Human Dioxin-Inducible Cytochrome P₁-450: Complementary DNA and Amino Acid Sequence

Abstract. Induction of cytochrome P_1 -450 has been linked to susceptibility to certain chemically induced cancers in mouse and man. Treatment of the human cell line MCF-7 with 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) results in high levels of aryl hydrocarbon (benzo[a]pyrene) hydroxylase (P_1 -450) activity. This cell line was used to isolate a human P_1 -450 full-length complementary DNA (cDNA) clone. The cDNA is 2566 nucleotides in length, encodes a polyadenylated messenger RNA (2.8 kilobases in length), and has a continuous reading frame producing a protein with 512 residues (molecular weight, 58,151). The human P_1 -450 cDNA and protein are 63 percent and 80 percent similar to mouse P_1 -450 cDNA and protein, respectively. Whereas the mouse TCDD-inducible P-450 gene subfamily has two members (P_1 -450 and P_3 -450), the human TCDD-inducible gene subfamily appears to have only one gene (P_1 -450).

The compound 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) is a potent inducer of many proteins including drugmetabolizing enzymes such as the cytochrome P-450 proteins (1-3). In mice, cytochromes P₁-450 and P₃-450 are dramatically induced by TCDD (4, 5). P_1 -450 is closely associated with aryl hydrocarbon (benzo[a]pyrene) hydroxylase (AHH) and is responsible for the metabolic activation of polycyclic hydrocarbons such as benzo[a]pyrene to form reactive carcinogenic intermediates (6). Induction of P₁-450 by TCDD may play a role in the initiation of certain types of environmentally caused malignancies; TCDD is a cocarcinogen in a mouse tumor model system (7) and is a very potent promoter of tumorigenesis (8). Genetically determined high inducibility of AHH is associated with enhanced risk of certain types of chemically induced cancers in mice (9) and in cigarettesmoking humans (10). The AHH gene system, like the α_1 -antitrypsin gene system (11), may be important in relating morbidity and mortality to environmental chemical exposure via cigarettes or industry (12).

As a first step in developing an assay for genetic susceptibility to chemically induced cancers we determined the complementary DNA (cDNA) and amino acid sequence of human P_1 -450. When the human breast carcinoma cell line MCF-7 is treated with TCDD, AHH activity is induced at a specific activity of >20 units per milligram of protein, which is a high value for a cell line derived from nonhepatic tissue (Fig. 1). Similar kinetics of AHH induction were observed when cells were treated with ben-Polyadenylate-enzo[*a*]anthracene. riched RNA (13) was isolated from 10^9 MCF-7 cells that had been treated with 50 nM TCDD for 24 hours. Doublestranded cDNA was synthesized after reverse-transcription of RNA; the DNA was methylated with Eco RI methylase,

and inserted into the bacteriophage cloning vector λ gt11 by use of Eco RI linkers (14). Clones that hybridized to a mixture of ³²P-labeled nick-translated full-length cDNA inserts of mouse P₁-450 and P₃-450 (5) were plaque-purified, digested with Eco RI, and subcloned into the Eco RI site of pBR322. Two Eco RI inserts from the largest clone were separately purified by agarose gel electrophoresis and used to prepare a library of DNA fragments (15) in M13 mp11. Sequencing was carried out by standard M13 cloning protocols and the dideoxynucleotide sequencing method (5, 16). Each stretch of DNA was sequenced at least once on both strands and usually 5 to 20 times. A Stu I fragment of 470 base



Fig. 1. Human AHH induction kinetics in MCF-7 cells treated with 50 nM TCDD, 50 μ M benzo[a]anthracene (BzAnth), or control medium. Specific activity denotes units (picomoles of phenolic benzo[a]pyrene products formed in 1 minute at 37°C) per milligram of cellular homogenate protein (27). The origin and development of the MCF-7 line (28), growth and treatment of the cells in culture (29), and preparation of the cells for AHH assay (27, 29) have been described.

pairs straddled the one Eco RI site in the cDNA; this fragment was sequenced to ensure that there was not a second Eco RI site. Nucleotide alignment and analysis of nucleotide and protein data were examined by standard computer programs (17).

The human P₁-450 cDNA was 2566 nucleotides long and had a continuous reading frame (from nucleotide 87 to 1625) that could produce a protein of 512 residues (Figs. 2 and 3). The termination codon for both human and mouse P1-450 messenger RNA (mRNA) was UAG (U, uracil; A, adenine; G, guanine; T, thymine; C, cytosine) (Fig. 2), although only UGA has been reported as the termination codon in the more than 280 published sequences of human and mouse mRNA's. UAA is the termination codon for hamster vimentin mRNA (18), however, providing further evidence that UGA is not the only termination codon in mammals. There were several termination codons (including two UGA codons) in two of the three possible reading frames in the 25 bases after the occurrence of the human UAG codon (Fig. 2); this is seen near the translation termination site of many eukaryotic and prokaryotic genes (19).

The human and mouse cDNA nucleotide sequences (Fig. 2) had 63 percent similarity overall, with much greater similarity (83 percent) in the translating regions than in the 5' nontranslating and the 3' nontranslating regions (34 percent similarity). Of 270 base changes in the translating region (17 percent divergence), 29, 16, and 55 percent occur in the first, second, and third codon positions, respectively. The large divergence in the nontranslating regions, as compared with that in the translating regions of human and mouse P₁-450 cDNA, is consistent with the finding that the region of cDNA encoding an important protein remains much more conserved than regions of cDNA that do not (19).

There was an 80 percent similarity between the human and mouse P_1 -450 deduced amino acid sequences (Fig. 3). In contrast, the human P_1 -450 protein sequence was 68, 79, 68, 35, and 27 percent similar to mouse P₃-450, rat P-450c, rat P-450d, rat P-450e, and rat P-450scc proteins, respectively (5, 20). Our sequencing data thus indicate that we have isolated the human P-450 cDNA that is equivalent to mouse P_1 -450 and rat P-450c cDNA. The amino acid composition of human and mouse P_1 -450 proteins was strikingly similar, including the presence of nine cysteine residues in both proteins. The mouse P₃-450 protein has six cysteine residues (5). The molecular weight of the human P_1 -450 protein (58,151) is about 1080 less than that of the mouse P_1 -450 protein (59,230).

When mouse genomic DNA is probed with mouse P₁-450 or P₃-450 full-length cDNA clones, the Southern blot hybridizations are consistent with the existence of only two genes in this P-450 family (5). In addition, when liver RNA from TCDD-treated mice is probed with these two full-length clones, mRNA 2.9 kilobases (kb) and 2.1 kb in length are found (21). When a probe derived from the 5'end of human cDNA (Fig. 4) was used for DNA or RNA blot hybridizations, however, all the data were consistent with the presence of a single TCDDinducible human P-450 gene. Digestion with Xba I and Hind III resulted in fragments 5.2 and 8.8 kb in size, respectively. These bands did not appear to differ among HeLa, human placenta, and MCF-7 genomic DNA's (Fig. 4A). Digestion with Kpn I resulted in two fragments (5.9 and 2.2 kb) in all three human genomic DNA preparations; these data are consistent with a known Kpn I site in the human P_1 -450 cDNA. The same result was obtained under lower hybridization stringencies (22).

The human 5' cDNA clone hybridized to both 2.1- and 2.9-kb mRNA from 3methylcholanthrene-treated C57BL/6J inbred mice (Fig. 4B). No hybridization to untreated MCF-7 cultures was observed, but in TCDD-treated cultures, a single size of mRNA (2.8 kb) hybridized strongly to the probe. A length of 2566 nucleotides for the cDNA, plus a poly(A) region of about 250 nucleotides, would produce the human P₁-450 mRNA of about 2.8 kb. These findings suggest a species difference in this TCDD-inducible P-450 gene subfamily; man probably has only P₁-450, whereas mouse has both P₁-450 and P₃-450. The functions of human P₁-450 thus might be divided between rodent P₁-450 and P₃-450. Alternatively, new functions may have evolved in P₃-450.

From the sequence data, it has been estimated that the TCDD-inducible P-450 gene family diverged from the phenobarbital-inducible P-450 gene family more than 200 million years ago (5, 20) and that the P_1 -450 and P_3 -450 genes separated (probably via gene duplication) about 65 million years ago (5). This estimate is consistent with the finding of at least two major TCDD-inducible P-450 proteins in mouse, rat, and rabbit (3, 5). Moreover, this calculation predicts that human P-450 proteins might not exhibit this pattern, because divergence of human from rodent would have occurred prior to the split of P₃-450 from P₁-450 5 APRIL 1985

BU BU Human: TGATOCTTGTGATOCCAGGGCTCCAGGGCCCCACGCCCAGGCTCCAGGCCCCCAGGATOCCTAGGATOCCTAGGATOCCTAGGATOCCTAGGATOCCTAGGAT Mouse: CTGAAGGTGGTAGTTCTTGGAGC TC GATCCT G T AA T A T------- C CTTACAGC CAG AC --TA GTA ATC AGG GCC TCA AGA CCT CAG GTC CCC AAA GGC CTG AAG AAT CCA CCA GGG CCA TGG GGC TGG CCT CTG ATT GGG CAC ATG CTG ACC CTG GGA AAG AAC CCG CAC CTG GCA CTG TCA AGG ATG AGC CAG CAG TAT COGG GAC GTG CTG CAG ATC CGA ATT CGC TCC ACA CCC GTG GTG GTG CTG AGC GGC CTG GAC ACC ATC CGG CAG GAC CTG GTG CGG CAG GGC CAT GAT TTC AAG GGC CGG CGC GAC CTC TAC ACC TTC ACC CTC ATC AGT AAT GGT CAG AGC ATG TCC TTC AGC CCA GAC TCT GGA CCA GTG TGG GCT GCC C3G C3G C3C CTG GCC CAG AAT 600CTG ATA AGC AGG TTG CAG GAG CTG ATG GCA GGG CCC GGG CAC TTT AAC COC TAC AGG TAT GTG GTA TCA GTG ACC AAT GTC ATC C C C AAC T A C A GT C G A G C C C AAC T C G A G TGT GCC ATT TGC TTT GGC CGG CGC TAT GAC CAC AAC CAC GAA CTG CTT AGC CTA GTC AAC CTG AAT AAT TTC GGG GAG GTG GTT GOC TCT GGA AAC CCA GCT GAC TTC ATC CCT ATT CTT COC TAC CTA CCC AAC CCT TCC CTG AAT GCC TTC AAG GAC CTG AAT GAG AAG TTC TAC AGC TTC ATG CAG AAG ATG GTC AAG GAG CAC TAC AAA CC TTT GAG AAG GGC CAC ATC COG GAC ATC ACA GAC AGC CTG ATT GAG CAC TGT CAG GAG AAG CAG CTG GAT GAG AAC GOC AAT GTC CAG CTG TCA GAT GAG AAG ATC ATT AAC ATC GTC TTG GAC CTC 1050 1080 1110 TTT GGA GCT GGG TTT GAC ACA GTC ACA ACT GCT ATC TCC TGG AGC CTC ATG TAT TTG GTG ATG AAC CCC AGG GTA CAG AGA AAG ATC CAA GAG GAG CTA GAC ACA GTG ATT GCC AGG TCA CGG CGG CCC CGG CTC TCT GAC AGA TCC CAT CTG CCC TAT ATG GAG GAC TTC ATC G A GAC ACA GTG ATT AC AT A GAG CCC CGG CTC TCT GAC AGA TCC CAT CTG CCC TAT ATG GAG GAC CTC ATC CTG GAG ACC TTC CGA CAC TCT TCC TTC GTC CCC TTC ACC ATC CCC CAC AGC ACA AGA GAC ACA AGT TTG AAA GGC TTT TAC ATC 1380 1440 1440 C38 TIT CTC ACC CCT GAT GGT GCT ATC GAC AAG GTG TTA AGT GAG AAG GTG ATT ATC TIT GCC ATG GGC AAG C3G AAG TG ATC GGT A C AGC C A C G C C C T T A C A 1470 GAG ACC GTT GCC CCC TGG GAA GTC TTT CTC TTC CTG CCTA CCC GTG CAA CCG GTG GAA TTC AGC GTG CCA CTG GAC GTG AAG GTG A A A A T AG T T CA G A 1560 GAÇ ATG ACC CAT CTAT GGG CTA ACC ATG AAG CAT GOC TGC TGC GAG CAC TTC CAA ATG CAG CTG CGC CTC TCC GAC ATG CAC CTC CAA ATG CAG CTG CGC CTC CAA ATG CAG CTG CGC CTC CAA ATG CAG CTG CGC CTC CAA ATG CAG CTG CTC CC CAT TCC 1650 1710 TGA380CTAGACTCTGTCTACCT63TCT63TCT63TCT63CA60CA60CA60CA60CC63CCTATGGTCTAAGATTCA TT CTGTGAT A CTT A60CA6AAACCA6ATCT636CATTGT60CT CCT 1980 2010 2040 СТОЗВОССАБТАССТАВСТСТБГАВОЗБТОСТБАСТВОСТГАВОСААСТАВОСААВОССАБТСАТТГВАСААТТВОВАВСТ------АВОА АТЕГ СТВЕТТАСТТГВАСАА Т А С СА АА ВОЗВОВАСА САВСТТА ТТСТ Б А А АВЭТЕСТС С СС СТВАВОСТААССА -6004 2070 2100 2130 2160 AGAGTGAAGGGAAGGACAGCCAGGATACTGGCACAGAGGTAGTCTCACTGCTTGAACTAGGCTGAGCAATCTGACCCTATGGGTCTAGGACACAGTTCCTGGGAACATCT TCCCAA T TTTAT CTG T 1G3GG CTCT TCA C TACATAGA A ACAG GTGCTC T T GCAT C T TTT TC AG AAGGGT GA AG AAGOGT GAGAC 2190 2220 2250 -ACATTOCTCTGCCCTTOCTGCAGGCAGGAACAAACAGGGCTGCCTTCTGGCCTTGTAAGACCCTTATTGCTGTOCTGGAGGGGCTGGGGGACTT A GAGGGAGAGAT CT GGCAGTAA C GGTCTAGA TAAAGGAATGACT TT GA AG G CT GTA COCA ACTCTGA TTGTGTCTAGAAAGAOOCAAGA GAOOGAGAGAT CT 2280 2310 2340 2370 GTGTCTG003039ATCAGAG003CACA609AGT0CACATATCCAG90CACA603ACTA6099CT03AGTGA6039999GGTATTTCAATTACCTTCTATTGGTCT00CTTCTCTACACTC ACT GCTACCAA GG GC TTGG CTCCT TTTGT G TGTT TT TTACTACCTTA CAAGTGCTA ATACAGTCA AG C TGCA CA6CTCT GGTC T TT A

2400 2430 2460 2490 TTGTAATAAAATGTCTATTTTTAATGTTTGTACACAACAATCCTTC-TATTCTAQCCTGCATTGAQCTTGCATCGCATAGAQCCTTAAGAACCATTGATTTAATGTAATAGC C TA TGT T C AG AA G T GTG TA GAG TG A GA G T TT ATGCAT T------------AGT A CCTTA

2520 2550 GAAAATTCTAACCCAGGTATCCAAAAATGTGTGAAGAACAACTACCTGAGCTACTAACGAATTGTTCAGA* AG G TAG TAATTTGC AGTGT CAAAG TTCTAACT AAGTA TC G T ATAGA A ACOCTACCTGAACTACTAACTAACTAACTAGAA

Fig. 2. Comparison of human and mouse P_1 -450 cDNA sequences. The initiation codons, the cysteine codons believed to participate in the enzyme active-sites (5, 20), the termination codons, and the putative poly(A) addition signal (AATAAA) are enclosed in blackened boxes. The human sequence is numbered; the mouse sequence is shown only at those positions where there was no correlation.

(23). The data in Fig. 4 support this prediction.

The time required for 1 percent divergence in amino acid sequence [unit evolutionary period (UEP) (24)] was estimated to be 2.1 and 2.4 million years by means of comparisons between rabbit and rat P-450 proteins (25) and between mouse and rat P-450 proteins (5), respectively. If human predecessors separated from the rodent line approximately 80 million years ago (24), the human and mouse P₁-450 proteins would have diverged between 33 and 38 percent. We were surprised to find only 20 percent divergence (Fig. 3). These results suggest the existence of an additional force or forces conserving the P₁-450 protein in these two species. The metabolic activation of foreign chemicals or detoxification may contribute to this conservation. In any event, there is greater similarity between human and mouse P1-450 proteins (80 percent) than between the mouse P₁-450 and P₃-450 proteins (73 percent); P₁-450 and P₃-450 are in the same TCDD-inducible P-450 subfamily.

Knowledge of the cDNA sequence should make it possible to look for restriction-fragment length polymorphisms

	20 MLFPISMSAT EFLLASVIFC MYGLAFV LVTV	
60 HMLTLGKNPH V	80 LALSRMSQQY GDVLQIRIGS S T L	
120 TFTLISNGQS S T K	140 MSFSPDSGPV WAARRRLAQN T N	160 GLKSFSIASD PASSTSCYLE EHVSKEAEVL A TAS NY
180 ISTLQELMAG VKKVE	200 PGHFNPYRYV VVSVTNVICA V D K L A	ICFGRRYDHN HQELLSLVNL NNNFGEVVGS
240 GNPADFIPIL Y V	260 RYLPNPSLNA FKDLNEKFYS S D D	
300 EKQLDENANV DRK	320 QLSDEKIINI VLDLFGAGFD DVT	340 TVTTAISWSL MYLVMNPRVQ RKIQEELDTV T .
360 IGRSRRPRLS DQ	380 DRSHLPYMEA FILETFRHSS PQ L	
4 2 0 QWQ 1 N H D Q K L V R E	440 WVNPSEFLPE RFLTPDGAID GDN R STL	
480 LAILLORVEF QI	500 SVPLGVKVDM TPIYGLTMKH K SP E T L	

Fig. 3. Comparison of human and mouse P1-450 protein sequences. The conserved COOHterminal cysteinyl fragment (5, 20) is enclosed in a blackened box. Residues of mouse that do not match those of human are shown (30). Abbreviations for amino acids: A, alanine; C, cysteine; D, aspartic acid; E, glutamic acid; F, phenylalanine; G, glycine; H, histidine; I, isoleucine; K, lysine; L, leucine; M, methionine; N, asparagine; P, proline; Q, glutamine; R, arginine; S, serine; T, threonine; V, valine; W, tryptophan; Y, tyrosine.

Fig. 4. Hybridization analyses with the human P₁-450 cDNA 5' phP₁450-5'. probe This probe is the Eco RI fragment from base 1 to base 1521 (Fig. 2). (A) Southern hybridization. blot Conditions were sufficiently stringent and exposure of the filter to the x-ray film was short enough to exclude almost completely the cross-hybridization of the human probe with C57BL/6J mouse DNA (lanes 1, 5, and 9). (B) Northern blot Pohybridization. ly(A)-enriched RNA



was prepared from the liver of C57BL/6J inbred mice that had received 3-methylcholanthrene (MeChol) (200 mg per kilogram of body weight) 18 hours before killing and from MCF-7 cells that had been exposed to control medium or 50 nM TCDD for 24 hours. Hybridization conditions and nick-translation of the probe were as described (5).

(RFLP's), which are probes for diagnosis and prediction in a growing number of clinical disorders and human family studies (11, 26). As has been shown in mice (9), the human population exhibits a wide range of AHH (P₁-450) inducibility (10). The finding of human RFLP's representing high and low AHH inducibility would be of importance in predicting the degree of risk for persons exposed to various environmental pollutants.

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References and Notes

- References and Notes
 A. Poland and J. C. Knutson, Annu. Rev. Pharmacol. Toxicol. 22, 517 (1982).
 D. W. Nebert, M. Negishi, M. A. Lang, L. M. Hjelmeland, H. J. Eisen, Adv. Genet. 21, 1 (1982); D. W. Nebert et al., Annu. Rev. Pharmacol. Toxicol. 21, 431 (1981).
 "Cytochrome P-450" refers to an unknown number of reduced nicotinamide adenine dinucleotide phosphate (NADPH)- and NADH-dependent microsomal and mitochondrial (multisubstrate) monooxygenases, which are hemoproteins with cysteinyl sulfur bound to iron. E. Hietanen, M. Laitinen, O. Hänninen, Eds., Cytochrome P-450, Biochemistry, Biophysics and Environmental Implications (Elsevier/North-Holland Biomedical Press, Amsterdam, 1982); tochrome P.450, Biochemistry, Biophysics and Environmental Implications (Elsevier/North-Holland Biomedical Press, Amsterdam, 1982); J. Rydström, J. Montelius, M. Bengtsson, Eds., Extrahepatic Drug Metabolism and Chemical Carcinogenesis (Elsevier/North-Holland Bio-medical Press, Amsterdam, 1983). Mouse "P,-450" and "P₃-450" are those forms of TCDD-inducible P-450 in the C57BL/6N inbred mouse that are most closely associated with ary lbydrothat are most closely associated with aryl hydro-carbon (benzo[a]pyrene) hydroxylase (AHH) carbon (benzo[a]pyrene) hydroxylase (AHH) and acetanilide 4-hydroxylase activity, respec-tively. Mouse P₁-450 and P₃-450 genes corre-spond to rat P-450c and P-450d, respectively [L. M. Reik, W. L. Levin, D. E. Ryan, P. E. Thomas, J. Biol. Chem. 257, 3950 (1982)], and rabbit forms 6 and 4, respectively [D. A. Hau-gen, T. A. Van der Hoeven, M. J. Coon, *ibid.* 250, 3567 (1975); R. L. Norman, E. F. Johnson and U. Muller-Eberhard, *ibid.* 253, 8640 (1978)]. The molecular biology of these genes has recent-The molecular biology of these genes has recent-ly been reviewed [D. W. Nebert and M. Negishi,
- ly been reviewed [D. W. Nebert and M. Negishi, Biochem. Pharmacol. 31, 2311 (1982)].
 M. Negishi and D. W. Nebert, J. Biol. Chem. 254, 11015 (1979).
 F. J. Gonzalez, P. I. Mackenzie, S. Kimura, D. W. Nebert, Gene 29, 281 (1984); S. Kimura, F. J. Gonzalez, D. W. Nebert, Nucleic Acids Res. 12, 2917 (1984); S. Kimura, F. J. Gonzalez, D. W. Nebert, J. Biol. Chem. 259, 10705 (1984) 1984)
- (1964). O. Pelkonen and D. W. Nebert, *Pharmacol. Rev.* 34, 189 (1982); H. V. Gelboin, *Physiol. Rev.* 60, 1107 (1980). 6.
- Rev. 36, 189 (1982); H. V. Geidoln, Physiol. Rev. 60, 1107 (1980).
 R. E. Kouri et al., Cancer Res. 38, 2777 (1978).
 A. Poland, D. Palen, E. Glover, Nature (London) 300, 271 (1982).
 R. H. Kouri and D. W. Nebert, in Origins of Human Cancer, H. H. Hiatt, J. D. Watson and J. A. Winsten, Eds. (Cold Spring Harbor Labo-ratory, Cold Spring Harbor, N.Y., 1977), vol. 4, p. 811; D. W. Nebert and N. M. Jensen, CRC Critical Reviews in Biochemistry, G. D. Fas-man, Ed. (CRC Press, Inc., Cleveland, Ohio, 1979), vol. 6, p. 401; D. W. Nebert, Environ. Health Perspect. 39, 11 (1981).
 R. E. Kouri et al., Banbury Rep. 16, 131 (1984); S. A. Atlas, E. S. Vesell, D. W. Nebert, Cancer Res. 36, 4619 (1976).
 V. J. Kidd, R. B. Wallace, K. Itakura, S. L. C. Woo, Nature (London) 304, 230 (1983).
 Institute of Medicine of the National Academy of Sciences, Genetic Influences on Responses to Converted Prevent of a Conference of the National Academy of Sciences, Genetic Influences on Responses to

- of Sciences, Genetic Influences on Responses to the Environment: Report of a Conference on Implications of Environmental/genetic Interac-

- tions (Institute of Medicine, National Academy of Sciences, Washington, D.C., 1981).
 13. H. Aviv and P. Leder, Proc. Natl. Acad. Sci. U.S.A. 69, 1408 (1972); F. J. Gonzalez and C. B. Kasper, Biochemistry 19, 1790 (1980).
 14. R. A. Young and R. W. Davis, Proc. Natl. Acad. Sci. U.S.A. 80, 1194 (1983).
 15. P. L. Deininger, Anal. Biochem. 129, 216 (1983).
 16. F. Sanger, S. Nicklen, A. R. Coulson, Proc. Natl. Acad. Sci. U.S.A. 74, 5463 (1977); J. Messing, R. Crea, P. H. Seeburg, Nucleic Acids Res. 9, 309 (1981).
 17. R. Staden, Nucleic Acids Res. 8, 3673 (1980); D. J. Brutlag, J. Clayton, P. Friedland, L. H.
- A. Stauen, Inucleic Acids Res. 8, 3673 (1980); D. J. Brutlag, J. Clayton, P. Friedland, L. H. Kedes, *ibid.* 10, 279 (1982); B. C. Orcott, D. G. George, J. A. Fredrickson, M. O. Dayhoff, *ibid.*, p. 157.
- 18. 19.
- 20.
- ibid., p. 157.
 Y. Quax-Jeuken, W. Quax, H. Bloemendal, ibid. 80, 3548 (1983).
 J. D. Watson, Molecular Biology of the Gene (Benjamin, Menlo Park, Calif., 1984).
 O. Gotoh, Y. Tagashira, T. Iizuka, Y. Fujii-Kuriyama, J. Biochem. (Tokyo) 93, 807 (1983);
 A. Kumar, C. Raphael, M. Adesnik, J. Biol. Chem. 258, 11280 (1983); Y. Mizukami, K. Sogawa, Y. Suwa, M. Muramatsu, Y. Fujii-Kuriyama, Proc. Natl. Acad. Sci. U.S.A. 80, 3958 (1983); K. Kawajiri et al., ibid. 81, 1649 (1984); K. Morohashi et al., ibid., p. 4647; Y. Yabusaki et al., Nucleic Acids Res. 12, 2929 Yabusaki et al., Nucleic Acids Res. 12, 2929 1984).
- M. Negishi and D. W. Nebert, J. Biol. Chem.
 256, 3085 (1981); R. H. Tukey, D. W. Nebert, M. Negishi, *ibid.*, p. 6969; R. H. Tukey and D. W. Nebert, *Biochemistry* 23, 6003 (1984).
- 22. A. K. Jaiswal, F. G. Gonzalez, D. W. Nebert, unpublished data.
- Trout has polycyclic hydrocarbon-inducible AHH (P_1 -450) activity but no evidence of "P-448" (defined here as P_3 -450) [J. T. Ahokas, H. Saarni, D. W. Nebert, O. Pelkonen, *Chem. Biol.* 23 Interact. 25, 103 (1979)]. This is further evolu-tionary evidence to suggest that P_1-450 is the original TCDD-inducible gene and that P_3-450
- Original 1CDD-inducible gene and that P₃-450 diverged from P₁-450.
 W. M. Fitch and C. H. Langley, Fed. Proc. Fed.
 Am. Soc. Exp. Biol. 35, 2092 (1976); A. C.
 Wilson, S. S. Carlson, T. J. White, Annu. Rev. Biochem. 46, 573 (1977); T. Miyata, J. Mol. Evol. 19, 28 (1982). 24.
- J. K. Leighton, B. A. DeBrunner-Vossbrinck, 25.
- J. K. Leighton, B. A. DeBrunner-Vossbrinck, B. Kemper, Biochemistry 23, 204 (1984).
 B. DeMartinville, A. R. Wyman, R. White, U. Francke, Am. J. Hum. Genet. 34, 216 (1982); C. D. Boehm, S. E. Antonarakis, J. A. Phillips III, G. Stetten, H. H. Kazazian, Jr., N. Engl. J. Med. 308, 1054 (1983); J. F. Gusella et al., Nature (London) 306, 234 (1983); P. S. Rotwein et al., N. Engl. J. Med. 308, 65 (1983); S. L. C. Woo, A. S. Lidsky, F. Güttler, T. Chandra, K. J. H. Robson. Nature (London) 306, 151 (1983); J. H. Robson, Nature (London) 306, 151 (1983);
 V. J. Kidd, M. S. Golbus, R. B. Wallace, K. Itakura, S. L. C. Woo, N. Engl. J. Med. 310, (20) (1994).
- 639 (1984).
 27. D. W. Nebert, Methods Enzymol. 52, 226
- D. W. Internet, (1978).
 L. W. Engel and N. A. Young, *Cancer Res.* 38, 4327 (1978).
 D. Wans and D. W. Nebert, *Mol. Pharma*-
- I. S. Owens and D. W. Nebert, *Mol. Pharmacol.* 11, 94 (1975); I. S. Owens, A. Niwa, D. W.
- col. 11, 94 (1975); I. S. Owens, A. Niwa, D. W. Nebert, Gene Expression and Carcinogenesis in Cultured Liver, L. E. Gerschenson and E. B. Thompson, Eds. (Academic Press, New York, 1975), p. 378; I. S. Owens and D. W. Nebert, Biochem. Pharmacol. 25, 805 (1976); C. Legraverend et al., J. Biol. Chem. 257, 6402 (1982).
 30. The mouse P₁-450 NH₂-terminal sequence determined on detergent-solubilized protein (M. Negishi, D. W. Nebert, J. E. Shively, unpublished data) was Y-G-L-P-A-F-V. ..., whereas the corresponding sequence determined on immunoaffinity-purified protein [K.-C. Cheng et al., Biochem. Biophys. Res. Commun. 123, 1201 (1984)] was P-S-M-Y-G-L-P-A-F-V. The original estimation of P₁-450 molecular weight (58,914) estimation of P_1 -450 molecular weight (58,914) and 521 residues (5) was therefore based on translation from the second of two "in-frame" ATG codons and the detergent-solubilized pro-Find the detergent solution and the detergent solution and the detergent solution has F_1 -450 has 524 residues and that the molecular weight is 59,230, based on the immunoaffinity-purified 59,230, based on the immunoaffinity-purified protein data. A similar observation of one or two NH₂-terminal residues being lost during detergent solubilization of rabbit P-450 form 4 has been reported [V. S. Fujita, S. D. Black, G. E. Tarr, D. R. Koop, M. J. Coon, Proc. Natl. Acad. Sci. U.S.A. 81, 4260 (1984)].
 31. HeLa cell DNA and human placental DNA were provided by O. Wesley McBride. The Southern blot was provided by C. Edgar Hildebrand.
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Human T-Cell Receptor α-Chain Genes:

Location on Chromosome 14

Abstract. The genes encoding the α chain of the human T-cell receptor have been mapped to chromosome 14, the chromosome on which the human immunoglobulin heavy chain locus resides. Thus, genes encoding two different classes of antigen receptor are present on the same chromosome. Furthermore, breaks involving chromosome 14 are frequently seen in tumors of T-cell origin. The potential relation of these chromosome abnormalities to α -chain genes is discussed.

The major histocompatibility complex (MHC) restricted antigen receptor on T lymphocytes is a disulfide-linked heterodimer and is composed of an α and a β chain (1). The complementary DNA (cDNA) clones encoding the β chain have been isolated and described in mice and man (2). The β -chain locus consists of several gene segments that encode variable (V), diversity (D), joining (J), and constant (C) regions of the β -chain protein, which is itself related to immunoglobulins. The β chain is encoded on chromosome 7 in man (3) and on chromosome 6 in mouse (4). The locus for murine immunoglobulin (Ig) kappa light chains is encoded on chromosome 6 as well (5), although the evolutionary significance of this linkage is not understood. The α -chain cDNA clones have been isolated and characterized in humans (6) and in mice (7). The sequence of these α chain cDNA's suggest that, like the β chain and the Ig's, the α chain is encoded in separate, noncontiguous gene segments (V, J, and C). We have now mapped the genes encoding the α chain and have assigned these genes to human chromosome 14.

We used a series of human-rodent somatic cell hybrids, containing characterizing combinations of human chromosomes (see legend to Fig. 1 for description of hybrids); the isolation and characterization of most of these hybrids have been described (8), and those not previously described are discussed in the legend to Fig. 1. Hybrid cells were grown in culture, and DNA was prepared from each hybrid (9). The presence or absence of human α -chain genes in each hybrid was determined by probing Southern blots of genomic DNA with a labeled α chain cDNA probe isolated from the human T-cell tumor, HPB-MLT. This human a-chain cDNA has been characterized by one of us (E.P.) and is described in (6).

Analysis of hybrid DNA's with an α chain cDNA probe by the Southern blot method (Fig. 1) shows three Hind III fragments (2.6, 4.4, and 9.6 kb) in human DNA. A cDNA fragment specific for the V region of this particular α chain hybridizes to the 9.6-kb Hind III fragment, whereas a cDNA fragment specific for the α -chain C region hybridizes to the 2.6- and 4.4-kb Hind III fragments (10).



Fig. 1. Southern analysis of human-rodent cell hybrids for human achain genes. Hybrid or tumor cell DNA was digested with Hind III and subjected to electrophoresis through 0.7 percent agarose gels. Samples of DNA were blotted and hybridized by the method of Wahl et al. (14), except that the acid depurination step was omitted and the gel was irradiated with shortwave ultraviolet for 8 minutes prior

to denaturation. The blots were hybridized with the 1.3-kb insert of pGA5, a cDNA clone that encodes the α chain from the human T-cell tumor HPB-MLT. This insert contains variable joining and constant region sequences and has been described (6). The upper, middle, and lower bands are 9.6, 4.4, and 2.6 kb, respectively. The faint bands seen in human DNA are apparently unrelated to the α -chain locus since they do not segregate with the major bands. They may represent α -chain pseudogenes, which are unlinked to the α -chain locus. The hybrid cell lines in (8) were used except for 32-1-A, a human lymphoblast-mouse L-cell hybrid; Cp23, a human fibroblast-Chinese hamster ade-B hybrid; Cp24, a human fibrosarcoma-Chinese hamster ade⁻B hybrid; 314-2, a human lymphocyte–Chinese hamster uri⁻C, ade⁻G hybrid; 706-D1, a human lymphocyte-Chinese hamster ade F hybrid.