able to regulate growth and, consequently, connectivity indicates that rather common simple molecules may also play prominent roles in regulating the pattern of neuronal connectivity.

P. G. HAYDON **D.** P. МсСовв S. B. KATER

Department of Zoology, University of Iowa, Iowa City 52242

References and Notes

- R. W. Gunderson and J. N. Barrett, J. Cell. Biol. 87, 546 (1980); E. R. Peterson and S. M. Crain, Dev. Brain Res. 2, 341 (1982),
 G. Lynch, B. Stanfield, C. W. Cotman, Brain Res. 59, 155 (1973); D. H. Hubel, T. N. Wiesel, S. Le Vay, Phil. Trans. R. Soc. London Ser. B 278, 377 (1977); M. Shankland, D. Bentley, C. S. Goodman, Dev. Biol. 92, 507 (1982); S. Denis-Donini, J. Glowinski, A. Prochiantz, J. Neuro-sci. 3, 2292 (1983).
- 3.
- Domini, J. Glowinski, A. Prochiantz, J. Neuro-sci. 3, 2292 (1983). C. E. Aguilar, M. A. Bisby, E. Cooper, J. Diamond, J. Physiol. (London) 234, 449 (1973). R. I. Hume, L. W. Role, G. D. Fischbach, Nature (London) 305, 632 (1983); S. H. Young and M. M. Poo, *ibid.*, p. 634. Experiments with high concentrations of seroto-nin ($10^{-4}M$) suggest that this neurotransmitter may affect the initiation of outgrowth [M. A.
- 5.
- nin (10 ⁷M) suggest that this neurotransmitter may affect the initiation of outgrowth [M. A. Kostenko, V. S. Musienko, T. I. Smolikhina, *Brain Res.* 276, 43 (1983)].
 R. D. Hadley, S. B. Kater, C. S. Cohan, *Science* 221, 466 (1983). The formation of electrical connections critically relies on a spatial and temporal coincidence of neurite outgrowth from both across and the second second second second second second sections of the second second second second second second sections of the second second second second second second sections of the second second second second second second sections of the second second second second second second second second sections of the second second second second second second second second sections of the second second
- both partner neurons.
 A. G. M. Bulloch and S. B. Kater, J. Neurophysiol. 48, 569 (1982); R. D. Hadley and S. B. Kater, J. Neurosci. 3, 924 (1983); P. G. Haydou and S. B. Kater, Soc. Neurosci. Abstr. 9, 371 (1983)
- R. G. Wong, R. D. Hadley, S. B. Kater, G. C. Hauser, J. Neurosci. 1, 1008 (1981).
- Central ganglia were treated with trypsin, the sheaths were cut with a tungsten microknife, 9 and identified neurons were removed and trans ferred to culture dishes with a glass micropi-
- S. B. Kater and R. D. Hadley, Trends Neurosci. 10. 5, 80 (1982). The rate of neurite elongation was quantified by measuring the advance of the leading edge of the growth cone either directly from the television monitor of the video microscopy system or from monochrome photographs taken at 20-minute frame intervals. The effect of serotonin on neurite elongation was assessed by comparing a control period of neurite outgrowth before serotonin was added (left of arrows in Fig. 1A) with a test period after serotonin was added (right of arrows in Fig. 1A). Neurons retain all of their normal electrophysio-
- logical properties while exposed to serotonin. Focal application of the carrier medium alone does not cause the effects seen on exposure to serotonin.
- G. Shaw and D. Bray, *Exp. Cell Res.* 104, 55 (1977); N. K. Wessells, S. R. Johnson, R. P. 13. (1977); N. K. Wessells, S. I Nuttall, *ibid*. 117, 335 (1978).
- M. V. L. Bennett, Ann. N. Y. Acad. Sci. 137, 509 (1966). The coupling coefficient is the ratio 14. of postsynaptic to presynaptic voltage (range 0.0 to 1.0) with d-c current injection. Coupling coefficients are reported here as means and standard errors.
- Serotonin significantly reduced the strength of 15. Schöhlin all spin tearlog reduced the strong for the electrical connection between neurons 5 and 19 [t(8) = 4.15, P < 0.002; t-test] and increased the frequency of zero coupling values [$\chi^{2}(1) = 9.0$, P < 0.01]. Of 12 neurons P5 studied, serotonin (10⁻⁶ to $\Sigma < 0.0^{-2}$).
- 16. $5 \times 10^{-5} M$) transiently inhibited the growth cone motile activity of eight neurons for periods of up to 100 minutes and totally inhibited the activity of the other four. 17. J. M. Lauder and H. Krebs, *Dev. Neurosci.* 1, 15. (JCT2)
- (1978).
- T. Kasamatsu and J. D. Pettigrew, J. Comp. 18.
- Kasamatsu and J. D. Petuglew, J. Comp. Neurol. 185, 139 (1979); T. Kasamatsu, J. D. Pettigrew, M. Ary, *ibid.*, p. 163.
 We thank C. S. Cohan, J. L. Denburg, R. K. Small, and C. F. Wu for their comments, S. Haas and L. Zachar for data analysis, and P. Cada for photographic assistance. Sumported by 19. Gade for photographic assistance. Supported by PHS grants NS15350, NS18819, and HD18577.
- 17 April 1984; accepted 15 June 1984

Extrapulmonary Gas Exchange Enhances Brain Oxygen in Pigeons

Abstract. Blood in mouth, nose, and eye tissues of birds cools by evaporation, then flows to a cephalic vascular heat exchanger, the ophthalmic rete. There, acting as a heat sink, blood from the evaporative surfaces cools arterial blood flowing countercurrent to it toward the brain. The brain thus remains cooler than the body core. Data for unanesthetized domestic pigeons (Columba livia) suggest that in addition to losing heat, blood perfusing the evaporative surfaces also exchanges oxygen and carbon dioxide with air. In the heat exchanger, this blood apparently gives up oxygen to, and gains carbon dioxide from, arterial blood. The consequent increase in oxygen and decrease in carbon dioxide in the brain's arterial blood enhance diffusion of these gases in, and oxygen supply to, the brain. Such events may help birds maintain the brain's oxygen supply during the high systemic demand of exercise and at the reduced oxygen availability of high altitude.

Evaporation from mouth, nose, and eye surfaces in birds cools the blood flowing just beneath these surfaces. The cooled blood then courses to the ophthalmic rete (OR), a network of small arteries and veins caudal to each eye (Fig. 1). Arterial blood in the OR conducts heat to venous blood flowing countercurrent to it, then proceeds to the brain at reduced temperature. This protects the brain from overheating when body temperature rises during exercise or high temperature exposure (1).

Oxygen and carbon dioxide tensions $(Po_2 \text{ and } Pco_2)$ in avian arterial blood at sea level are about 85 and 30 torr, compared to 159 and 0 torr in air (2). Blood circulating in the dense vascular beds beneath the moist cephalic surfaces must therefore encounter large gradients for these gases, and it seemed reasonable to infer that air and blood at these surfaces might exchange O₂ and CO₂, as well as water vapor and heat. Venous blood flowing to the OR would then not only have a lower temperature but would also have an increased Po_2 and a reduced Pco₂. Our preliminary calculations indicated that Po_2 and Pco_2 in the OR veins would be higher and lower, respectively, than in OR arteries, suggesting that the OR is a site of O_2 diffusion from venous to arterial blood, and of CO₂ diffusion in the opposite direction, as well as a site of heat exchange. Furthermore, as shown in Fig. 1, changes in temperature, Pco_2 , and pH would affect hemoglobin O_2 binding in a manner conducive to both O₂ loading at the moist surfaces and O₂ transfer from venous to arterial blood in the OR (3).

The data presented here support this idea. They show that Po_2 in cerebral arterial blood of domestic pigeons (Columba livia: mean mass, 0.34 kg) is greater than in carotid arterial blood and suggest that the Po₂ difference is correlated with the extent to which cephalic mucosal surfaces contact air. An elevated Po₂ in cerebral arteries would increase the Po₂ in brain capillaries, improving O_2 diffusion to tissue. If arterial O_2 saturation also increases, total O_2 flow to brain would also increase.

To test the idea, we compared gas tensions in arterial blood before it entered the OR (pre-OR) and after it left the OR (post-OR) to determine whether enhancement of gas exchange occurs in the OR itself. Pre-OR arterial gas tensions were measured in carotid blood. However, post-OR arteries were inaccessible for sampling. We therefore reasoned that gas tensions in cerebrospinal fluid (CSF) sampled near the lateral ventricle's choroid plexus, a site of CSF formation (4), would represent corresponding values in choroid plexus blood, at least some of which comes from the OR. Accordingly anaerobic samples of CSF and arterial blood were simultaneously sampled from unanesthetized birds, and Po₂, Pco₂, and pH were measured (5).

Figure 2 shows results for 11 control birds at room temperature (experiment 1). Variances for mean differences were not significantly different from each other; we therefore used *t*-tests for comparing means. The mean Po_2 of the CSF was 114 torr (± 3 standard deviation). This is 39 percent greater [t(10) = 24.8;P < 0.0001 than the mean Po_2 in carotid blood [82 torr (± 3)]. We also found that Pco₂ was significantly lower in CSF [23.3 torr (± 2.2)] than in carotid blood $[31.4 \text{ torr } (\pm 2.3)]$, a 26 percent decrease [t(10) = -8.5, P < 0.0001]. The changes in cerebral arterial Po₂ and Pco₂ make possible increases in O₂ diffusion to and CO₂ diffusion from brain tissue.

Our results may also be explained by possible carbonic anhydrase activity in the ependymal cells of the choroid plexus. It has been suggested that the ependymal carbonic anhydrase reaction produces HCO_3^- , which moves into CSF, and H⁺, which enters choroid capillary plasma (6). Erythrocytic bicarbonate also moves into plasma to maintain ionic balance, combining with H^+ to form CO_2 . The CO_2 moves into erythrocytes where carbonic anhydrase catalyzes its conversion to HCO_3^- , replacing $HCO_3^$ lost to plasma. Meanwhile H^+ produced in the same reaction facilitates the release of O_2 from hemoglobin, which moves into CSF and increases its PO_2 . This is the mechanism most likely to



Fig. 1. (A) Simplified diagram of blood flow in pigeon head. Arrows show flow direction. Arterial blood (unshaded) flows to brain directly and via the bilateral ophthalmic retia (only one shown), and to the nasal and oral mucosa (stippled). Venous blood (shaded), cooled at the cephalic mucosa to about 36°C in this example, flows through the OR, where arterial blood at about 41°C is cooled by countercurrent heat exchange before flowing to brain. (B) Pigeon blood oxyhemoglobin dissociation curves showing changes in blood O_2 saturation (So₂) and tension (Po₂) evoked by changes in temperature, CO₂ tension (Pco_2) , and pH (3). Letters a through f on curves correspond to locations on the diagram. The hypothesis presented here is as follows. Arterial blood at $Po_2 = 80$ torr and $So_2 = 85$ percent [point a (2)] cools and loses CO₂ to air at mucosa, driving the curve leftward (point b). This blood also loads O₂ from air, raising Po2 and So2 to point c. On returning to the rete the curve moves rightward (point d) as it obtains heat and CO_2 from arterial blood. The latter moves to point b because of its heat and CO₂ loss, establishing a retial O₂ gradient from point d to point b. If retial gas exchange is complete, arterial blood from the OR will enter the brain virtually saturated at $Po_2 = 115$ torr (point e), whereas venous blood from the OR will flow to the heart at point f. If retial exchange is not complete, points e and f for arterial and venous retial outflow will be closer together. Thus, final brain Po₂ depends on the proportioning of arterial flow between direct and retial routes, and on completeness of gas and heat exchange in the mucosa and OR.

operate in the brain's glial cells for direct oxygenation of interstitial fluid (6).

To control for the possibility that this model may explain our CSF results in pigeons, we performed two additional experiments. In experiment 2, we plugged the nares with Plasticine and taped the beak to prevent contact between air and oronasal surfaces. Experiment 3 was identical to experiment 2, except that we also covered the closed eyelids with sheet paraffin to prevent contact between air and eye surfaces. Birds breathed freely via tracheostomies (5) in both experiments. The experimental treatments did not significantly affect arterial values of Po_2 and Pco_2 (Fig. 2). Cerebrospinal Po2, in contrast, fell significantly in experiment 2, although not all the way to the arterial level. In experiment 3, CSF Po₂ fell even farther and became indistinguishable from carotid arterial Po_2 . These results indicate that the nose, mouth, and eyes participate in the processes that determine CSF Po_2 in these animals.

Cerebrospinal Pco_2 increased from below the carotid level in experiment 1 to above the carotid level in experiments 2 and 3 (Fig. 2). We think that this finding resulted from our preventing gas exchange between the evaporative surfaces and air in experiments 2 and 3, so that venous Pco_2 in the OR was greater than Pco_2 in arterial blood. The arterial blood therefore must have gained rather than lost CO_2 in the OR before it flowed to the brain, contrary to the model in Fig. 1. We also found that pH in CSF was lower than in arterial blood in all the experiments-probably because of the absence of protein buffers in CSF-and did not change (Fig. 2). This suggests that CSF pH homeostasis is the probable end result of CSF HCO_3^- regulation caused by shifts in CA activity during disturbances in dissolved gas tensions.

Gas tension results from experiments 1, 2, and 3 are consistent only with the hypothesis that moist cephalic surfaces take up O_2 and release CO_2 and that O_2 and CO_2 exchange occurs within the OR. The results of experiments 2 and 3 would not have been observed if acid-base changes in the ependymal cells were the sole explanation for enhancement of CSF PO_2 .

Sealing the eyes against contact with air (experiment 3) affected Po_2 but not Pco_2 in the CSF (Fig. 2). Because CO_2 solubility in aqueous solutions is about 30 times greater than O_2 solubility, a large amount of CO_2 could diffuse from plasma to air without markedly altering the molar CO_2 content of the plasma. On the other hand, during O_2 diffusion from

air to blood, a small increase of molar O₂ concentration in plasma, caused by diffusion from air, would lead to a large increase in plasma Po₂. Because total surface area of the eyes is small compared to nasal and oral surfaces, the molar CO₂ content of blood might not be significantly reduced in the time of blood transit through the eyes, whereas CO_2 loss from blood flowing through nose and mouth tissues might be enough to reduce Pco₂ significantly. Therefore sealing the nose and mouth would evoke a significant reduction in the loss of blood CO_2 as well as in the uptake of O_2 , whereas additionally sealing the eyes would reduce further the uptake of O_2 but not the loss of CO_2 .

The physical properties of avian blood support the possibility of augmented cerebral oxygen supply. Because of the



Fig. 2. Oxygen and CO_2 tensions and pH in anaerobic samples of cerebrospinal fluid (unshaded bars) and carotid arterial blood (shaded bars) obtained simultaneously from unanesthetized pigeons (environmental temperature, 23°C and barometric pressure, 665 torr). Bar heights represent means; vertical lines extend +1 standard deviation. Numbers across top represent numbers of individual birds used in each experiment. Each bird was used once. Experiment 1 was performed on intact control birds; experiment 2 was performed on tracheostomized birds with mouth and nose sealed against contact with air; and experiment 3 was identical to experiment 2, except that eyes were also sealed.

comparatively large size and nondeformability of avian erythrocytes, at equal tube diameters and hematocrits the relative viscosity of avian erythrocyte suspensions is four to six times greater than in mammalian erythrocyte suspensions (7). In apparent compensation, the fraction of tissue volume occupied by brain capillaries is about twice as high in birds as in mammals (8). In a tissue like the brain, contained in an inelastic skull, further increases in capillary density would increase O₂ delivery, but the resulting increase in tissue volume occupied by capillaries would reduce neuronal space. It therefore seems plausible that further evolutionary improvements in O_2 delivery would be brought about through increases in the diffusion gradient; this could be achieved by means of the proposed mechanism.

We conclude that the Po_2 is increased and the Pco_2 is decreased in a portion of the arterial supply to the pigeon brain, thereby facilitating diffusion of O₂ into and CO₂ out of brain tissue by increasing the respective gradients. Blood O₂ affinity is lower in birds than in similar-sized mammals, and arterial blood is not likely under most conditions to be saturated with $O_2(9)$. We therefore suggest further that, in raising arterial Po_2 , the mechanism must also increase hemoglobin O₂ saturation. This would (i) sustain steadystate O₂ delivery at lower cerebral blood flows than would otherwise be necessary, and (ii) increase brain O₂ delivery under conditions in which an increase in cerebral blood flow alone could not meet the O_2 demand, such as at high altitudes. MARVIN H. BERNSTEIN

HARRY L. DURAN*

Department of Biology, New Mexico State University, Las Cruces 88003 **BERRY PINSHOW**

Jacob Blaustein Institute for Desert Research, Ben-Gurion University, 84990 Sede Boger Campus, Israel

References and Notes

 Brain cooling has been shown in numerous studies [for example, D. L. Kilgore, M. H. Bernstein, D. M. Hudson, J. Comp. Physiol. 110, 209 (1976)]. Four kinds of evidence imply the involvement of nose and mouth: (i) ablation experiments [M. H. Bernstein, I. Sandoval, M. B. Curtis, D. M. Hudson, J. Comp. Physiol. 129, 115 (1979)]; (ii) histological observations of 129, 113 (1979), (i) histological observations of dense vascular networks in nose and mouth tissue (M. H. Bernstein, unpublished results);
(iii) studies of cephalic vascular anatomy [for example, C. Bech and U. Midtgard, J. Comp. Physiol. 145, 89 (1981); P. G. H. Frost, W. R. Siegfried, P. J. Greenwood, J. Zool. 175, 231 (1975); D. L. Kilgore, M. H. Bernstein, D. M. Hudson, J. Comp. Physiol. 110, 209 (1976); U. Midtgard, Z. Arad, E. Skadhauge, J. Comp. Physiol. in press; S. A. Richards, J. Zool. 153, 221 (1968); Brain Res. 23, 265 (1970); A. Vitums, M. Shin-Ichi, D. S. Farner, Anat. Anz. 116, 309 (1965); K. G. Wingstrand and O. Munk, Biol. Skrift. 14, 1 (1965)], and (iv) studies of cerebral blood flow, especially during heat stress [for example, C. Bech and K. Johansen, Acta Physiol. Scand. 110, 351 (1980)]. Ablation experivascular networks in nose and mouth ol. Scand. 110, 351 (1980)]. Ablation experi-

ments and experiments on intraocular and brain temperatures in pigeons during corneal ventila-tion [B. Pinshow, M. H. Bernstein, G. E. Lo-pez, S. Kleinhaus, Am. J. Physiol. 242, R577 (1982)] have established the involvement of the

- eyes.
 P. L. Lutz and K. Schmidt-Nielsen, Respir. Physiol. 30, 383 (1977); P. J. Butler, N. H. West, D. R. Jones, J. Exp. Biol. 71, 7 (1977); Y. Weinstein, M. H. Bernstein, P. E. Bickler, D. V. Gonzales, F. C. Samaniego, M. A. Esco-bedo, Am. J. Physiol., in press.
 B. Pinshow, M. H. Bernstein, Z. Arad, in prepa-ration
- H. F. Cserr, M. Bundgaard, J. K. Ashby, M. Murray, J. Physiol. 238, R76 (1980); K. Ko-shiba, H. Oshima, H. Uematsu, J. Electron Microsc. 29, 129 (1980). 4.
- 5. We obtained anaerobic samples of cerebrospinal fluid from the lateral ventricle via an indwelling alast cannula; this is a modification of the tech-nique of J. H. Brakkee, V. M. Wiegant, W. H. Gispen, Lab. Anim. Sci. 29, 78 (1979). We drew Chippen, Lab. Anim. Sci. 29, 78 (1979). We drew anaerobic blood samples, via an indwelling poly-ethylene catheter in one carotid artery, into a chilled, heparinized glass syringe. Cannulas were implanted and tracheostomies were per-

formed in animals given local anesthesia (mepivacaine HCl, 1 percent), and the animals were allowed to recover for 1 day. All samples were obtained from unanesthetized animals and analyzed immediately with a blood gas acid-base analyzer (Radiometer BMS3Mk2) adjusted to body temperature and calibrated with certified gas and pH standards immediately prior to each measurement.

- L. Jankowska and P. Grieb, Am J. Physiol. 236, 6. F220 (1979).
- P. Gaehtgens, F. Schmidt, G. Will, *Pfluegers* Arch. 390, 278 (1981); P. Gaehtgens, G. Will, F. 7.
- Arch. 590, 276 (1961), p. 283.
 W. Lierse, Acta Anat. 54, 1 (1963).
 P. L. Lutz, Am. Zool. 20, 187 (1980).
 Supported by NSF grants PCM-8118956 and PCM-8402659 and by Minority Biomedical Re-10. search Support Program grant RR08136 from the National Heart, Lung, and Blood Institute through the Division of Research Resources, National NIH.
- Present address: Department of Pharmacology, College of Medicine, Ohio State University, Columbus 43210.

12 March 1984; accepted 8 August 1984

Induction of Pituitary Lactotrope Differentiation by Luteinizing Hormone α Subunit

Abstract. Addition of gonadotropin releasing hormone to cultures of fetal rat pituitary induced differentiation of lactotropes as revealed by immunocytochemistry. Antiserum to luteinizing hormone (LH) (recognizing native LH), but not antiserum to LH- β (recognizing both native LH and its β subunit), inhibited this induction. Further addition of highly purified LH-a subunit in culture medium also induced lactotrope differentiation. Thus, the α subunit may have a specific biological activity of its own with probable practical use in clinical investigations.

Induction of pituitary lactotrope differentiation by gonadotropin releasing hormone (GnRH) has been reported (1). This effect of GnRH is possibly subsequent to a primary action on gonadotropes and induces a further release of stimulating factor or factors. Such a chain of events has been described for the stimulation of the release of prolactin

from adult rat lactotropes (2). Moreover in human anencephalic fetuses, where hypothalamic GnRH is lacking, lactotropes are numerous and active (3). In these types of fetuses, gonadotropes are known to store the α subunit almost exclusively (4, 5). A possible relation between the abundance of this subunit and the proliferation of lactotropes was therefore considered. We now describe the role of the α subunit as mediator in the action of GnRH and in the differentiation of lactotropes.

Adenohypophysial primordia were explanted from 13-day-old rat fetuses and cultured in a synthetic medium as described (1, 6). Cultures were maintained for 8 days to reach the expected term of gestation when the first lactotropes would normally appear in vivo (7). The culture medium was renewed every 2 days.

Fig. 1. Immunocytochemistry with antiserum to rat prolactin diluted 1:800 at 4°C overnight (light microscopy). This antiserum has been studied and used previously (1). Typical primordium cultured in medium containing LH- α (10⁻⁷M) (the experiment was repeated four times, and a total of 20 primordia was studied). (A) Several immunoreactive cells are scattered in the cultured tissue ($\times 250$). (B) The immunoreactive cells are irregularly shaped and intensely stained (×1200).

