

8. CAMREX stands for Carbon in the Amazon River Experiment.
9. R. H. Meade, T. Dunne, J. E. Richey, U. dos Santos, E. Salati, *Science* **228**, 488 (1985).
10. E. M. Thurman and R. L. Malcolm, *Environ. Sci. Technol.* **15**, 224 (1981).
11. J. R. Ertel, J. I. Hedges, A. H. Devol, J. E. Richey, N. Ribeiro, *Limnol. Oceanogr.*, in press; J. R. Ertel, thesis, University of Washington, Seattle (1985).
12. The  $^{14}\text{C}$  activity of the sample ( $A_s$ ) was compared to the age-corrected  $^{14}\text{C}$  activity of the National Bureau of Standards (NBS) oxalic acid standard ( $A_{ox} = 0.95$  NBS activity) after normalization to a  $\delta^{13}\text{C}$  of  $-25$  per mil for the samples and  $-19$  per mil for the standard. The results are expressed as  $\Delta^{14}\text{C} = [(A_s/A_{ox}) - 1] 1000$  per mil [M. Stuiver and H. A. Polach, *Radiocarbon* **19**, 355 (1977)]. All uncertainty intervals about reported  $\Delta^{14}\text{C}$  values correspond to standard deviations based either on the distribution of repeated measurements of the same sample or on the uncertainties of individual measurements, whichever was larger. In the latter case, the standard deviation was determined from the results of the 15 to 20 half-minute data collection cycles that constituted a single measurement. Carbon-14 contamination in the AMS sample preparation and combustion procedure was tested for by identically analyzing an anthracite sample. No significant contamination (0.26 percent modern) was found. For particulars of the AMS sample preparation and measurement procedures, see G. W. Farwell, P. M. Grootes, D. D. Leach, F. H. Schmidt, M. Stuiver, *Radiocarbon* **25**, 711 (1983); G. W. Farwell, P. M. Grootes, D. D. Leach, F. H. Schmidt, *Nucl. Instrum. Methods* **233**(B5), 144 (1984).
13. R. Nydal and K. Lövsæth, *J. Geophys. Res.* **88**, 3621 (1983).
14. I. Levin *et al.*, *Radiocarbon* **27**, 1 (1985).
15. Maximum average residence times were calculated as the maximum time since the introduction of bomb carbon (30 years) divided by the minimum average fractional turnover within the carbon pool [100/(minimum percentage of bomb carbon); Table 1]. In this application, the usual assumption of a well-mixed pool may not be well met, and appreciable overlap in residence times of individual constituents of the various carbon forms may occur.
16. R. Herrera, C. F. Jordan, H. Klinge, E. Medina, *Interscencia* **3**, 223 (1978); C. F. Jordan, *Ecology* **63**, 647 (1982).
17. B. J. O'Brien, J. D. Stout, K. M. Goh, in *Flux of Organic Carbon by Rivers to the Ocean*, G. E. Likens *et al.*, Eds. (Department of Energy, Washington, DC, 1981), pp. 46-74; N. Gilet-Blein, G. Marien, J. Evin, *Radiocarbon* **22**, 919 (1980).
18. W. H. McDowell and T. Wood, *Soil Sci.* **137**, 23 (1984); C. S. Cronan and G. R. Aiken, *Geochim. Cosmochim. Acta* **49**, 1697 (1985); B. K. G. Theng, *The Formation and Properties of Clay-Polymer Complexes* (Elsevier, New York, 1979); J. I. Hedges, *Geochim. Cosmochim. Acta* **42**, 69 (1978).
19. Dissolved total organic matter containing significant levels of bomb carbon has been reported in several unpolluted U.S. rivers [E. C. Spiker and M. Rubin, *Science* **187**, 61 (1975); E. C. Spiker, in *Flux of Organic Carbon by Rivers to the Ocean*, G. E. Likens *et al.*, Eds. (Department of Energy, Washington, DC, 1981), pp. 75-108]. A  $^{14}\text{C}$  age less than 30 years also has been reported for Suwannee River fulvic acid [E. M. Thurman and R. L. Malcolm, in *Aquatic and Terrestrial Humic Materials*, R. F. Christman and E. T. Gjessing, Eds. (Ann Arbor Science, Ann Arbor, MI, 1983), pp. 1-23].
20. M. Stuiver and P. Quay, *Earth Planet. Sci. Lett.* **53**, 349 (1981).
21. We thank R. Meade, C. Cranston, B. Forsberg, L. Mertes, R. Victoria, L. Martinelli, R. Sampaio, and the captain and crew of the *L/M Amanai* for help in collecting field samples; D. Wilbur and W. Clark, who helped prepare samples for  $^{14}\text{C}$  analysis; M. Stuiver for cooperation in isotopic measurements; and E. M. Thurman for guidance concerning the DOC isolation procedure. Funded primarily by NSF grant DEB-801-7522 with supplementary support from the Brazilian Conselho Nacional de Desenvolvimento Científico e Tecnológico, the M. J. Murdock Charitable Trust, the U.S. Department of Energy, and NSF grants EAR-811-5994 and OCE-82-9294. Contribution 12 from the CAMREX Program, and contribution 1603 from the University of Washington School of Oceanography.

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## Sea-Air Partitioning of Mercury in the Equatorial Pacific Ocean

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The partitioning of gaseous mercury between the atmosphere and surface waters was determined in the equatorial Pacific Ocean. The highest concentrations of dissolved gaseous mercury occurred in cooler, nutrient-rich waters that characterize equatorial upwelling and increased biological productivity at the sea surface. The surface waters were supersaturated with respect to elemental mercury; a significant flux of elemental mercury to the atmosphere is predicted for the equatorial Pacific. When normalized to primary production on a global basis, the ocean effluxes of mercury may rival anthropogenic emissions of mercury to the atmosphere.

RECENTLY WE PRESENTED PRELIMINARY evidence for the evasion of mercury from the surface waters of the equatorial Pacific Ocean. This evidence supported model predictions of a low-latitude ocean source of mercury and suggested that the volatilization process may be mediated by marine biota (1). Here we report results confirming that biologically productive regions of the equatorial Pacific are a source of atmospheric mercury. Moreover, the principal species appears to be elemental mercury ( $\text{Hg}^0$ ). The largest dissolved quantities of  $\text{Hg}^0$  were associated with cooler temperatures and higher nutrient and chlorophyll *a* levels, suggesting that  $\text{Hg}^0$  efflux is tied to equatorial upwelling and phytoplankton activity.

Evidence for an atmospheric source of mercury in the equatorial Pacific was obtained on a research cruise of the O.S.S. *Researcher* from 8 June to 3 July 1984. The equatorial cruise track from  $155^\circ$  to  $93^\circ\text{W}$  and the track at  $85^\circ\text{W}$  from  $4^\circ 30'\text{S}$  to

$4^\circ 30'\text{N}$  are shown in Fig. 1. Seawater for measurement of dissolved gaseous mercury (DGM) was collected in the surface mixed layer at every  $5^\circ$  of longitude along the equator and at closer intervals ( $<2^\circ$ ) along the track at  $85^\circ\text{W}$ . Water samples were also

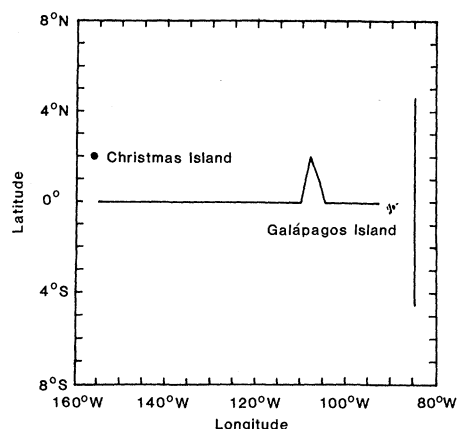


Fig. 1. Cruise track of the O.S.S. *Researcher* in the equatorial Pacific (June to July 1984).

obtained for determination of temperature, total mercury (2), nitrate plus nitrite, and chlorophyll *a*. Total gaseous mercury (TGM) and organic gaseous mercury (OGM) in the air were sampled at bow level (10 m above the surface) on both cruise legs while the ship was under way. The broad coverage of these two cruise tracks allowed quantification of the horizontal extent and range of DGM in surface water and TGM and OGM in the overlying air in the equatorial Pacific.

Samples of DGM in surface seawater were collected in acid-cleaned, Teflon-lined, 30-liter Go-Flo bottles suspended on Kevlar line and anchored with a plastic-bagged concrete weight. The seawater was then purged with mercury-free air in a 4-liter flow-through glass bubbler that had been scrupulously acid-cleaned (3). The sparging efficiency for  $\text{Hg}^0$  was 90 to 95 percent, as determined with dissolved oxygen as a tracer and a volatilization rate ratio of  $\text{Hg}^0$  to dissolved oxygen of 0.94 (4). Total DGM was purged from 30- to 60-liter aliquots, trapped on gilded quartz sand columns, and analyzed by cold vapor atomic absorption spectrophotometry (5). Another 30- to 60-liter aliquot was used to separate the total DGM into an organic and an inorganic fraction with a silver-gold stack method (6). In this arrangement the silver column collects  $\text{Hg}^0$  while passing volatile organic mercury species such as  $(\text{CH}_3)_2\text{Hg}$  to be trapped on the gold column. Subsequent measurements are made with a two-stage gold amalgamation technique (5).

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Atmospheric concentrations of TGM and OGM were determined by the gold and the silver-gold stack procedure (6). Total mercury in surface seawater was collected in the Go-Flo bottles. Aliquots were acidified with ultrapure, sub-boiling distilled nitric acid and stored in acid-cleaned, 2-liter Teflon bottles for later analysis (2). Sea surface temperature was measured by a thermistor mounted on the hull of the ship at a depth of 5 m. Temperature profiles (250 m) were obtained with expendable bathythermographs or with reversing thermometers mounted on Niskin bottles in a rosette frame. The nutrient samples of nitrate plus nitrite were collected in acid-washed (10 percent HCl) polyethylene bottles and frozen for later analysis with a Technicon autoanalyzer (7). Chlorophyll *a* was measured fluorometrically (8).

The atmospheric measurements of TGM along the equatorial cruise track (Fig. 2A) and along the meridional cruise track at 85°W (Fig. 2B) ranged from 0.85 to 1.13 ng of mercury per standard cubic meter (SCM), with an average of  $1.02 \pm 0.08$  (mean  $\pm$  SD,  $n = 23$ ). This variation corresponds to our estimate of overall experimental error associated with the collection, volume determination, and analysis of TGM at

these concentrations. The value compares well with open-ocean results for the Southern Hemisphere of  $1.05 \pm 0.09$  ng/SCM ( $n = 4$ ) at latitudes greater than 10°S during our 1980 equatorial Pacific study (1) and with  $1.07 \pm 0.08$  ng/SCM ( $n = 16$ ) measured by us in the South Pacific westerlies at New Zealand. Values of  $1.05 \pm 0.22$  ng/SCM ( $n = 64$ ) have been reported for the South Atlantic (9). The predominant form of the mercury in the air is  $Hg^0$  (80 to 96 percent), as defined by the silver-gold separation technique (Fig. 2, A and B).

Elevated atmospheric TGM (1.5 to 2.3 ng/SCM) found during our October 1980 study (1) at 160°W in the equatorial upwelling region was not observed. The differences in TGM between the cruises most probably reflect temporal and spatial variability in the processes affecting the volatilization of mercury in the equatorial zone. For example, equatorial upwelling can show significant short-term (10- to 20-day) variations in response to fluctuations in the strength of the trade winds (10). In addition, significant areal and temporal changes in equatorial primary production have been reported (11).

The total DGM in the surface seawater (Fig. 2A) generally increases along the equa-

tor from about 60 fM between 155° to 125°W to 225 fM near the Galapagos Islands (90°W). At the 85°W track (Fig. 2B), total DGM is about 50 fM at 4°30'N and gradually increases to 135 fM at 3°S. The silver-gold speciation experiments show that all the DGM is  $Hg^0$ . The increasing trend of DGM coincides with decreasing surface sea temperature and increasing concentration of nitrate plus nitrite along the equator farther east toward the Galapagos Islands (Fig. 2A) and south of the equator along the 85°W track at 3°S (Fig. 2B). Cooler, nutrient-rich waters found east of 130°W are indicators of the upwelling of water from lower depths (12). The 20°C isotherm along the equator was at a depth of 100 at 155°W and 25 m at 93°W. This shallowing of the thermocline allowed cool water to be closer to the sea surface in the eastern part of the equatorial Pacific.

The chlorophyll *a* distribution, contoured every 100  $\mu g/m^3$  for the top 100 m (Fig. 2, A and B), is variable and reflects the patchiness of the phytoplankton populations. The sampling depths for DGM are superimposed on the chlorophyll *a* distribution. High concentrations of DGM and chlorophyll *a* occur along the equator at 95°, 100°, 105°, and 115°W (Fig. 2A) and at 2° and

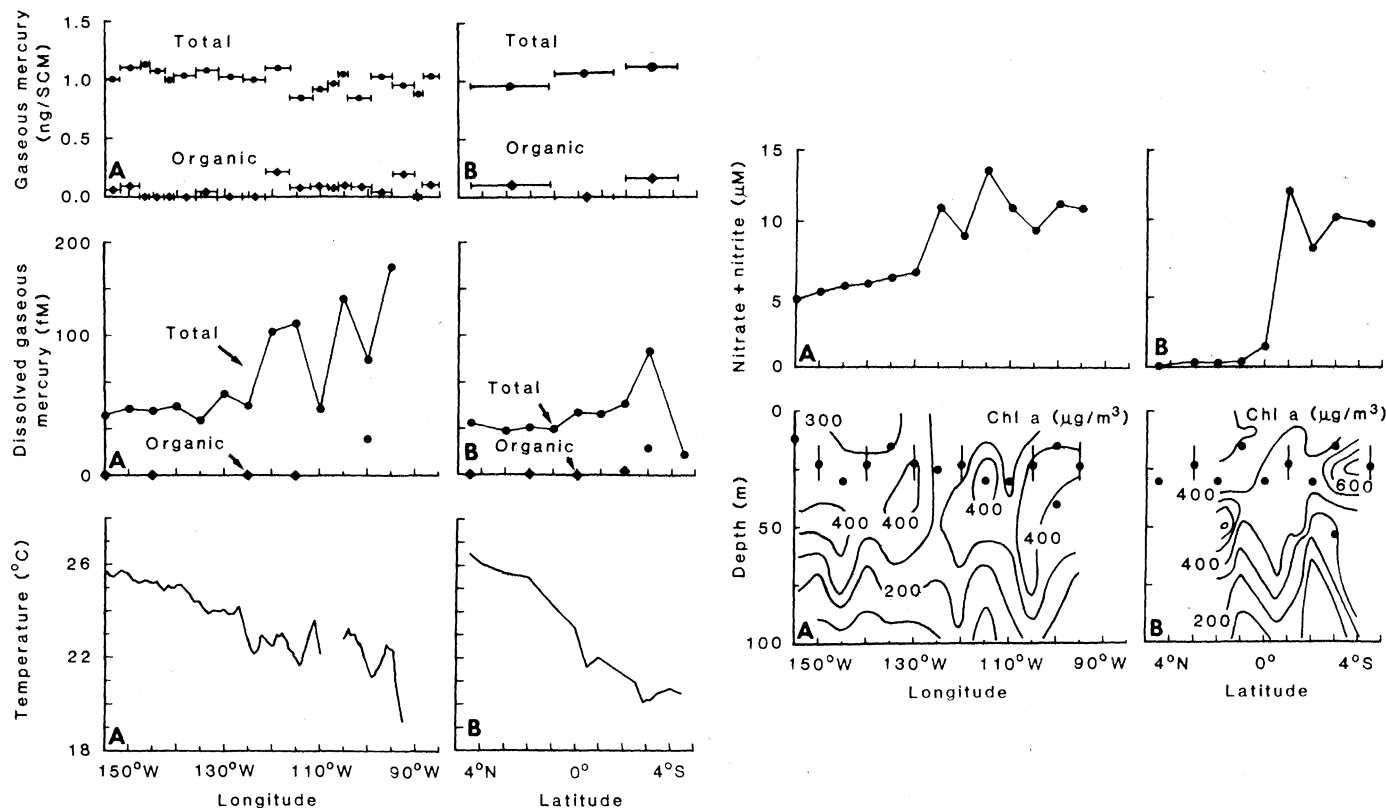


Fig. 2. Experimental results from the equatorial Pacific plotted versus longitude from 155° to 93°W along the equator (A) and versus latitude from 4°30'N to 4°30'S along 85°W (B). The parameters, in descending order, are: total and organic gaseous mercury concentrations in the atmosphere in nanograms per standard cubic meter; dissolved gaseous mercury in surface seawater in femtomoles per liter; sea-surface temperature in degrees Celsius;

nitrate plus nitrite in micromoles per liter; and chlorophyll *a*, contoured every 100  $\mu g/m^3$ . Superimposed on the chlorophyll *a* field are the sampling depths of the dissolved gaseous mercury. A circle denotes one sample, while a circle with a line indicates that two samples taken at 15 and 30 were combined for the dissolved gaseous mercury measurement.

3°S (15 m) along the 85°W track (Fig. 2B). Among all samples, DGM and chlorophyll a have a correlation coefficient of 0.448 ( $P = 0.0415$ ,  $n = 21$ ).

The correspondence between DGM and chlorophyll a, although weak, suggests that dissolved mercury in the surface waters of the equatorial Pacific may be volatilized by phytoplankton to an inorganic volatile form that is probably  $\text{Hg}^0$ . Laboratory studies have shown that algae are able to convert dissolved mercury to  $\text{Hg}^0$  (13). The elevated levels of DGM in cooler, nutrient-rich waters also suggest that DGM could be produced at depth, perhaps through bacterial activity, and subsequently be transported in upwelling waters to the sea surface. Dissolved gaseous mercury may also be formed from the reduction of dissolved mercury species by organic compounds. While the production of DGM is associated with biological activity, there is no specific information on the principal volatilization mechanisms.

Total mercury concentrations (2) in the surface water of the equatorial Pacific are low, ranging from 1.0 to 3.5 pM. These values compare well with our previous results in the equatorial Pacific (155°W) for reactive Hg in surface seawater of 0.7 to 2.0 pM (1). The concentrations of DGM range from 28 to 225 fM and represent a significant fraction of total mercury (2 to 12 percent).

Elemental mercury is the only inorganic gaseous form of mercury that is sufficiently volatile to be purged from seawater under the experimental conditions. Dissolved  $\text{HgCl}_2$  gas, although predicted to be about 2.6 percent of the total dissolved mercury in seawater (14), is too soluble to be removed during the purging step. The Henry's law constant [ $H = C_g/C_w$ , where  $C_g$  and  $C_w$  are the concentrations for  $\text{HgCl}_2$  in air and in fresh water ( $\text{Cl}^- = 0.2 \times 10^{-3}M$ ), respectively] for  $\text{HgCl}_2$  gas is  $2.9 \times 10^{-8}$  at 25°C (15). Only inorganic volatile mercury species with relatively large values of  $H$  are viable candidates for DGM, with  $\text{Hg}^0$  ( $H = 0.37$  at 25°C) being the prime candidate.

It is reasonable to assume that all the DGM in the surface water and all the inorganic gaseous mercury in the overlying air is  $\text{Hg}^0$ . The degree of saturation of mercury in the seawater relative to air can be calculated by

$$S = \left[ \left( \frac{C_w H}{C_g} \right) - 1 \right] 100 \quad (1)$$

where  $C_w$  and  $C_g$  are the concentrations of  $\text{Hg}^0$  in seawater and air, respectively, and  $H$  is the Henry's law constant for  $\text{Hg}^0$  (16). Positive values of  $S$  indicate supersaturation

in the water while negative values indicate undersaturation. Using the measured values of  $C_w$  and  $C_g$  obtained on the cruise and values of  $H$  as a function of temperature in seawater for  $\text{Hg}^0$  (17), we calculate a range of values for  $S$  from 79 to 1669. The surface seawater is greatly supersaturated with  $\text{Hg}^0$  for all stations. This indicates that the equatorial Pacific Ocean is a source of mercury to the atmosphere.

The flux of  $\text{Hg}^0$  from the ocean to the atmosphere can be estimated with a thin-film gas-exchange model following the approach of Broecker and Peng (18):

$$F = \left( \frac{D}{Z} \right) (C_t - C_b) \quad (2)$$

where  $F$  is flux of  $\text{Hg}^0$  into (+) or out of (−) the ocean,  $D$  is diffusivity of  $\text{Hg}^0$  in water,  $Z$  is water film thickness at the air-sea interface, and  $C_t$  and  $C_b$  are the concentrations of  $\text{Hg}^0$  at the top and the bottom of the film, respectively. Using an average value of  $Z$  for this region of  $50 \pm 22 \mu\text{m}$  ( $n = 11$ ) (18) and a value of  $D$  for  $\text{Hg}^0$  (19) of  $2.9 \pm 0.1 \times 10^{-5} \text{ cm}^2/\text{sec}$  at 23.6°C [the average sea-surface temperature ( $n = 22$ ) of all the stations], we calculate a piston velocity ( $D/Z$ ) for  $\text{Hg}^0$  of  $5.1 \pm 2.2 \text{ m/day}$ . This is comparable to piston velocities of other gases such as oxygen ( $4.0 \pm 1.7 \text{ m/day}$ ,  $n = 11$ ) and radon ( $2.4 \pm 1.0 \text{ m/day}$ ,  $n = 11$ ) in the equatorial region (18).

The calculated annual flux of  $\text{Hg}^0$  ranges from 4.4 to 78.7  $\mu\text{g}/\text{m}^2$ , with an average of  $29 \pm 19 \mu\text{g}/\text{m}^2$  ( $n = 22$ ). Taking the area of the equatorial Pacific Ocean (length from 82° to 172°W and width from 5°N to 5°S) to be roughly  $1.1 \times 10^{13} \text{ m}^2$ , we estimate an average annual mercury flux from the equatorial Pacific to the atmosphere of  $0.32 \pm 0.26 \times 10^9 \text{ g/year}$  ( $n = 22$ ). This is about 4 percent of the total annual mercury flux to the atmosphere ( $8 \times 10^9 \text{ g/year}$ ) (1).

We can estimate a flux of mercury from the world ocean to the atmosphere by assuming that the volatilization of mercury is proportional to primary production. About 11 percent of the total primary production occurs in the equatorial Pacific, on the basis of a primary production in the equatorial zone of  $2 \times 10^{15} \text{ g}$  of carbon per year (20) and a primary productivity for the world ocean of  $1.8 \times 10^{16} \text{ g}$  of carbon per year (21). This yields an annual mercury efflux of  $2.9 \pm 1.8 \times 10^9 \text{ g/year}$  ( $n = 22$ ) for the world ocean. This is about 36 percent of the yearly mercury flow to the atmosphere and is comparable to anthropogenic emissions estimated by Watson (22) at  $2.4 \times 10^9 \text{ g/year}$ .

In summary, evasion is an important feature of the marine biogeochemical cycle of mercury in the equatorial Pacific ocean. On

a broader scale, oceanic effluxes of mercury may significantly affect the global cycling of this metal. The exchange behavior of mercury between air and sea should be further investigated, particularly in differing ocean regimes, to elucidate the mechanisms responsible for the volatilization of mercury in the marine environment.

#### REFERENCES AND NOTES

1. W. Fitzgerald, G. Gill, J. Kim, *Science* **224**, 597 (1984).
2. G. Gill and W. Fitzgerald, *Deep-Sea Res.* **32**, 287 (1985). Mercury measurements in seawater are often divided into two fractions, reactive and total mercury. Reactive mercury measurements are broadly defined as representing those species of mercury that are readily reducible (as with  $\text{SnCl}_2$ ) in acidified seawater. Total mercury is determined in unfiltered seawater after prolonged acid digestion (>1 month of storage at pH 1.2) that is required to cleave covalent carbon-mercury bonds to liberate inorganic mercury for reduction and detection. Operationally, the difference between total and reactive mercury is the measure of the presence of reasonably stable organo-mercury associations, such as methylated mercury compounds.
3. C. Patterson and D. Settle, *Natl. Bur. Stand. (U.S.) Spec. Publ.* **422** (1976), p. 321.
4. S. Okouchi, S. Sasaki, Y. Harano, *Bull. Chem. Soc. Jpn.* **57**, 338 (1984).
5. W. Fitzgerald and G. Gill, *Anal. Chem.* **51**, 1714 (1979).
6. R. Braman and D. Johnson, *Environ. Sci. Technol.* **8**, 991 (1974); W. Fitzgerald, G. Gill, A. Hewitt, in *Trace Metals in Sea Water*, C. Wong, E. Boyle, K. Bruland, J. Burton, E. Goldberg, Eds. (Plenum, New York, 1983), pp. 297–315.
7. P. Glibert and T. Loder, *WHOI Tech. Rep.* 77-47 (1977).
8. C. Yentsch and D. Menzel, *Deep-Sea Res.* **10**, 221 (1963); O. Holm-Hansen, C. Lorenzen, R. Holmes, J. Strickland, *Cons. Int. Explor. Mer.* **30**, 3 (1965).
9. F. Slemr, W. Seiler, G. Schuster, *J. Geophys. Res.* **86**, 1159 (1981).
10. K. Wyrki and G. Eldin, *J. Phys. Oceanogr.* **12**, 984 (1982).
11. R. Barber and F. Chavez, *Science* **222**, 1203 (1983).
12. K. Wyrki, *J. Phys. Oceanogr.* **11**, 1205 (1981).
13. D. Ben-Bassat and A. Mayer, *Physiol. Plant.* **33**, 128 (1975); *ibid.* **40**, 157 (1977); W. Macka, H. Wihlidal, J. Washutti, E. Bancher, *Experientia* **34**, 602 (1978).
14. D. Dyrssen and M. Wedborg, in *The Chemistry and Biogeochemistry of Estuaries*, E. Olsson and I. Caro, Eds. (Wiley, New York, 1980), pp. 71–120.
15. A. Iverfeldt and O. Linqvist, in *Proceedings of the International Symposium on Gas Transfer at Water Surfaces*, W. Brutsaert and G. Jirka, Eds. (Reidel, Dordrecht, 1984), pp. 533–538.
16. S. Hoyt and R. Rasmussen, in *Mapping Strategies in Chemical Oceanography*, A. Zirino, Ed. (American Chemical Society, Washington, 1985), pp. 31–56.
17. I. Sanemasa, *Bull. Chem. Soc. Jpn.* **48**, 1795 (1975).
18. W. Broecker and T.-H. Peng, *Tracers in the Sea* (Eldigio, Palisades, NY, 1982), pp. 110–131.
19. The diffusivity of  $\text{Hg}^0$  is calculated by the Othmer-Thaker correlation, which empirically relates the diffusivity of a substance to its molal volume at its boiling point [D. Othmer and M. Thaker, *Ind. Eng. Chem.* **45**, 589 (1953)].
20. F. Chavez, R. Barber, J. Kogelschatz, V. Thayer, B. Cai, *Trop. Ocean-Atmos. Newsl.* **28**, 1 (1984).
21. J. Ryther, *Science* **166**, 72 (1969).
22. W. Watson, in *The Biogeochemistry of Mercury in the Environment*, J. Nriagu, Ed. (Elsevier/North-Holland, Amsterdam, 1979), pp. 42–77.
23. We thank the captain, officers, crew, and survey technicians of the O.S.S. *Researcher* for their assistance; S. Piotrowski for the opportunity to participate on the cruise; R. Barber (NSF grant OCE 81 10 702) for generously sharing the chlorophyll a data; D. Wilson and S. DeLuca for lugging Go-Flo bottles; and T. Loder and S. Welch for analyzing nutrient samples. Supported by the National Science Foundation as part of the Sea-Air Exchange Program (SEAREX).

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