

Orth *et al.* (8) briefly reported findings on a blind subject who appeared to have a "free-running" component of cortisol secretion despite a normal sleep-wake cycle. In several other studies, psychologically normal subjects not isolated from time cues, but with varying degrees of blindness, did not show evidence of "free-running" circadian rhythms (9). Nevertheless, our subsequent survey of 50 subjects with varying degrees of blindness revealed that 38 complained of a significant sleep-wake disorder. Of these, 20 reported that their symptoms were cyclic or episodic, 14 had no light perception, 18 were blind from birth, and 13 had retrolental fibroplasia (as did J.X.).

The syndrome suffered by J.X. is not necessarily restricted to the blind. When symptoms are less severe, the syndrome would rarely be suspected, and without sophisticated long-term monitoring or autorhythmometry (10) the diagnosis will be difficult to confirm. Nevertheless, the disorder might not be uncommon, and the social and economic impact of even minor symptoms might be substantial.

In view of the many publications which have demonstrated that animal (including human) biological systems are influenced by lunar rhythms (11), it is notable that J.X. maintains a circadian rhythm that cannot be significantly distinguished from the period of the lunar day (24.84 hours). Furthermore, throughout the ad-lib sleep study, there was a remarkable coincidence between his sleep onset and a local low tide.

Full details of this unusual case are in preparation (12).

L. E. M. MILES  
D. M. RAYNAL  
M. A. WILSON

Department of Medicine and Sleep  
Research Center, Stanford University  
School of Medicine, and  
Veterans Administration Hospital,  
Palo Alto, California 94305

#### References and Notes

1. E. Hoddes, V. Zarcone, H. Smythe, R. Phillips, W. C. Dement, *Psychophysiology* **10**, 431 (1973).
2. J. Aschoff, *Aerosp. Med.* **40**, 844 (1969); W. B. Webb and H. W. Agnew, Jr., *ibid.* **45**, 617 (1974).
3. W. B. Webb and H. W. Agnew, Jr., *ibid.* **45**, 701 (1974).
4. F. Halberg, in *Symposium on Biological Cycles and Psychiatry, Symposium Bel-Air III* (Masson, Geneva, 1968), pp. 73-126.
5. O. Remler, *Klin. Monatsbl. Augenheilkd.* **113**, 116 (1948).
6. R. W. Bryson and D. F. Martin, *Lancet* **1954-II**, 365 (1954).
7. W. B. Webb and H. W. Agnew, *Sleep Res.* **1**, 179 (1972).
8. D. N. Orth, P. H. King, W. E. Nicholson, 58th Annual Meeting of the Endocrine Society, New York, June 1975, abstract 504.
9. D. T. Krieger and S. J. Glick, *Clin. Endocrinol.* **33**, 847 (1971); E. D. Weitzman, M. Perlow, J.

- F. Sassin, D. Fukushima, B. Burak, L. Hellman, *Trans. Am. Neurol. Assoc.* **97**, 197 (1972); F. Hollwich and B. Dieckhues, in *Chronobiology*, L. E. Scheving, F. Halberg, J. E. Pauly, Eds. (Igaku Shoin, Tokyo, 1974), pp. 285-292; B. D'Alessandro, A. Bellastella, U. Esposito, C. F. Colucci, N. Montalbetti, *Br. Med. J.* **2**, 274 (1974); M. L. Simenhoff, *J. Appl. Physiol.* **37**, 374 (1974).
10. F. Halberg, R. Lauro, F. Carandente, *Ric. Clin. Lab.* **6**, 207 (1976).
11. E. Bunning, *The Physiological Clock* (English Universities Press, London, 1973), pp. 177-188;

- F. A. Brown, Jr., in *Chronobiology*, L. E. Scheving, F. Halberg, J. E. Pauly, Eds. (Igaku Shoin, Tokyo, 1974), pp. 689-693.
12. M. A. Wilson, D. M. Raynal, L. E. M. Miles, in preparation.
13. We thank E. Chiang for assistance in assembling the sleep recording data, and Dr. W. C. Dement for his advice and encouragement. This work was supported by NIMH grant MH29861 and by a research fellowship from the Medical Research Council of New Zealand to M.A.W.

8 July 1977

## Memory Formation: Evidence for a Specific Neurochemical System in the Amygdala

**Abstract.**  $\beta$ -Adrenergic antagonists injected into the amygdala complex of rats trained in a passive avoidance task produced time-dependent and dose-dependent decreases in retention of the task. In addition, the effects observed with  $\beta$ -adrenergic antagonists were both stereospecific and reversed by norepinephrine. The results support a role for an amygdala  $\beta$ -adrenergic system in memory processes.

Clinical observations of retrograde amnesia following brain trauma (1), and permanent loss of long-term memory formation capabilities with temporal lobe resection in humans (2), have guided investigations with laboratory animals in which attempts have been made to elucidate the neuroanatomical and neurochemical substrates underlying memory formation. Although a wide variety of disruptive agents such as electroconvulsive shock, anesthetics, and protein synthesis inhibitors have been used in these investigations, the results generally agree that the sooner the agent is applied after an experience, the greater is the amnesia for the experience (3). This time-dependent retrograde amnesia gradient confirms and extends the earlier clinical findings of retrograde amnesia

following human brain trauma. However, owing to the widespread and non-specific effects of many of these amnesic agents (4), our knowledge of the specific neuroanatomical and neurochemical substrates underlying the memory process has not been greatly enhanced by their use.

Researchers have also investigated systematically the neuroanatomical and neurochemical systems involved in long-term memory formation in animals (5, 6). Here we report that microinjections of  $\beta$ -adrenergic blocking agents into the amygdala of rats that have received a single training experience produce retrograde amnesia which is both time-dependent and dose-dependent. Furthermore, the amnesia produced by these agents is demonstrated to be stereospecific and re-

Table 1. Latencies on day 2. The control groups were as follows: 1, no surgery; 2, surgery only; 3, vehicle only.

| Group          | Dose (nmole)         | N  | Median (seconds) | Interquartile range (seconds) |
|----------------|----------------------|----|------------------|-------------------------------|
| Control groups |                      |    |                  |                               |
| 1              |                      | 17 | 268              | 139 to 576                    |
| 2              |                      | 9  | 279              | 80 to 411                     |
| 3              |                      | 8  | 216              | 145 to 315                    |
| Propranolol    |                      |    |                  |                               |
| 4              | 8.5                  | 9  | 166              | 34 to 576                     |
| 5              | 17.0                 | 7  | 113              | 35 to 291                     |
| 6*             | 34.0                 | 9  | 30               | 11 to 180                     |
| 10†            | 34.0 (6 hours delay) | 9  | 135              | 87 to 397                     |
| 12†            | 34.0 (dextro isomer) | 8  | 235              | 126 to 322                    |
| Alprenolol     |                      |    |                  |                               |
| 7              | 8.5                  | 6  | 155              | 45 to 297                     |
| 8*             | 17.0                 | 8  | 69               | 11 to 187                     |
| 9*             | 34.0                 | 9  | 44               | 8 to 74                       |
| 11†            | 34.0 (6 hours delay) | 8  | 246              | 166 to 414                    |
| 13†            | 34.0 (dextro isomer) | 6  | 182              | 134 to 302                    |

\*Mann-Whitney U tests (two-tailed) performed between these groups and the control groups (pooled data from groups 1 to 3) revealed significant differences ( $P < .01$ ). †While each of these groups did not differ significantly from the pooled control data (groups 1 to 3), groups 10 and 12 each differed significantly from group 6. Likewise, groups 11 and 13 differed significantly from group 9.

versed by microinjections of *l*-norepinephrine.

Untrained male Sprague-Dawley rats ( $N = 160$ ) were housed singly and assigned to one of 13 groups (7). Group 1 received no surgery. Groups 2 to 12 were surgically prepared with bilateral cannulas positioned at the dorsal surface of the amygdala complex. At the completion of the experiment cannula placement was verified histologically according to previously established criteria (8).

One week after surgery all groups were trained in a passive avoidance task (9). On day 1, each animal was placed into the lit compartment of a two-compartment apparatus. As it stepped into a dark compartment through an open door which then closed, the animal received a footshock (1 ma, 2 seconds). Retention of passive avoidance was measured 24 hours later (day 2). An increase in latency (time between start of trial and entry into dark compartment) indicated retention of the training experience (10). Group 1 (no surgery) and group 2 (surgery only) served as control groups. Group 3 (vehicle control group) received a 0.5- $\mu$ l injection of a Krebs-Ringer phosphate solution (11) bilaterally in the amygdala complex immediately after training. Groups 4 to 6 received bilateral injections of the  $\beta$ -adrenergic blocking agent *dl*-propranolol (8.5, 17, or 34 nmole, respectively) immediately after training, and groups 7 to 9 received identical bilateral injections of a second  $\beta$ -adrenergic blocking agent, *dl*-alprenolol (8.5, 17, or 34 nmole). Groups 10 and 11 received 34-nmole injections of either *dl*-propranolol or *dl*-alprenolol 6 hours after training, and groups 12 and 13 received 34-nmole injections of the dextro isomer of either propranolol or alprenolol immediately after training (12). The volume and rate of infusion for all injections was 0.5  $\mu$ l per minute.

Retention test data are shown in Table 1. A Kruskal-Wallis one-way analysis of variance (13) performed on the latency data obtained on day 2 for all groups revealed a significant difference among groups (14). Since an additional Kruskal-Wallis one-way analysis of variance performed on the latency data obtained on day 2 for the control groups (groups 1, 2, and 3) revealed no significant difference among these groups, the data from these groups were pooled for further statistical analyses. Subsequent two-tailed Mann-Whitney U tests (13) indicated that, in comparison with the control groups, injections of 34 nmole of *dl*-propranolol or of either 17 or 34 nmole

Table 2. Latencies on day 2. The groups were as follows: 1, no surgery; 2, vehicle only; 3, combined *l*-norepinephrine (25 nmole) and *dl*-propranolol (34 nmole); 4, *l*-norepinephrine (25 nmole).

| Group | N  | Median (seconds) | Interquartile range (seconds) |
|-------|----|------------------|-------------------------------|
| 1     | 9  | 355              | 158 to 563                    |
| 2     | 9  | 334              | 128 to 491                    |
| 3*    | 10 | 175              | 81 to 496                     |
| 4*    | 10 | 201              | 144 to 264                    |

\*Did not differ significantly from the pooled data from groups 1 and 2.

of *dl*-alprenolol immediately after training produced a significant retention deficit ( $P < .01$ ), while lower doses of these two blocking agents did not significantly affect retention. The similar magnitude of effect on retention for equivalent doses of *dl*-propranolol and *dl*-alprenolol parallels results obtained with these two agents in other adrenergic systems, indicating that they are approximately equipotent  $\beta$ -adrenergic blocking agents (15).

The dose-related decrease in retention obtained at the higher doses of *dl*-propranolol and *dl*-alprenolol appeared to be time-dependent because if the injection of the 34-nmole dose was delayed for 6 hours after training retention was not impaired relative to controls. In addition, the retention deficit obtained with *dl*-propranolol and *dl*-alprenolol administered immediately after training was stereospecific because 34 nmole of the dextro isomer of either propranolol or alprenolol had no significant effect on retention 24 hours later.

The possibility that nonspecific effects caused by local anesthetic properties of the drugs accounted for the results is unlikely, because the dextro isomer had no effect on retention (16). Furthermore, electroencephalogram recordings taken from the sites of injection after the administration of either *dl*-propranolol or *dl*-alprenolol (34 nmole) in additional groups of animals did not reveal any abnormal afterdischarge activity. This absence of abnormal afterdischarge activity excludes the possibility that these drugs produce retention deficits by inducing the spread of abnormal electrical activity from the site of injection to extra-amygdala areas.

Our next experiment provided a further test of the hypothesis that the effects obtained with *dl*-propranolol and *dl*-alprenolol in our first experiment were due to the specific  $\beta$ -adrenergic blocking activity of these two agents. Since the  $\beta$ -

blocking activity of propranolol and alprenolol is competitive and reversible it would be predicted that increased adrenergic activity immediately after training might at least partially reverse the effects on retention produced by the administration of the  $\beta$ -adrenergic antagonists after training. We therefore assessed the effects on retention of injections of *dl*-propranolol in combination with *l*-norepinephrine.

Untrained Sprague-Dawley rats ( $N = 50$ ) served as subjects. Surgical, behavioral, histological, and data analysis procedures were identical to those in the previous experiment. Prior to training, the animals were assigned to one of four groups: control groups 1 (no surgery) and 2 (vehicle only); group 3, which received bilateral amygdala injections of *l*-norepinephrine (25 nmole) combined with *dl*-propranolol (34 nmole); and group 4, which received bilateral amygdala injections of *l*-norepinephrine (25 nmole). All injections were given immediately after training. A Kruskal-Wallis analysis of variance performed on the data obtained on day 2 (Table 2) revealed no significant differences among groups (14). Comparison of group 3 (*l*-norepinephrine and *dl*-propranolol) with group 6 of the previous experiment (*dl*-propranolol alone, 34 nmole; Table 1) revealed a significant difference between the two treatments ( $P < .05$ ). This reversal of the *dl*-propranolol retention deficit by injection of the adrenergic agonist *l*-norepinephrine in combination with *dl*-propranolol provides further support for the argument that the stereospecific amnesic effect observed in the first experiment was produced by the activity of these  $\beta$ -adrenergic blocking agents on an adrenergic-sensitive system in the amygdala (17).

Our results support the interpretation that  $\beta$ -adrenergic blockade in the amygdala of rats disrupts long-term memory formation in passive avoidance conditioning. While the possibility that drugs injected via the intracerebral cannula method produce their effects by spreading to extra-amygdala regions cannot be ruled out, autoradiographic studies (18) of intracerebral amygdala injections of [ $^{14}$ C]propranolol of twice the volume used in these experiments indicated that the drug was largely contained within the amygdala complex. Furthermore, others have reported that intraventricular injections of *dl*-propranolol at doses equal to and above those used in the present experiments have no effects on retention of passive avoidance conditioning (19),

suggesting that extra-amygdala spread into the ventricles is not responsible for the effects described herein.

Recent reports from other laboratories confirming the presence of norepinephrine in amygdala nuclei of rats (20) and studies on the localization and characterization of  $\beta$ -adrenergic receptors in the amygdala complex (21) strongly support the possibility that our results indeed reflect the effects of local manipulation of a specific  $\beta$ -adrenergic-sensitive neurochemical system within the amygdala of rats.

Previous observations that low-level electrical stimulation of the amygdala after training disrupts long-term memory formation have implicated this neuroanatomical region in the memory process (5).

Furthermore, pharmacological studies have indicated that the integrity of whole brain norepinephrine systems is necessary for long-term memory formation (6). Our results thus confirm and extend these previous findings by implicating an amygdala adrenergic system in long-term memory formation.

MICHELA GALLAGHER  
BRUCE S. KAPP, RICHARD E. MUSTY  
PATRICIA A. DRISCOLL  
Department of Psychology, University  
of Vermont, Burlington 05401

References and Notes

1. W. R. Russell and P. W. Nathan, *Brain* **69**, 280 (1946).  
2. W. Penfield and B. Milner, *Arch. Neurol. Psychiatry* **79**, 475 (1958); W. B. Scoville and B. Milner, *J. Neurol. Neurosurg. Psychiatry* **20**, 11 (1957).  
3. S. H. Barondes and D. H. Cohen, *Science* **160**, 556 (1968); S. L. Chorover and P. H. Schiller, *J. Comp. Physiol. Psychol.* **59**, 73 (1965); J. L. McGaugh, *Science* **153**, 1351 (1966); C. A. Pearlman, Jr., S. K. Sharpless, M. E. Jarvik, *J. Comp. Physiol. Psychol.* **54**, 109 (1961); L. J. Herz, *J. Neurobiol.* **1**, 111 (1969).  
4. H. E. Adams, P. R. Hoblit, P. R. Sutker, *Physiol. Behav.* **4**, 113 (1968); S. L. Chorover and A. M. DeLuca, *J. Comp. Physiol. Psychol.* **69**, 141 (1969); C. Cotman, G. Banker, S. Zornetzer, J. L. McGaugh, *Science* **173**, 454 (1971); L. B. Flexner and R. H. Goodman, *Proc. Natl. Acad. Sci. U.S.A.* **72**, 4660 (1975).  
5. P. E. Gold, J. Macri, J. L. McGaugh, *Behav. Biol.* **9**, 671 (1973); M. J. Handwerker, P. E. Gold, J. L. McGaugh, *Brain Res.* **75**, 324 (1974).  
6. C. T. Randt, D. Quartermain, M. Goldstein, B. Anagnoste, *Science* **172**, 498 (1970); R. K. Dismukes and A. V. Rake, *Psychopharmacologia* **23**, 17 (1972); L. Stein, J. D. Belluzzi, C. D. Wise, *Brain Res.* **84**, 329 (1975).  
7. Rats were obtained from the Canadian Breeding Laboratories, Quebec.  
8. Stereotaxic coordinates according to the Pellegrino and Cushman [L. J. Pellegrino and A. J. Cushman, *A Stereotaxic Atlas of the Rat Brain* (Appleton-Century-Crofts, New York, 1967)] rat brain atlas were -0.6 posterior to bregma, 4.7 lateral, and 7.0 ventral. Cannula placements for all animals that were operated on were established under the microscope with the aid of the Pellegrino and Cushman rat brain atlas. Cannula tip placements were rated as unacceptable if they were (i) more than 0.5 mm dorsal or ventral to the dorsal surface of the amygdala complex; (ii) more than 0.8 mm anterior or 0.6 mm posterior to the coronal plane -0.6 mm posterior to bregma; or (iii) lateral to the corpus callosum or

with any medial damage to the optic tract. Only animals with bilaterally acceptable cannula placements were included in the data analysis.  
9. B. S. Kapp, J. D. Kaufman, D. A. Repole, *Physiol. Behav.* **13**, 47 (1974).  
10. If an animal did not enter the dark compartment on day 2 within 600 seconds, the test was terminated and the animal was given a score of 600.  
11. W. W. Umbriet, R. H. Burris, J. F. Stauffer, *Manometric Techniques* (Burgess, Minneapolis, 1957), p. 149. The concentration of calcium in the Krebs-Ringer phosphate solution taken from Umbreit *et al.* was adjusted to 0.055M [J. K. Merlis, *Am. J. Physiol.* **131**, 67 (1940)].  
12. Drugs were generously supplied as hydrochloride salts from the following sources: *dl*-propranolol and *d*-propranolol (Ayerst Laboratories), *dl*-alprenolol (Astra Pharmaceutical), *d*-alprenolol (Hassle). *l*-Norepinephrine was obtained from Sigma.  
13. S. Siegel, *Nonparametric Statistics for the Behavioral Sciences* (McGraw-Hill, New York, 1956).  
14. A Kruskal-Wallis one-way analysis of variance performed on the latency data obtained on day 1 revealed no significant differences among groups.  
15. J. D. Fitzgerald, *Clin. Pharmacol. Ther.* **10**, 292 (1969); R. J. Lefkowitz, *Biochem. Pharmacol.* **24**, 1651 (1975); M. Zatz, J. W. Kebabian, J. A. Romero, R. J. Lefkowitz, J. Axelrod, *J. Pharmacol. Exp. Ther.* **196**, 714 (1976).  
16. A. L. Bassett and B. F. Hoffman, *Annu. Rev. Pharmacol.* **11**, 143 (1971).  
17. A group subjected to noncontingent footshock

(NCFS) and a group that received no surgery and no footshock (NFS) were removed from the apparatus after they entered the dark compartment on day 1 without receiving footshock. The NCFS group promptly received a noncontingent footshock (1 ma, 2 seconds) in a dissimilar apparatus, and then a combined injection of *l*-norepinephrine and *dl*-propranolol. Latencies on day 2 for these NCFS and NFS groups were uniformly low and did not significantly differ either from each other or from their own respective latencies on day 1. These data indicate that the increased latencies in group 3 (the combined *l*-norepinephrine and *dl*-propranolol group) were not a function of any aversive properties accompanying the combined drug treatment.  
18. J. S. Richardson, thesis, University of Vermont (1971).  
19. J. D. Belluzzi and C. D. Wise, Annual Meeting of the Eastern Psychological Association, New York, 1975.  
20. Y. Ben-Ari, R. E. Zigmond, K. E. Moore, *Brain Res.* **87**, 96 (1975); M. Brownstein, J. M. Saevedra, M. Palkovits, *ibid.* **79**, 431 (1974).  
21. R. W. Alexander, J. N. Davis, R. J. Lefkowitz, *Nature (London)* **258**, 437 (1975); D. B. Bylund and S. H. Snyder, *Mol. Pharmacol.* **12**, 568 (1976).  
22. This research was supported by the Heublein Training Fund in Neuropsychology and by a research scientist development award KO2 MH00118 (NIMH) to B.S.K.  
21 December 1976; revised 26 April 1977

Increasing Frequency of Thyroid Goiters in Coho Salmon (*Oncorhynchus kisutch*) in the Great Lakes

Abstract. Coho salmon collected during the 1976 spawning runs from Lakes Michigan, Ontario, and Erie had overt goiter frequencies of 6.3, 47.6, and 79.5 percent, respectively. These represent significant increases over the frequencies observed in previous years. Epizootiological data suggest that environmental goitrogens (possibly pollutants) may be involved in the etiology of the thyroid disorder.

Coho salmon (*Oncorhynchus kisutch*) is a highly prized sport and commercial marine fish. In the late 1960's the species was successfully introduced into the Great Lakes (1), and it has provided a spectacular multimillion-dollar sports fishery to North American anglers. Since it was introduced into the Great Lakes, several investigators have reported thyroid hyperplasia (goiters) in this species (2-4). Recently, Drongowski *et al.* (3) and Sonstegard and Leatherland (4) described severe hypothyroidism associat-

ed with thyroid hyperplasia in coho from Lakes Michigan and Ontario. While the hypothyroid condition (goiters) found in the coho salmon is undoubtedly due in part to the low availability of iodine in the Great Lakes Basin (5, 6), we report studies here which strongly suggest the involvement of environmental goitrogens (possibly pollutants) in the etiology of the thyroid disorder. In addition, our data (Table 1) suggest an increase in the frequency of occurrence of overt goiters over that in previous years.

Sexually mature coho salmon were collected during the fall spawning runs from Lakes Michigan, Ontario, and Erie. The presence of goiters was determined by retracting the operculum of each fish and examining the base of the gill arches for evidence of swelling or nodules (see Fig. 1). Fish with one or more distinct nodules more than 1 cm in diameter were recorded as having overt goiters. The frequency of overt goiters ranged from 6.3 percent in Lake Michigan to 47.6 percent in Lake Ontario to a striking 79.5 percent in Lake Erie.

If low iodine availability were the sole factor contributing to goiter develop-

Table 1. Goiter frequencies of Great Lakes coho salmon. Frequencies were determined from sexually mature fish examined during the fall spawning run. The number of fish examined is given in parentheses. Abbreviation: T.R., this report.

| Lake     | Year | Frequency (%) | Reference |
|----------|------|---------------|-----------|
| Erie     | 1972 | 44 (117)      | (2)       |
|          | 1976 | 79.5 (117)    | T.R.      |
| Ontario  | 1975 | 24 (51)       | (4)       |
|          | 1976 | 47.6 (63)     | T.R.      |
| Michigan | 1973 | 1 (100)       | T.R.      |
|          | 1976 | 6.3 (111)     | T.R.      |