ± 0.8 (n = 8), at 20°C and pH 7.8, which indicates that the hemoglobin exhibited a high affinity for O<sub>2</sub> and a high degree of cooperativity (13).

The significance of these affinity measures becomes apparent when the PO<sub>2</sub> at which the hemoglobin begins to deliver O<sub>2</sub> is compared with the PO<sub>2</sub> at which O<sub>2</sub> supplied by diffusion becomes limited. At a PO<sub>2</sub> of 2.1 mmHg, this hemoglobin is 95 percent saturated, as computed from the Hill equation, which suggests that O<sub>2</sub> delivery will start at a PO<sub>2</sub> value slightly higher than 2.1 mmHg. The PO<sub>2</sub> value at which O<sub>2</sub> supply becomes limited by diffusion was estimated from Harvey's equation for a long cylinder (14):

$$C_o = (Ar^2)/(4D)$$

where C<sub>o</sub> is the oxygen partial pressure necessary to supply a given O<sub>2</sub> consumption rate A for a cylinder of radius r with a diffusion coefficient D. For the *T. alternata* c-v nerve, A was approximately 150 μl of O<sub>2</sub> per gram per hour, r averaged 0.007 cm, and D was conservatively estimated to be 1.1 × 10<sup>-5</sup> ml cm<sup>-2</sup> min<sup>-1</sup> (atm/cm)<sup>-1</sup> (at standard temperature and pressure, dry) (15). The value of C<sub>o</sub> is 0.0028 atm or 2.13 mmHg PO<sub>2</sub>, almost exactly the PO<sub>2</sub> at which the hemoglobin begins to unload. Continuously monitoring transmitted light intensity at a specified wavelength with an X-Y recorder or by recording absorption spectra at intervals of 3 to 5 minutes while flushing the gas slide with humidified 99.999 percent N<sub>2</sub> revealed that it took 29.8 ± 6.4 minutes (n = 9) for 99 percent of the hemoglobin to unload O<sub>2</sub> (16). Hemoglobin was oxygenated within 20 seconds of being exposed to air.

We recorded in situ electrical activity and simultaneous fractional saturation of the hemoglobin from entire c-v nerves by using another gas slide containing a miniature nerve chamber having 12 miniature platinum electrodes. Chamber gas tensions were controlled through gas ports, and temperature was controlled with internal fluid coils. A light beam was passed through the nerve to monitor heme saturation while the nerve was positioned on the electrodes for external

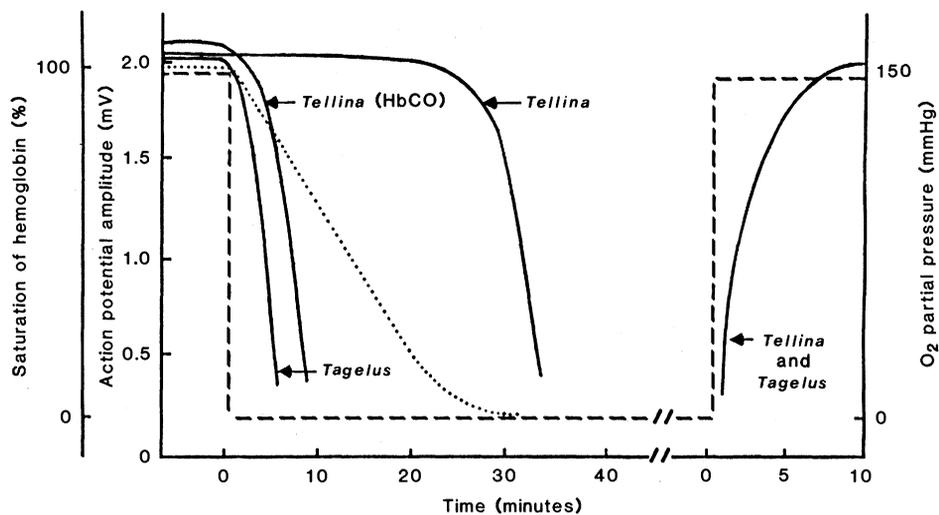


Fig. 2. Changes in action potential amplitude dependent on ambient O<sub>2</sub> or O<sub>2</sub> stored on the hemoglobin. Action potential amplitudes, solid lines; saturation of hemoglobin, dotted line; PO<sub>2</sub> in the chamber, dashed line. The action potential trace for *Tellina* is accompanied by a trace of the hemoglobin saturation for that experiment. The other two action potential amplitude traces of *Tellina* HbCO and *Tagelus* are similar because the hemoglobin was not functional or was not present. Nerve chamber volume was 0.1 ml, and gas flow was 70 ml/min, so PO<sub>2</sub> change was immediate. Carboxyhemoglobin (HbCO) was formed by exposing the nerve to 2 to 3 mmHg in air for 30 minutes, after which a carboxyhemoglobin spectrum was recorded (P<sub>50</sub> for CO = 0.4 mmHg). During this CO exposure, c-v nerves from both clams showed no change in electrical excitability.

stimulation and recording of action potentials. Action potentials from *T. alternata* c-v nerves were monitored with a Tektronix 5103N oscilloscope, photographed with a Coleman Oscillotron, and compared with the action potentials from the hemoglobin-less homologous c-v nerves from *T. plebeius*. In air, externally recorded action potentials from c-v nerves of both animals were 2 to 3 mV in amplitude, were propagated at 20 to 40 cm/sec, and exhibited refractory periods of 100 to 150 seconds at 20°C. These values agree with those of action potentials recorded from slow fibers in other bivalves (10). Freshly extracted nerves from both clams propagated action potentials at 1 to 3 pulses per second for more than 2 hours in humidified air.

Representative action potentials recorded sequentially during four experiments when the nerves were exposed to either pure N<sub>2</sub> or ouabain are overlaid and presented in Fig. 1. When the nerve chamber was flushed with N<sub>2</sub>, action potential amplitude in c-v nerves from *T. plebeius* immediately began to decline, and propagation ceased in 7.0 ± 3.7 minutes (n = 8) (Fig. 1A). In contrast, action potentials in the *T. alternata* nerve were propagated for 36.3 ± 9.6 minutes (n = 6) after N<sub>2</sub> flushing began. Amplitude did not decline by more than 10 percent for at least 15 to 20 minutes (Fig. 1B). When stimulated at 1 pulse/sec in air with a drop of 1 mM ouabain in seawater placed on the nerve between the stimulating and recording electrodes, the ability to propagate action po-

tentials beyond that point ceased in ~60 seconds in both nerves (Fig. 1, C and D). These data indicate that the nerves from both clams depend on operational Na<sup>+</sup>-K<sup>+</sup> pumps to maintain membrane potentials and that, under anoxic conditions, *T. alternata* nerves can remain active five to six times as long as those from *T. plebeius*. Figure 2 illustrates changes in action potential amplitude as dependent on either ambient O<sub>2</sub> or O<sub>2</sub> stored on the hemoglobin. When the nerve chamber was flushed with N<sub>2</sub>, the action potentials from *T. plebeius* ceased in about 5 minutes, but those from *T. alternata* ceased only after the hemoglobin was deoxygenated. However, when more than 95 percent of the hemoglobin was bound with CO to eliminate the O<sub>2</sub>-binding capability, and then the nerve chamber was flushed with N<sub>2</sub>, action potentials ceased in 5 to 10 minutes, a response similar to that of the *T. plebeius* c-v nerve under anoxic conditions. When the nerves were reoxygenated, action potentials from both *T. alternata* and *T. plebeius* returned to their initial amplitudes at similar rates.

The small diameter axons in these nerves seem to depend on aerobic metabolism to maintain membrane potentials, probably as a result of the high ratio of surface area to volume (17), and the high-affinity, slowly unloading hemoglobin in the *T. alternata* c-v nerve seems to begin to release O<sub>2</sub> just when diffusion becomes insufficient, providing at least enough O<sub>2</sub> during deoxygenation to support axon function.

The criteria necessary in the design of this particular O<sub>2</sub> storage system probably include: (i) a high-affinity hemoglobin that slowly dissociates and rapidly reassociates O<sub>2</sub> to reduce the amount of free O<sub>2</sub> diffusing out of the nerve and that, combined with high cooperativity, restricts O<sub>2</sub> unloading to a very low and narrow PO<sub>2</sub> range so that the diffusion gradient is always small; (ii) diffusion distances, which are smaller from glial cells to axons than from glial cells across the collagen sheath to the outside, that favor O<sub>2</sub> consumption by axon bundles; and (iii) O<sub>2</sub> consumption rates of active nerves that are equivalent to the O<sub>2</sub> unloading rates. A measure of how much of the unloaded O<sub>2</sub> is actually consumed by the nerve as opposed to how much is lost could indicate how efficiently the hemoglobin functions in storage. Preliminary calculations based on O<sub>2</sub> unloading rates, O<sub>2</sub> consumption rates, heme concentration, and the anatomy of the nerve suggest that more than 80 percent of the unloading O<sub>2</sub> could be consumed by the nerve during high activity, demonstrating an appropriate match between hemoglobin O<sub>2</sub> supply and neural O<sub>2</sub> demand. The molecular phenomena influencing the long unloading time and a model relating the important variables to long-term O<sub>2</sub> supply by hemoglobin will be discussed elsewhere (18).

*Tellina alternata*, unlike the filter-feeding *T. plebeius*, does not inhabit a permanent burrow, because its mode of deposit feeding quickly depletes the surrounding region of food and it must continually burrow through anoxic sediment to new locations. When laterally burrowing several centimeters below the sediment surface, it does not maintain siphonal contact with the surface and therefore does not seem to have access to a source of O<sub>2</sub>. It seems reasonable that its slowly unloading neural O<sub>2</sub> stores may enable continued nervous function and burrowing activity even in anoxic sediments for an extended period of time, allowing it to reach fresh deposits.

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  11. Data from electron micrographs of c-v nerve cross sections revealed similar numbers and sizes of axons in both bivalves.
  12. The red pigment is referred to as hemoglobin because it displays a typical oxyhemoglobin spectrum (peaks: a = 575 nm; b = 540 nm) and carboxyhemoglobin spectrum. The deoxy spectrum is unusual in having two peaks (560 nm and 530 nm), which have been ascribed to a nitrogenous or sulfurous ligand binding to the ferrous heme iron when it is deoxygenated [J. B. Wittenberg and B. A. Wittenberg, *Comp. Biochem. Physiol.* **51A**, 425 (1975)].
  13. The affinity characteristics of the hemoglobin do not seem to be pH-sensitive from 6.6 to 7.8. However, because this is an in situ preparation, it is difficult to be certain that the intracellular pH is in equilibrium with the pH of the solution that bathes the intact nerve. Although this hemoglobin has been isolated as a monomer having a molecular weight of 20,000 in its oxygenated state, it displays several characteristics in addition to high cooperativity that suggest that it forms large aggregational polymers upon

- deoxygenation (D. W. Kraus and J. M. Colacino, in preparation). The existence of this property in tissue hemoglobins that deliver O<sub>2</sub> in a precise manner has been reported [R. M. G. Wells *et al.*, *J. Comp. Physiol.* **142**, 515 (1981); L. T. Tam and A. F. Riggs, *J. Biol. Chem.* **259**, 2610 (1984)].
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## Nitrogen Fixation by *Azotobacter vinelandii* Strains Having Deletions in Structural Genes for Nitrogenase

PAUL E. BISHOP, R. PREMAKUMAR, DENNIS R. DEAN, MARTY R. JACOBSON, JOHN R. CHISNELL, THOMAS M. RIZZO, JENNIFER KOPCZYNSKI

Phenotypic reversal of Nif<sup>-</sup> mutant strains to Nif<sup>+</sup> under molybdenum-deficient conditions has been cited as evidence that *Azotobacter vinelandii* possesses two nitrogen fixation systems: the conventional molybdenum-enzyme system and an alternative nitrogen-fixation system. Since explanations other than the existence of an alternative system were possible, deletion strains of *A. vinelandii* lacking the structural genes for conventional nitrogenase (*nifHDK*) were constructed. These strains were found to grow in molybdenum-deficient nitrogen-free media, reduce acetylene (at low rates), and incorporate molecular nitrogen labeled with nitrogen-15. Thus it can be concluded that the phenotypic reversal phenomenon cannot be due to altered phenotypic expression of *nif* mutations under molybdenum-deficient conditions, but is due to the existence of an alternative nitrogen-fixation system in *A. vinelandii* as originally proposed.

SEVERAL YEARS AGO WE REPORTED that Nif<sup>-</sup> mutant strains of *Azotobacter vinelandii* containing mutational lesions in the structural genes for nitrogenase underwent phenotypic reversal to Nif<sup>+</sup> when cultured in molybdenum-deficient nitrogen-free media (1, 2). To account for this phenotypic reversal, we proposed the existence of an alternative N<sub>2</sub> fixation system expressed under conditions of molybdenum starvation and repressed in the presence of molybdenum (1). It can be argued, however, that phenotypic reversal of Nif<sup>-</sup> mutants under conditions of molybdenum starvation is due to increased leakiness of the mutant phenotypes and not to derepression of an alternative N<sub>2</sub>-fixation system. Mutant

strains containing deletions in the structural genes for nitrogenase (*nifHDK*) should distinguish between these two explanations since mutational alterations caused by deletions should preclude leaky mutant phenotypes. Here we report N<sub>2</sub> fixation by Nif<sup>-</sup> mutant strains containing deletions in *nifHDK*.

Deletions in *nifHDK* were constructed by removing internal restriction fragments from cloned DNA's containing these genes.

P. E. Bishop, R. Premakumar, M. R. Jacobson, J. R. Chisnell, T. M. Rizzo, J. Kopczynski, U.S. Department of Agriculture, Agricultural Research Service, and Department of Microbiology, North Carolina State University, Raleigh, NC 27695.  
D. R. Dean, Department of Anaerobic Microbiology, Virginia Polytechnic Institute, Blacksburg, VA 24061.