

lating K⁴²-labeled cells was also determined. The K⁴² and P³² (14.3-day half-life) were obtained from Oak Ridge. Each isotope was prepared for human injection in neutral sterile isotonic saline solution.

Ten ml of heparinized blood was incubated with about 50 µc carrier-free P³², and a similar volume was incubated with 100–200 µc K⁴² contained in about 50 mg of stable potassium carrier. With this carrier level, the presence of serum potassium had no effect on the uptake of potassium by the cells. The method of incubation and preparation of the blood for injection was a modification of the method described by Reeve and Veall (5).

The percentage of K⁴² taken up by the red blood cells ranged from 2% to 6% under varying conditions in which approximately the same amount of carrier potassium was used. The factors determining the rate of uptake are at present under study.

A weighed volume of the saline-suspended K⁴²-labeled cells was injected, and heparinized blood samples were withdrawn about 5 and 15 min later. This procedure was then repeated with the P³²-labeled cells. The radioactivity of the K⁴² in the injected and withdrawn samples of blood was determined immediately. The radioactivity of the P³² in the injected and withdrawn samples was determined 5 days later, at which time the K⁴² in the withdrawn blood was no longer detectable. The blood volume in each case was calculated in the usual manner. For the determinations of the biological half-life of the K⁴²-labeled cells, multiple specimens were taken for a 12- to 15-hr period.

The results of the almost simultaneous blood volume studies are given in Table 1. These two determinations

TABLE 1

Blood volumes

Patient	K ⁴² (ml)	P ³² (ml)	Av (ml)	Difference from av (%)
F. S.	5,940	6,400	6,170	3.7
J. R.	7,655	7,800	7,727	0.9
J. P.	4,945	4,820	4,882	1.3
O. H.	4,170	4,080	4,125	1.1
R. F.	4,750	4,870	4,810	1.3
J. B.	5,950	5,300	5,625	5.7

give essentially the same values within the errors inherent in the methods.

The biological half-life of the K⁴²-labeled cells *in vivo* ranged from 28 to 35 hr. This is consistent with studies on the *in vitro* rate of potassium uptake in red blood cells (6) and with studies that we will report elsewhere on the *in vivo* uptake of K⁴² by the RBC after plasma specific activity has reached a constant.

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The Metabolism of *Blastomyces dermatitidis*, Antagonists to the Growth-inhibiting Effect of Trimeton Maleate¹

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Previous reports from this laboratory (1, 2) presented evidence that trimeton maleate² (1-phenyl-1- α -pyridyl-3-dimethylaminopropane maleate, Schering) could completely inhibit the growth of *Blastomyces dermatitidis*³ and to a lesser extent the growth of other fungi, which results are similar to those reported by Carson and Campbell (3) using different antihistaminics. In our series (2) partial inhibition of *B. dermatitidis* was evident with 0.0003 M trimeton maleate, with complete suppression of growth of the fungus at 0.015 M concentration.

For investigation of the therapeutic implications of these findings, mice were infected with *B. dermatitidis* and treated with trimeton maleate. Twenty-eight albino mice of the Swiss strain (18–20 g) were infected by the intraperitoneal route with a heavy suspension of the yeast phase of *B. dermatitidis* suspended in 4% maltose, 1% peptone water. The organisms used for the mouse inoculations had been grown on blood agar in the incubator at 37° C for 2 weeks. The animals were divided into 2 groups for therapy, with paired controls receiving no treatment. Therapy was administered by subcutaneous injection of trimeton maleate, with a daily dosage of 40 mg/kg, divided into 2 injections/day. One group of the treated mice received therapy from the day of infection; in the other group therapy was initiated on the tenth day after the date of infection. For the prophylactic trial, 20 mice were divided into 2 groups. One group received 40 mg/kg trimeton maleate daily for 5 days preceding infection with *B. dermatitidis*; the other group received no pre-treatment, and treatment was instituted at the time of experimental infection.

All animals were sacrificed on the twenty-first day after infection, since in the experience of Spring (4), infection with *B. dermatitidis* is at a maximum in mice at this time. At autopsy, infection could be readily demonstrated by the presence of widespread nodules and partly necrotic masses affecting the mesentery, the retrosplenic and retrohepatic regions. Infection was determined by positive wet mounts, positive cultures, and by demonstrating the microorganism in histological sections.

These experiments indicated that trimeton maleate,

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³ We are indebted to Harry Seneca for his kindness in supplying the strain of *Blastomyces dermatitidis* used in this study.

TABLE 1
COMPARISON OF EXTENT OF GROWTH OF *Blastomyces dermatitidis* ON TRIMETON MALEATE TREATED MEDIUM* IN THE PRESENCE OF ADDED NEOPEPTONE AND ASHED NEOPEPTONE

Addition of neopeptone			Addition of ashed neopeptone		
Amt neo-peptone added (mg)	Colony diam (mm) Tube 1	Colony diam (mm) Tube 2	Amt ashed neo-peptone added† (mg)	Colony diam (mm) Tube 1	Colony diam (mm) Tube 2
200	6	7	7.8	6	7
400	17	12	15.6	11	10
600	17	19	23.4	19	20

Control Cultures Run Simultaneously‡	
Additions	Observations
No addition	Good growth covering entire slant
0.03 M trimeton maleate (final concentration)	No growth
400 mg neopeptone	Enhanced growth covering entire slant
15.6 mg ashed neopeptone	Enhanced growth covering entire slant

* Medium: 4% glucose, 1% neopeptone agar, with a final concentration of 0.03 M trimeton maleate. Standard amount of medium used was 20 ml/tube.

† 7.8 mg ashed neopeptone is equivalent to 200 mg neopeptone.

‡ Medium: 4% glucose, 1% neopeptone agar used; amount, 20 ml.

under the conditions of the experiment, failed to prevent the infection, and furthermore the infected animals did not respond favorably to the therapy with trimeton maleate.

In order to elucidate the mechanism of the *in vitro* inhibitory effect of trimeton maleate on the growth of *B. dermatitidis*, a series of experiments was conducted to determine a means of reversing this inhibition and to search for an explanation of the mechanism. For that purpose the following substances were incorporated in an 0.03 M trimeton maleate, 4% glucose, 1% neopeptone agar: (1) neopeptone (Difco); (2) ashed mineral residue of neopeptone (Difco); (3) histamine dihydrochloride; (4) 1-histidine monohydrochloride (Tables 1 and 2). Before incorporation of the added substances, the pH in each instance was adjusted to the pH level of the medium. Experimental tubes were inoculated at one point from a fresh culture of *B. dermatitidis* and incubated at room temperature for one month, when the extent of growth was determined by inspection.

Reversal of the inhibitory effect by additions of neopeptone (200–600 mg neopeptone added per 20 ml culture medium) induced us to investigate whether the mineral content or the organic part of the neopeptone was involved in this mechanism. After adding ashed neopeptone (ashed residue of 200–600 mg neopeptone per 20 ml culture medium) to the trimeton-containing glucose medium, growth of *B. der-*

matitidis was obtained that was equal in each case to the growth with the corresponding amount of neopeptone. Thus the ashed residue evidently caused a marked reversal of the inhibitory effect of trimeton maleate. Growth with the largest amount of ashed residue added (equivalent of 600 mg neopeptone) was not quite equal to the growth in the untreated control, but the extent of growth in the experimental tubes varied directly with the concentration of added ashed residue (Table 1).

Subsequent trials with the following cations were and are being performed: Mg, Na, K, Ca, Li, Ba, Ni, Sr, and Fe. With the exception of nickel and lithium, the remaining metals (added in molar concentrations equal to the concentration of trimeton maleate in the medium as well as in twice that amount) showed appreciable reversal of the growth inhibition by trimeton maleate. The effect of the addition of several anions on the suppression of growth of *B. dermatitidis* by trimeton maleate is being studied.

Although actually we have no proof to offer that the cations tested are part of an enzyme system of *B. dermatitidis*, in the light of our experiments such a possibility can be considered. Similar investigations were conducted by Abelson and Aldous (5), who demonstrated that the toxicity of Ni, Co, Cd, Zn, and Mn for *Escherichia coli* could be lowered by adding large amounts of magnesium. In a similar way the toxicity of Ni and Co could be diminished in the presence of magnesium in the case of three other organisms tested: *Aerobacter aerogenes*, *Torulopsis utilis*, and *Aspergillus niger*. It is of additional interest to mention that Utter and Werkman (6) have demonstrated that Mg activated an important enzyme converting phosphoglycerate to phosphopyruvate. This enzyme acts partially without the addition of the divalent metal. The addition of small amounts of Ni

TABLE 2
ADDITIONS OF HISTAMINE AND HISTIDINE TO TRIMETON MALEATE TREATED MEDIUM, INOCULATED WITH *Blastomyces dermatitidis*

Tube No.	Molar concentration of trimeton maleate	Molar concentration of histamine	Molar concentration of histidine	Observations
1	0	0	0	Good growth*
2	0.03	0	0	No growth
3	0.03	0.03	0	" "
4	0.03	0.06	0	" "
5	0.03	0.09	0	" "
6	0	0.06	0	Enhanced growth
7	0	0.09	0	" "
8	0.03	0	0.03	No growth
9	0.03	0	0.06	" "
10	0.03	0	0.09	" "
11	0	0	0.06	Enhanced growth
12	0	0	0.09	" "

* Results reported represent 6 replicate tubes for each condition with uniform findings.

