toriness of chronic schizophrenia to treatment with Frenquel and reserpine (*5*) suggests that changes may take place which are not readily reversed.

The chromatographic work does point to a physiological alteration in schizophrenia which can be controlled by means of a suitable diet. It remains for future work to establish whether such an alteration is of basic importance to the disease.

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References and Notes

1. M. K. Young, Jr., *et al., Univ. of Texas Publ. No. 5109* (1951), pp. 189–197; I. Sano, *Folia Psychiat. et Neurol. Japonica* 8, 218 (1954).
2. P. L. McGeer, E. G. McGeer, W. C. Gibson, *Science* 123, 1029 (1956).
3. We wish to thank the laboratory and dietetic staff of the Provincial Mental Hospital and of Crease Clinic; without their cooperation this project would have been impossible.
4. L. I. Woolf *et al., Brit. Med. J.* 1955I, 57 (1955); L. E. Holt, Jr., *et al., Proc. Soc. Exptl. Biol. Med.* 48, 726 (1941).
5. F. Rinaldi *et al.*, paper presented before the American Psychiatric Association, Atlantic City, May 1955.

20 December 1955

Induced Spawning in the Indian Catfish

In the course of an investigation of the relationship between structural changes in the pituitary and the development of ova in the common Indian catfish, *Heteropneustes* (*Saccobranchus*) *fossilis* (Bloch), we have attempted to induce spawning under laboratory conditions (*1*). This work is preliminary to a wider project designed to bring about the spawning of other major Indian carps (*Labeo, Cirrhina, Catla*) in confining waters. Thus restricted, either these species may become gravid but fail to ovulate, or they may discharge unfertilized eggs (*2*). So far as we are aware, no attempt has previously been made in India to induce spawning of food fishes by injecting fresh (or acetone dry) pituitaries of the same species. Injections of mammalian pituitary preparations result in ova that are not viable (*3*).

Heteropneustes breeds in large tanks just after the monsoon showers. Ghosh and Kar (*4*) report that the active phase of the ovary extends from late April to July, but we have collected gravid females in August and September (1955). These were taken from a local tank (Bellandur, area 893 acres) from which all of our supply comes. The occurrence of gravid fish in our collections after the normal breeding season is doubtless due to the delay in the break of the monsoon. Weights of the gravid fish ranged from 70 to 135 g. They were held in aquaria (60 by 30 by 33 cm) that were supplied with city water from overhead cisterns, and the water was renewed daily. No additional aeration was provided, and all experiments were conducted at room temperature (24° to 25°C). The animals were not fed during the experimental period.

After a series of replications, it was found that a single injection of one-quarter of a fresh pituitary from a gravid *Heteropneustes* homogenized in 0.5 ml of standard Holtfreter solution was quite effective.

The injections were administered intramuscularly, since it was difficult to give intraperitoneal injections without damaging the egg-laden ovisacs. Controls injected with an equal quantity of Holtfreter solution did not spawn. Only specimens weighing more than 80 g were utilized, since it was found that smaller animals did not always respond to the injections.

Because spawning usually happened during the night, we have not been able to compute the time lapse necessary for the gravid females to spawn after each injection. However, our experiments indicate that at least 12 to 14 hours must elapse before the injected females spawn in aquaria. Eggs stripped from such females the following morning were fertilized in sperm suspension in Holtfreter solution and were cultivated both in Holtfreter solution and city water, since spring water was not easily accessible to us. Development proceeded normally in both media, and hatching took place within 32 to 34 hours.

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References and Notes

1. We wish to express our thanks to B. R. Seshachar, Central College, for kind encouragement.
2. S. L. Hora, *Proc. Nat. Inst. Sci. India* 11, 303 (1945).
3. A. Husain, *ibid.* 11, 320 (1945).
4. A. Ghosh and A. B. Kar, *Proc. zool. Soc. Beng.* 5, 29 (1952).
* Research scholarship, Ministry of Education, Government of India.

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Postirradiation Protection of Rabbits by Injection of "Splenic" Plasma

Jacobson (*1*) has attributed the "protection" of animals by shielding of the spleen during irradiation to a "humoral factor" elaborated by the spleen. It has also been shown that injection of certain tissue homogenates (of spleen or bone marrow, for example) soon after irradiation is similarly protective. However, it seems that the presence of cells or cell fragments in the injected material is necessary to bring about protection. If the postulated humoral factor is elaborated by the spleen, it should be present in higher concentration in the blood leaving the spleen. Therefore, plasma of blood taken from the splenic vein should exhibit a protective action.

Table 1. Comparative data in series I and II.

No. of animals	Plasma injection* (cm^3)	30-Day survival (%)	Avg. survival (day)
First series			
27		22	14.4
13	6 A	54	22.2
10	6	20	16.0
15	3†	20	14.4
Second series			
25		4	8.9
25	6 A	24	14.4
25	6 B	18	12.2

* Intraperitoneal, except where otherwise indicated.
† Intravenous. Eight rabbits received A plasma and one B plasma. The other six had been injected before it was thought necessary to distinguish between A and B plasma.

Such an experiment has been carried out using adult male chinchilla rabbits (*2*). In essence, heparinized blood was obtained from a nonirradiated donor rabbit as follows. The distal gastric coronary vein and artery were ligated at the stomach wall and then severed beyond the ligature on the spleen side. Blood from the splenic vein and artery was allowed to flow into a test tube in ice water through a polyethylene tube. This "splenic blood" is really a mixture of blood from the splenic vein, the splenic artery, and the proximal gastric coronary veins. The first 35 cm^3 of blood thus collected constitutes the A sample and the second 35 cm^3 the B sample. Each sample was centrifuged for 20 minutes at 2300 rev/min, and the plasma was separated.

After centrifugation of the splenic blood, approximately 18.0 cm^3 of plasma was carefully removed by pipette for injections. After the three 6.0-cm^3 doses of plasma had been injected, a residual drop of the plasma was placed on a slide, covered with a cover slip, and examined microscopically. No cells were found in the specimens examined. In one case, a more significant search for cells in splenic plasma was made as follows. Measured amounts of the plasma were drawn into erythrocyte- and leukocyte-diluting pipettes, and then the plasma