- 3. C. W. Tabor, S. M. Rosenthal, H. Tabor, unpublished observations. 4.
- unpublished observations. H. Kihara and E. E. Snell, Proc. Natl. Acad. Sci. U.S. 43, 867 (1957); E. J. Herbst and E. E. Snell, J. Biol. Chem. 181, 47 (1949). (Earlier references cited.) C. W. Tabor, H. Tabor, S. M. Rosenthal, J. Biol. Chem. 208, 645 (1954); H. Tabor, Phar-macol. Revs. 6, 299 (1954). (Earlier refer-ences cited.) 5. ences cited.)
- We wish to thank Dr. C. Levinthal for call-6. ing this problem to our attention
- 8.
- Ing rules problem to our attention.
 A. D. Hershey, Virology 4, 237 (1957).
 H. Tabor, S. M. Rosenthal, C. W. Tabor, Federation Proc. 15, 367 (1956).
 R. C. Greene, J. Am. Chem. Soc. 79, 3929 (1957) 9.
- 1957) 10.
- (1997).
 (H. Tabor, S. M. Rosenthal, C. W. Tabor, *ibid.* 79, 2978 (1957).
 H. J. Vogel and D. M. Bonner, J. Biol. Chem. 218, 97 (1956). Glucose was added to the medium to make a 0.5 percent solution. Prepared by successive 4000, 20,000, 5000, and 00000 a centrifurgetion. The optical dentity 11.
- 12. 20,000 g centrifugations. The optical density at 260 m μ of 10¹¹ phage per milliliter was bout 1.2/cm.
- 13. The phage were ashed with $Mg(NO_a)_2$ by the procedure of Assoc. of Off. Agr. Chemists (1955). Inorganic phosphate, which was negligible before ashing, was assayed by the pro-cedure of C. H. Fiske and Y. Subbarow [J.
- G. R. Wyatt, Biochem. J. 48, 584 (1951).
 Prepared by H. Tabor, S. M. Rosenthal, C. 14 15.
- W. Tabor (unpublished synthesis). We wish to thank Dr. P. Hartman for a prep-aration of PLT-22 and Dr. L. Baron for 16.
- phage 98. We wish to thank Drs. H. Tabor and C. W. Tabor for their advice and helpful criticism. 17.

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Gibberellin-Induced Systemic Fruit Set in a **Male-Sterile Tomato**

Although recent reports (1-3) suggest that gibberellin or gibberellin-like substances (hereafter referred to as gibberellin) occur naturally in plants, investigators differ in their opinions regarding the movement of these compounds. The probability that gibberellin, like auxin, produces physiological effects distal from the site of synthesis indicates the need for further investigations of its movement in plants (4, 5). Hitchcock and Zimmerman (6) and Ferri (7) have demonstrated the movement of auxin through the plant by application to the soil, to roots, or to cuttings.

In the present study (8) the ability of gibberellin, applied as flower sprays, to set fruit parthenocarpically (9, 10) or to increase the growth of "dormant" tomato fruit (11) was regarded as a biological assay of its systemic movement.

The technique was refined by using male-sterile tomato plants (Lycopersicon esculentum) of the variety Earlypak (12) which were identified and selected at anthesis from a segregating backcross generation. These plants were normal in every respect except that the pollen grains were aborted (as indicated by an acetocarmine test). A few parthenocarpic fruit may set naturally on this muTable 1. Effect of gibberellin applied both to the foliage and soil on induction of fruit set in male-sterile Earlypak tomato.

Gibberellin per plant	Place of application	Average per plant			
		Total No. of clusters with fruits	Total No. of fruits	No. of fruits on treated lateral	No. of fruits on untreated lateral
100 µg	Expanded leaves	6.4	14.8	10.8	4.8
100 µg	Stem apices	7.0	12.0	7.0	5.0
100 µg	Flower peduncles	2.0	5.0	5.0	0.0
100 mg	Soil	8.0	36.0		
0	Control (untreated)	0.3	0.7		

tant, but any appreciable increase in numbers of fruit under isolated greenhouse conditions could be attributed to applied gibberellin.

Immediately preceding anthesis of the first flower cluster, the main stems of the tomato plants were pruned in order to stimulate the growth of two lateral branches from the cotyledonary axils. These branches were nearly alike with regard to time of flowering, number of flowers per cluster, and number and length of internodes. Basipetal and acropetal movement from a treated lateral would be reflected in a stimulation of fruit set on an untreated lateral. The plants were grown during the spring and summer in a greenhouse held at approximately 65°F at night. Day temperatures were held between 65° and 85°F.

In preliminary experiments to confirm previous results (13), floral sprays containing 500 µg of gibberellin per milliliter resulted in characteristic parthenocarpic fruit development. Subsequently, the effect of gibberellin (14) on inducing systemic fruit set was evaluated (i) by applying, with a micropipette, 100 µg per plant to the first or second fully expanded leaf above the second open flower cluster, to stem apices, and to the peduncle of a single inflorescence (15)and (ii) by applying 100 ml of a solution containing 1000 μ g/ml (100 mg) to the soil (Table 1). An excess of gibberellin was applied in order to compensate for the rapid degradation in the soil reported by Brian et al. (16). One milliliter of polyoxyethylene sorbitan monolaurate (Tween-20) per 100 ml of solution was added as a wetting agent for both plant and soil treatments.

Increased parthenocarpic fruit set on both treated and untreated laterals was induced by applying gibberellin to the foliage, but not by treating peduncles (Table 1). Greatest fruit set resulted from the soil application. Fruit from these treatments in every way resembled that resulting from direct floral sprays. All treatments, in addition to floral sprays, resulted in significant increases in size of "dormant" fruits. Johnson and Liverman (11) reported that a "dormancy" of developing fruits induced by high temperature or by far red irradiation could be overcome by spraying them with gibberellin. No quantitative studies were made to determine whether promotion of fruit growth in our experiments was comparable to that reported by Johnson and Liverman.

The marked increase in fruit set on an untreated lateral of a male-sterile plant indicates that gibberellin initiates a physiological response distant from the point of treatment. The results do not necessarily imply that gibberellin per se is directly responsible for systemic induction of fruit set. The use of male-sterile plants to assay for systemic fruit setting may have application for evaluating other growth-regulating substances.

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References and Notes

- B. O. Phinney et al., Proc. Natl. Acad. Sci. U.S. 43, 398 (1957).
 M. Radley, Nature 178, 1070 (1956).
- 3. 4.
- R. Lona, Ateneo Parmense 28, 111 (1957).
 R. Watanabe and N. J. Scully, Plant Physiol. Suppl. 32, 1vi (1957).
 P. M. Neely and B. O. Phinney, *ibid*. Suppl. 5.
- 32, xxi (1957). A. E. Hitchcock and P. W. Zimmerman, Con-6.
- tribs. Boyce Thompson Inst. 7, 447 (1935). M. G. Ferri, ibid. 14, 51 (1945).
- M. G. Ferri, *ibid.* 14, 51 (1945). This investigation was supported in part by a research grant from Merck, Sharp and Dohme, Inc. S. H. Wittwer *et al.*, *Plant Physiol.* 32, 39
- 9. (1957).
- 10. 11.

- (1957). L. Rappaport, Calif. Agr. 10, 4 (1956). J. L. Johnson and S. P. Liverman, Science 125, 1086 (1957). Appreciation is expressed to Dr. C. M. Rick for supplying seeds of $m_{s_{44}}$ [C. M. Rick, Rept. Tomato Genetics Coop. 6, 26 (1956)]. L. Rappaport, Plant Physiol. 32, 440 (1957). The preparation, supplied by Merck, Sharp and Dohme, Inc., contained the potassium salt of gibberellic acid (Gibrel) and about 80 percent apparently inactive materials.
- sait of globerenic acid (Glorel) and about ob percent apparently inactive materials. S. Zalik, G. A. Hobbs, A. C. Leopold, Proc. Am. Soc. Hort. Sci. 58, 201 (1951). P. W. Brian et al., J. Sci. Food Agr. 5, 602 (1954) 15. 16. (1954).

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SCIENCE, VOL. 127