

neuronal activity responsible for actual behavior in response to light [7]; Y. Goh and D. L. Alkon, in preparation].

7. D. L. Alkon, *Biol. Bull. (Woods Hole, Mass.)* **159**, 505 (1980).
8. ———, *Science* **205**, 810 (1979).
9. J. J. Shoukimas and D. L. Alkon, *Soc. Neurosci. Abstr.* **6**, 17 (1980).
10. J. A. Connor and C. F. Stevens, *J. Physiol. (London)* **213**, 21 (1971).
11. S. H. Thompson, *ibid.* **265**, 465 (1977).
12. J. J. Shoukimas and D. L. Alkon, in preparation.
13. The treatment condition of each animal was revealed after all voltage-clamp recordings had been analyzed and the quantitative measurements made.
14. A number of criteria for acceptability of impalement were used: (i) the resting potentials recorded by the voltage-recording and current passing electrodes should be approximately equal, (ii) the response to light recorded by each electrode should be approximately the same, (iii) the leak current at 0 mV should be ≤ 10 nA, (iv) the settling time of the voltage clamp should be ≤ 20 msec, and (v) the holding current at -60 mV should be ≤ 5 nA.
15. Corrections of I_A values for "leak" current were obtained by extrapolation from a linear portion of the current-voltage relation, which was generated with small positive and negative command pulses. I_A was also corrected for I_B , which was

estimated from current values measured 1 second from the onset of the second of the twin pulses. This estimate agreed very closely with I_B values obtained with a test pulse (to 0 mV) occurring 30 msec after a 5.0-second preliminary pulse to -10 mV. This correction for I_B was arrived at after numerous previous experiments (J. J. Shoukimas and D. L. Alkon, unpublished observations) that established I_B characteristics in the absence of I_A (after blocking with 4-aminopyridine).

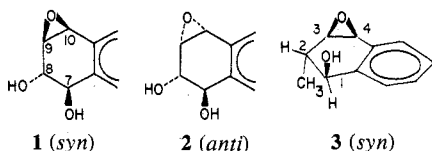
16. The time constant for inactivation of I_A is much greater than the time constant of inactivation. Furthermore, there were no apparent between-group differences in the rising phase of I_A . Between-group differences in the decrement of I_A for the second of the twin pulses, therefore, were considered to largely reflect changes of inactivation of I_A rather than activation.
17. Other electrophysiological studies are also suggesting differences among the three type B cells with regard to changes during associative learning (T. Crow and D. L. Alkon, in preparation; J. Farley and D. L. Alkon, in preparation).
18. D. L. Alkon, *Science* **210**, 1375 (1980); J. Farley and D. L. Alkon, *ibid.*, p. 1373.
19. D. L. Alkon, J. J. Shoukimas, E. Heldman, unpublished observations.
20. M. Klein and E. R. Kandel, *Proc. Natl. Acad. Sci. U.S.A.* **77**, 6912 (1980).

23 July 1981; revised 9 September 1981

Internal Hydrogen Bond Formation in a *syn*-Hydroxyepoxide

Abstract. The existence of an internal hydrogen bond in a compound representative of a *syn* diol epoxide (a possible intermediate in chemical carcinogenesis by certain polycyclic aromatic hydrocarbons) has been demonstrated by x-ray crystallographic and nuclear magnetic resonance studies. This internal hydrogen bond was found in 3,4-epoxy-2-methyl-1,2,3,4-tetrahydro-1-naphthol and was shown to persist in dioxane-water solutions containing up to 80 mole percent water. In this structure, the 1-hydroxy and 2-methyl groups are shown to occupy axial positions. In the *anti* diol epoxide, which has no internal hydrogen bond, analogous groups are equatorial. Crystals of the compound were unstable in the x-ray beam while the data were being collected (even at low temperatures), presumably as a result of decomposition.

Initiation of carcinogenesis by benzo[a]pyrene (BaP) is considered by many to proceed by alkylation of DNA by an intermediate diol epoxide metabolite of BaP (1). Recent studies have shown that rat liver microsomes convert BaP, in part, to two diastereomeric diol epoxides [(-)-1 and (+)-2] of high enantiomeric purity (2). The isomers are designated *syn* if the benzylic hydroxyl (the hydroxyl at position 7 of 1) and epoxide groups project on the same side of the tetrahydrobenzene ring system (1), and *anti* if they lie on opposite sides (2). Isomer 2 has been identified as a potent carcinogen in newborn mice, and is thought to be the ultimate carcinogen of BaP in this system (3).

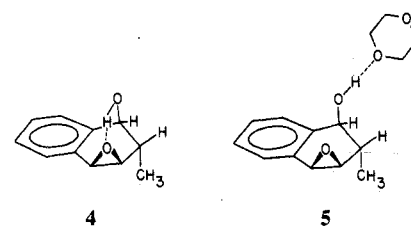


Hulbert (4) pointed out that internal hydrogen bonding between the benzylic hydroxyl and epoxide groups is possible in *syn* diol epoxide 1. Since this hydro-

gen bonding would weaken the C-O bonds of the epoxide, it was pointed out that nucleophilic attack at C(9) or C(10) of 1 would be facilitated. Indeed, the *syn* isomer (1) is about 160 times more reactive toward nucleophilic reaction at C(10) by the *p*-nitrothiophenolate ion in *t*-butyl alcohol than its *anti* isomer 2, which appears not able to form such a stable intramolecular hydrogen bond (5). Presumably, then, the greater reactivity

of the *syn* isomer causes it to interact with other macromolecules before it can reach the critical target for carcinogenesis. The structure of the *anti* BaP diol epoxide 2 has been studied by x-ray diffraction techniques (6). In addition, the probable three-dimensional structures of both isomers 1 and 2 have been constructed mathematically by merging known crystal structures (7).

We report here the crystal structure of a tetrahydronaphthalene epoxide 3 containing a benzylic hydroxyl group located *syn* to the epoxide group (3). This compound is structurally related to the *syn* naphthalenediol epoxide (1), with the C(8) hydroxyl group of 1 replaced in 3 by a methyl group on C(2) (equivalent to C(8) of 1). Compound 3 was prepared from *trans*-2-methyl-1-acetoxy-1,2,3,4-tetrahydronaphthalene by the following series of reactions: *N*-bromosuccinimide (NBS) bromination at C(4), dehydrobromination, and epoxidation to yield a mixture of the acetate of 3 and a diastereomeric acetate with the epoxide and acetate groups in a *trans* relation. The isomeric acetates were separated by alumina chromatography, and the *syn* acetate was hydrolyzed to 3. Purification of 3 was effected by crystallization from an ether-pentane solution (8).



The ^1H nuclear magnetic resonance (NMR) spectrum of 3 in either CCl_4 or $[\text{H}_8]\text{dioxane}$ revealed a small coupling of 2 Hz between H(1) and H(2), which suggests that the C(1) hydroxyl and C(2) methyl groups occupy axial positions. The ^1H NMR spectrum of 3 in CCl_4 showed an H(1)-OH coupling of 12 Hz

Table 1. Atomic parameters as fractions of cell edges with estimated standard deviation values in parentheses.

Atom	x	y	z	Atom	x	y	z
O(1)	0.4476(3)	0.7269(3)	-0.1283(4)	H(O1)	0.417(5)	0.647(5)	-0.051(7)
O(2)	0.4245(3)	0.6013(3)	0.2567(4)	H(C1)	0.679(3)	0.930(3)	-0.014(5)
C(1)	0.6138(4)	0.8298(4)	0.0350(6)	H(C2)	0.488(4)	0.888(3)	0.201(5)
C(2)	0.5727(4)	0.8758(4)	0.2448(6)	H(C3)	0.500(3)	0.771(3)	0.536(5)
C(3)	0.5247(4)	0.7542(4)	0.3805(6)	H(C4)	0.639(4)	0.607(4)	0.525(6)
C(4)	0.6229(4)	0.6575(4)	0.3922(6)	H(C5)	0.868(5)	0.558(5)	0.438(7)
C(5)	0.8891(4)	0.6194(4)	0.3145(6)	H(C6)	1.101(5)	0.592(4)	0.226(6)
C(6)	1.0161(5)	0.6420(4)	0.1948(8)	H(C7)	1.117(4)	0.756(4)	-0.073(6)
C(7)	1.0182(5)	0.7257(5)	0.0265(7)	H(C8)	0.892(4)	0.850(3)	-0.149(5)
C(8)	0.8902(5)	0.7887(4)	-0.0239(6)	H1(C11)	0.861(5)	1.006(5)	0.450(7)
C(9)	0.7608(4)	0.7669(4)	0.0936(6)	H2(C11)	0.765(4)	1.101(4)	0.314(5)
C(10)	0.7585(4)	0.6810(4)	0.2623(6)	H3(C11)	0.707(4)	1.029(3)	0.540(5)
C(11)	0.7295(5)	1.0175(4)	0.3933(7)				

